EFFECT OF BIOLOGICAL SOIL CRUSTS ON THE SEED GERMINATION OF THREE PLANT SPECIES UNDER LABORATORY CONDITIONS

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Abstract: Semiarid grasslands in northeastern Mexico contain endemic plants and animals, and are an important refuge for resident and migratory animals. Here, as in other semiarid areas, biological soil crusts (BSC) are a key component of the ecosystem. However, findings about their effect on the germination of vascular plants are contradictory. We asked whether seed germination of some native plant species would be inhibited by the presence of BSC as it has been found in other studies; in turn, we evaluated the effect of five lichens (Endocarpon pusillum Hedw., Placidium sp., Psora cerebriformis W.A. Weber, Psora decipiens (Hedw.) Hoffm., Xanthoparmelia chlorochroa (Tuck.) Hale), one cyanobacteria (Nostoc commune Vauch.), and one hepatic (Oxymitra sp.), from the dominant BSC of the area on the germination percentage (germinability) and speed of germination (t50) of three native plant species: Frankenia gypsophila I.M. Johnst., Muhlenbergia arenicola Buckl., and Sartwellia mexicana A. Gray. Germination tests were carried out in an environmental chamber at 26 °C and at a constant humidity of 60% with 12:12 hours light and darkness. BSC did not affect germination percentage of the three evaluated plant species when compared to bare soil. Speed of germination ranged between 3.5 and 5 days and there were no differences between substrates.

Keywords: Endocarpon pusillum, northeastern Mexico, Nostoc commune, Placidium, semiarid grasslands.

Biological soil crusts (BSC) are communities of organisms that live in the soils of arid and semiarid areas, and are the main living cover in many arid areas of the world (Belnap, 2006). BSC communities consist of soil surface-dwelling cyanobacteria, green algae, microfungi, bacteria, lichens, and bryophytes (Hawkes, 2003; Zhang, 2005; López-Cortés et al., 2010) that grow on top or within the top layers of soil (Belnap et al., 2001; Quiñones-Vera et al., 2009). They influence local hydrological cycles, including soil porosity, roughness, aggregate stability, texture, pore formation, and water retention (Belnap, 2006), contribute to soil retention (Belnap and Gardner, 1993; Zhang et al., 2006), and facilitate the accumulation of soil nutrients (Jafari et al., 2004). Due to the effects of microtopography, and water and nutrients fluxes, BSC affect the distribution of resources in the soil, contributing to vegetation cluster patterns
(Deines et al., 2007) and probably improve water infiltration and retention in the soil (Casenave and Valentin, 1989; Montaña et al., 1995; Rosentreater and Belnap, 2001; Reyes-Gómez et al., 2007; Yair, 2008; Chamizo et al., 2010).

BSC can also affect the establishment and survival of vascular plants, and available data show varied results for BSC. Studies have shown BSC having either facilitative, inhibitory or no effect on vascular plant establishment and cover (During and Van Tooren, 1990; Belnap et al., 2001). Existing evidence suggests that the effects of crusts on seed germination vary depending on the soil crust composition and plant characteristics (Lesica and Shelly, 1992; Zaady et al., 1997; Prasse and Bornkamm, 2000; Rivera-Aguilar et al., 2005; Serpe et al., 2006; Deines et al., 2007), as seeds vary in their germination requirements and consequently may respond differently to the conditions created by the crust (Baskin and Baskin, 1998). Positive effects of the crust on vascular plants may be the result of improved soil moisture conditions and nutrient availability, as well as a decrease in predation (During and Van Tooren, 1990; Belnap and Harper, 1995). Negative effects could be attributed to competition for water and nitrate, changes in the red/far-red ratio or the presence of inhibitory compounds (During and Van Tooren, 1990; Callaway and Walker, 1997).

Speed of germination results for different BSC and plant species have also been controversial. For example, Godínez-Alvarez et al. (2012) found no differences in $t_{50}$ for Agave marmorata Roezl. and Neobuxbaumia tetetzo (F.A.C. Weber) Backeb. germinated on a cyanobacteria crust and on bare soil. Deines et al. (2007) compared germination of two grass species (Bromus tectorum L. and Vulpia microstachys (Nutt.) Benth.) on three different substrates (lichen crust, mixed crust, and bare soil) and found a longer germination time on the lichen crust than on bare soil and no differences between the mixed crust and bare soil for both grass species.

Recent studies have demonstrated that BSC dominated by Diplolischistes sp. reduced plant emergence (Escudero et al., 2007; Serpe et al., 2008). This might be due to the sealing of the soil pores by these crusts that might reduce water availability and thus inhibit germination (Eldridge et al., 2010). Similarly, Prasse and Bornkamm (2000) in the Negev Desert in Israel and Li et al. (2005) in the Tengger Desert of China found greater seedling emergence after removal or destruction of the BSC. St. Clair et al. (1984) observed significantly higher germination for three grass species on algal crusts compared with mixed crusts of lichens, mosses and algae. Zamfir (2000) showed that seedling emergence was much lower on lichen mats compared with moss mats for three of the four species studied.

These uneven results highlight the relevance of evaluating more specific associations of BSC and vascular plant species in order to assess seed germination, seedling emergence, and plant establishment. Hence, in this study we determined the effect of seven dominant BSC from the Northern Mexican Plateau (Molina-Guerra, 2013) on the germination percentage and speed of germination ($t_{50}$) of three native plants. While the physical barrier of crusts is likely to affect soil moisture and several stages of seedling growth, in here we asked whether germination would be inhibited by the presence of lichens as it has been found in other studies (Deines et al., 2007).

Material and methods

Study area. The BSC were collected in the states of Nuevo León, Coahuila, and San Luis Potosí within the Southern part of the Chihuahuan Desert in northeastern Mexico. The collection area was within 25° 13’ 51” N and 101° 17’ 28” W. Mean annual temperature for the region is 17.2 °C, with a minimum of -1.8 °C in January and maximum of 35.1 °C in May. Average annual rainfall is 386.43 mm, with March being the driest month (8.43 mm) and August being the wettest (58.06 mm; SMN, 2000).

According to the Lof P. and van Baren (1987; FAO-UNESCO classification), soils in the area are mainly solonchack and calcic phaeozem, and smaller areas of chromic vertisol and luvic chernozem (INEGI, 1981a, b; 2002). These soils have been classified into three broad groups: (1) those with a high content of calcium carbonate and no gypsum; (2) soils with similar contents of gypsum and calcium carbonate; and (3) those with high gypsum content and very little or no calcium carbonate (Pando et al., 2013). Sampling areas are between 1,800 and 2,000 m altitude.

The vegetation is mainly made from communities of halophytic/gypsophyllus grasslands associated with microphyll and rosetophyll desert scrub (Estrada-Castillón et al., 2010), where the most abundant plant families are: Poaceae, Chenopodiaceae, and Frankeniaceae (Rzedowski, 1978). These grasslands contain several endemic plants (Dalea gypsofila, D. radicans, Frankenia gypsophila, Mahaeranthera heterophylla, and M. crutchfii eldii; Estrada-Castillón et al., 2010), and animals (Cynomys mexicanus, Spizella worthenii; Baillie and Groombridge, 1996).

Field sampling. The dominant BSC were collected with a cylinder designed by the authors. The size of the cylinder was adjusted to the size of the Petri dishes where the samples were placed (100 × 15 mm). All Petri dishes were covered with mixed crust where at least 50% was occupied by the treatment species. Samples to be collected were previously sprayed with distilled water to facilitate their extraction. The cylinder was placed on top of the soil and pressed until the lower part of the cylinder touched the soil crust. Once the crust was extracted it was placed on a Petri dish and sealed with tape. The components of the collected mixed crusts were identified by experts from two mycology laboratories, there were: five lichens Endocarpon pusillum, Verrucaria-
ceae; Placidium sp., Verrucariaceae; Psora cerebriformis, Psoraceae; Psora decipiens, Psoraceae; and Xanthoparmelia chlorochroa, Parmeliaceae (all squamulose except X. chlorochroa which is usually foliose); one cyanobacteria: Nostoc commune, Nostocaceae; and one hepatic: Oxymitra sp., Oxymitracaceae. Additionally, samples of bare soil from the three broad groups of soils in the region were collected to be used as controls.

Three native plant species that co-exist with the BSC to be used as controls. Seeds of the first two plant species have been found to have a high germination percentage (> 80%) under laboratory conditions, using agar as a medium (Contreras et al., 2012). The seeds were collected in September 2011 from plants growing within the study region; each seed lot contained a minimum of 50 individuals. The seeds were manually extracted, cleaned and selected using a stereoscopic microscope. They were then stored in plastic containers at room temperature.

There were no pre-treatments for either the seeds or the BSC before germination tests. Germination tests were carried out in an environmental chamber (Lumistell® ICP-S4) at 26 °C (following Contreras et al., 2012) and at a constant humidity of 60% with 12:12 h light and darkness. The seeds were checked every 24 h (Baskin and Baskin, 1998) over one month. Light intervals were defined to resemble average day and night number of hours for these latitudes. Temperature was established according to Contreras et al. (2012), who worked on germination of these plant species even though her work did not involve soil biological crusts.

Experiment set up and analysis. There were 30 treatments arranged randomly in the germination chamber (ten substrates (BSC or soil) × three plant species) with five replicates. The substrates were: S1: soil with gypsum and calcium carbonate (Sgc) (45% gypsum and 35% calcium carbonate); S2: Psora cerebriformis; S3: Placidium sp.; S4: Endocarpon pusillum; S5: Nostoc commune; S6: soil with calcium carbonate and no gypsum (Sc) (0% gypsum and 62% calcium carbonate); S7: soil with gypsum and no calcium carbonate (Sg) (50% gypsum and 5% calcium carbonate); S8: Xanthoparmelia chlorochroa; S9: Psora decipiens; and S10: Oxymitra sp. Seeds were from three plant species: Frankenia gypsophila, Muhlenbergia arenicola, and Sartwellia mexicana. The substrates S2-S5, as well as S8-S10 were labeled with the name of the species occupying at least 50% of the Petri dish.

Ten seeds of each plant species were sown on the surface of the dominant BSC in each Petri dish (50 seeds per treatment). The seeds were sown evenly spaced on the surface of the selected species of BSC, except for those on bare soil treatment. For samples not covering 100% of the surface, seeds were specifically placed on top of the particular soil crust component. Germination was recorded when the seed radicle emerged and was visible (Jurado and Westoby, 1992; Welbaum et al., 1998). From these observations we obtained:

1) germinability, calculated as the percentage of germinated seeds at the end of the experiment (Flores and Jurado, 1998; Flores and Briones, 2001); and 2) Speed of germination or half-time of germination (t50), calculated as the time (in days) at which 50% of the seeds germinated (Grime et al., 1981; Thompson and Grime, 1983; Jurado and Westoby, 1992; Flores and Jurado, 1998; Flores and Briones, 2001; Pérez-Sánchez et al., 2011).

Germination percentage. As a whole, there were no significant differences in germinability between substrates (F = 1.09, df = 9, P = 0.368). Germinability differed between substrates for Frankenia gypsophila (F = 3.712, df = 49, P = 0.002) and Muhlenbergia arenicola (F = 3.094, df = 49, P = 0.007), and there were no differences for Sartwellia mexicana (F = 1.650, df = 49, P = 1.34). Frankenia gypsophila germinability was similar in all but one substrate: Nostoc commune where germinability was lower than in the others (66%; Table 1). The lowest germinability of Muhlenbergia arenicola was on bare soil with gypsum (18%), there were no differences between the other substrates.

Frankenia gypsophila showed the highest germination percentage (91.4% ± 13 sd), followed by Muhlenbergia arenicola (56.6% ± 17 sd) and Sartwellia mexicana (13% ± 9 sd). This ranking was consistent when evaluating germination percentage by substrate, except for Nostoc commune where M. arenicola had a higher germinability percentage (78%) than F. gypsophila (66%).
Speed of germination or half-time of germination ($t_{50}$). The $t_{50}$ was analyzed only for those seed species whose seed germination percentage was above 15% in all substrates. This threshold was established because germination percentages below 15% could generate $t_{50}$ values that did not represent the species speed of germination, but rather reflected that those seeds might require other treatments to promote germination. Hence, $t_{50}$ is not shown for *Sartwellia mexicana* seeds.

*Frankenia gypsophila* and *Muhlenbergia arenicola* seed germination started on the second day after sowing and had an average $t_{50}$ of 4.86 and 3.65 days, respectively. According to Jurado and Westoby (1992), germination for *M. arenicola* was fast, and moderate for *F. gypsophila* across substrates. Speed of germination ($t_{50}$) was equal for all substrates ($F = 1.66$, df $= 9$, $P = 0.109$) and their values ranged between 3.5 and 5 days (Table 1).

### Discussion

**Germination percentage or germinability.** Results from this research indicated that, under laboratory conditions, the BSC did not affect the seed germination percentage of the three evaluated plant species from the Northern Mexican Plateau.

In agreement with this study’s findings, Li *et al.* (2005) found no differences in germination percentage and seedling establishment between soils with biological crusts and bare soil for *Eragrostis poaeoides* Beauv. and *Bassia dasyphylla* O.Kuntze under wet conditions in the Tengger Desert in Northern China. Deines *et al.* (2007) found negative effects for BSC on the germination of *Bromus tectorum* L. and *Vulpia microstachys* Nutt., where germination percentage, analyzed under laboratory conditions, was about three times lower on the BSC dominated by the lichen *Diploschistes muscorum* (Scop.) R.Sant., than on bare soil. Germination on bare soil was found to be equal to mixed biological crusts which contain the lichens *Aspicilia*, *Caloplaca*, *Candelariella*, *Collema*, and *Placidium*, the cyanobacterium *Microcoleus*, and a low growing form of the moss *Syntrichia caninervis* Mitt. Zaady *et al.* (1997) found that germination of three mucilaginous seeds decreased on soils with a smooth crust dominated by cyanobacteria.

The effects of the BSC on germination may vary according to plant species, the type of biological crust (either smooth or rough), and the species that dominate in the crust (Rivera-Aguilar *et al.*, 2005; Deines *et al.*, 2007), as well as the size and morphology of the seeds and the presence of microsites in the crust suitable for germination (Li *et al.*, 2005). Escudero *et al.* (2007) and Serpe *et al.* (2008) showed that BSC reduced seedling emergence when it was dominated by species like *Diploschistes* sp., because this lichen might seal the soil, thus reducing water availability during the germination period of some plants (Otsus and Zoel, 2004; Morgan, 2006). Contrastingly, the presence of both algal and moss crusts significantly enhanced the germination of *Bassia dasyphylla* (Fisch. & Mey.) O.Kuntze and *Artemisia ordosica* Krasch. compared with uncrusted soil (Su *et al.*, 2009).

Godínez-Alvarez *et al.* (2012) found that biological soil crusts increased germination of one species (*Agave marmorata*) but did not show any effect on the germination of two other species from the same region (*Prosopis laevigata* (H. & B.) Jonhst. and *Neobuxbaumia tetetzo*). Germination of

### Table 1. Germination percentage and speed of germination ($t_{50}$) (average and standard error, $n = 5$) per plant species by substrate.

<table>
<thead>
<tr>
<th>Substrate</th>
<th><em>Frankenia gypsophila</em> %</th>
<th>S.E.</th>
<th><em>Muhlenbergia arenicola</em> %</th>
<th>S.E.</th>
<th><em>Sartwellia mexicana</em> %</th>
<th>S.E.</th>
<th>$t_{50}$ %</th>
<th>S.E.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bare soil with gypsum and calcium carbonate</td>
<td>100</td>
<td>0.00</td>
<td>60</td>
<td>7.07</td>
<td>10</td>
<td>5.47</td>
<td>56.7</td>
<td>4.0</td>
</tr>
<tr>
<td><em>Psora cerebriformis</em> W.A. Weber</td>
<td>90</td>
<td>4.47</td>
<td>48</td>
<td>11.14</td>
<td>10</td>
<td>3.16</td>
<td>49.3</td>
<td>4.7</td>
</tr>
<tr>
<td><em>Placidium</em> sp.</td>
<td>98</td>
<td>2.00</td>
<td>52</td>
<td>11.58</td>
<td>0</td>
<td>0.00</td>
<td>50.0</td>
<td>3.6</td>
</tr>
<tr>
<td><em>Endocarpon pussillum</em> Hedwig</td>
<td>100</td>
<td>0.00</td>
<td>70</td>
<td>3.16</td>
<td>2</td>
<td>2.00</td>
<td>57.3</td>
<td>3.5</td>
</tr>
<tr>
<td><em>Nostoc commune</em> Vauch.</td>
<td>66</td>
<td>11.66</td>
<td>78</td>
<td>10.20</td>
<td>14</td>
<td>5.10</td>
<td>52.7</td>
<td>4.9</td>
</tr>
<tr>
<td>Bare soil with gypsum</td>
<td>96</td>
<td>4.00</td>
<td>18</td>
<td>5.83</td>
<td>30</td>
<td>16.43</td>
<td>48.0</td>
<td>4.7</td>
</tr>
<tr>
<td><em>Xanthoparmelia chlorochroa</em> (Tuck.) Hale</td>
<td>70</td>
<td>15.81</td>
<td>54</td>
<td>9.27</td>
<td>20</td>
<td>5.48</td>
<td>48.0</td>
<td>5.0</td>
</tr>
<tr>
<td><em>Oxymitra</em> sp.</td>
<td>96</td>
<td>2.45</td>
<td>70</td>
<td>8.94</td>
<td>14</td>
<td>5.10</td>
<td>60.0</td>
<td>4.1</td>
</tr>
<tr>
<td>Bare soil with calcium carbonate</td>
<td>100</td>
<td>0.00</td>
<td>54</td>
<td>14.35</td>
<td>12</td>
<td>5.83</td>
<td>55.3</td>
<td>3.6</td>
</tr>
<tr>
<td><em>Psora decipiens</em> (Hedwig) Hoffm.</td>
<td>98</td>
<td>2.00</td>
<td>62</td>
<td>7.35</td>
<td>18</td>
<td>4.90</td>
<td>59.3</td>
<td>4.1</td>
</tr>
</tbody>
</table>
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A. marmorata was significantly higher on a cyanobacteria crust (72.5%) than on a mixed crust (30%) that contained cyanobacteria and moss (mainly Bryum and Pseudocressidium species). Germination on bare soil (52.5%) had intermediate values that did not differ significantly from any of the BSC treatments. Godínez-Alvarez *et al.* experiment was also conducted in a laboratory and at constant humidity for all treatments. Zhang *et al.* (2010) found contrasting results using moist and dry conditions for five species in the China Gurbantunggut Desert, the presence of BSC resulted in less seeds germinating in two out of five species under dry conditions, and in three out of five species under moist conditions.

**Speed of germination (t₅₀).** Seeds of Muhlenbergia arenicola and Frankenia gypsophila showed average t₅₀ of 3.65 days (fast) and 4.86 days (moderate), respectively. These short germination times could reflect the response of the seeds to one single event of profuse rainfall, typical for an environment where wet soils are uncommon and humid periods are brief (Jurado and Westoby, 1992), which are typical climatic characteristics for the area where the seeds chosen for this study came from.

Speed of germination was similar across substrates. Likewise, Godínez-Alvarez *et al.* (2012) found that the number of days to 50% of the total germination (t₅₀) for Agave marmorata and Neoduxbaunmia tetetzo, from semiarid Valle de Tehuacan in Mexico, were between 1.88 and 5.5 days for the different species and treatments with no statistical differences between them. In contrast, in the same area of Valle de Tehuacan in Mexico, Rivera-Aguilar *et al.* (2005) evaluated seed germination rate using quadratic regressions for two native plant species (Mimosa luisana Brandegee and Myrtillocactus geometrizans (Martius) Console) concluded that seeds placed in BSC had a faster germination rate compared to those placed in bare soil. Contrarily, Deines *et al.* (2007) found that mean germination time (MGT) of *Vulpia microstachys* Nutt. and *Bromus tectorum* L. was faster on mixed crusts and on bare soil than on crusts of lichens, with an average of 5-7 days for the first two conditions, and 11 days for the crust of lichens. Also working with grasses (*Festuca idahoensis* Elmer, *Festuca ovina* L., *Elymus wavavaiensis* J. Carlson & Barkworth, and *Bromus tectorum* L.), Serpe *et al.* (2006) found that MGT was shorter for seeds germinating on bare soil than on BSC made of short or large mosses (*Tortula ruralis* Hedw. and *Bryum argenteum* Hedw.), and Briggs and Morgan (2011) found that two out (*Salvia verbenaca* and *Vittadinia gracilis*) of the five species tested, exhibited shorter t₅₀ on disturbed soil crust than on moss and foliose lichen. The other species tested were *Austrodanthonia* sp. (Poaceae), *Maireana excavata* (Chenopodiaceae), and *Leptorhynchos scaber* (Asteraceae), all common in the semi-arid woodlands to the northwest of Melbourne, Australia.

These somehow uneven results on speed of germination (t₅₀) seem to be a consequence of differences on the way the experiments were carried out (e.g. providing or not constant humidity), as well as the roughness of the evaluated BSC and the dominant species on the crust.

Further experimental and field evidence is required to better understand the implications of BSC on seed germination and seedling establishment.

**Conclusions**

The purpose of this paper was to assess whether the more conspicuous types of BSC found in the semiarid grasslands of northeastern Mexico inhibit or delay seed germination of vascular plants as has been reported for other biological soil crusts. By analyzing the effect of five lichens, one cyanobacteria, and one hepatic on the germination of native plants, we conclude that under high substrate humidity conditions, BSC did not inhibit or delay seed germination of the vascular plants used for this study.

*Muhlenbergia arenicola* and *Frankenia gypsophila* had fast and moderate average speed of germination or half-time of germination (t₅₀) of 3.65 days and 4.86 days, respectively, according to the expected germination times in environments where there are only brief periods of high soil humidity. Values for t₅₀ were between 3.5 and 5 days and did not differ between substrates.

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