

This is the Author's Pre-print version of the following article: *Cynthia L. Pérez-Ruiz, Ernesto I. Badano, Juan P. Rodas-Ortiz, Pablo Delgado-Sánchez, Joel Flores, David Douterlungne, Jorge A. Flores-Cano, Climate change in forest ecosystems: A field experiment addressing the effects of raising temperature and reduced rainfall on early life cycle stages of oaks, Acta Oecologica, Volume 92, 2018, Pages 35-43, Pages 22116-22125*, which has been published in final form at: [10.1016/j.actao.2018.08.006](https://doi.org/10.1016/j.actao.2018.08.006)

© 2018 This manuscript version is made available under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) license <http://creativecommons.org/licenses/by-nc-nd/4.0/>

1 **Climate change in forest ecosystems: a field experiment addressing the effects of raising**
2 **temperature and reduced rainfall on early life cycle stages of oaks**

3

4 **Cynthia L. Pérez-Ruiz^a, Ernesto I. Badano^{a,*}, Juan P. Rodas-Ortiz^a, Pablo Delgado-**
5 **Sánchez^b, Joel Flores^a, David Douterlungne^c, Jorge A. Flores-Cano^b**

6

7 ^a IPICYT-División de Ciencias Ambientales, Instituto Potosino de Investigación Científica y
8 Tecnológica, A.C., Camino a la Presa San José 2055, Lomas 4^a Sección, C.P. 78216, San Luis
9 Potosí, SLP, México.

10 ^b Facultad de Agronomía y Veterinaria, Universidad Autónoma de San Luis Potosí, Km. 14.5
11 Carretera San Luis-Matehuala, Apdo. Postal 32, Soledad de Graciano Sánchez, C.P. 78321, San
12 Luis Potosí, SLP, México.

13 ^c CONACYT-IPICYT, División de Ciencias Ambientales, Camino a la Presa San José 2055, CP
14 78216, San Luis Potosí, SLP México.

15

16 *Corresponding author. IPICYT/División de Ciencias Ambientales, Camino a la Presa San José
17 2055, Lomas 4^a Sección, C.P. 78216, San Luis Potosí, SLP, México. E-mail address:

18 ernesto.badano@ipicyt.edu.mx (E.I. Badano).

19 **Abstract**

20 Higher temperatures and reduced rainfalls that are expected with the advance of climate change
21 can impair the emergence and establishment of tree seedlings in forest ecosystems. These
22 climatic changes can also decrease the availability of soil resources and reduce the performance
23 of seedlings. We evaluated these effects in a temperate forest from Mexico with two native oak
24 species (*Quercus crassifolia* and *Quercus eduardii*). As recently emerged oak seedlings are
25 highly sensitive to changing environmental conditions, our field experiment was conducted
26 across the season in which seedling emergence occurs (October-February). In the field, we used
27 open-top chambers to increase temperature and rainout shelters to reduce rainfall, while controls
28 were exposed to the current climate. Experimental plots of both treatments were established
29 beneath the forest canopy because most oaks recruit in understory habitats. In these plots, we
30 sowed acorns of both species in October 2015 and recorded seedling emergence and survival
31 until February 2016, also monitoring temperature, precipitation and contents of water and
32 nitrogen in the soil. On seedlings that survived until the end of the experiment we measured their
33 growth, photosynthetic efficiency and foliar contents of water, carbon and nitrogen. Both the
34 emergence and survival of *Q. crassifolia* seedlings were lower in climate change plots than in
35 controls, but no differences were found for *Q. eduardii*. However, seedlings of both species had
36 lower growth rates, photosynthetic efficiencies and contents of water, nitrogen and carbon in
37 climate change simulation plots. These results indicate that climate change can impair tree
38 seedling establishment in oak forest, also suggesting that their development will be constrained
39 by reduced water and nitrogen availability.

40

41 **Keywords**

42 Drought; Growth rate; Leaf traits; Open-top chambers; Rainout shelters; Warming

43 **1. Introduction**

44 The prevalence of forest ecosystems mainly depends on the emergence and establishment of tree
45 seedlings, but climate change can alter these processes (Clark et al., 2016). In temperate forests
46 from North America, most global circulation models predict that the average air temperature will
47 rise 5° C in the course of this century, while annual precipitation levels in these ecosystems are
48 projected to decrease up to 20% (IPCC, 2013). These ecosystems will then face warmer and drier
49 conditions in the future, but field experiments assessing how tree seedlings will deal with these
50 changes are still scarce.

51 Water availability is a key factor that regulates the emergence of tree seedlings in forests and
52 climate change can strongly impair this process (Pérez-Ramos et al., 2013; Clark et al., 2016).
53 This is particularly critical for tree species that produce recalcitrant seeds because higher
54 temperatures and lower precipitation levels can enhance seed water loss and induce embryo
55 mortality, hence reducing seedling emergence (Zavala-Chávez, 2008; Joët et al., 2013). Further,
56 if seedlings emerge under these conditions, they can display a number of functional responses
57 that would ultimately reduce their performance in climate change scenarios. Increased
58 temperatures usually enhance foliar transpiration but, when plants are also subjected to water
59 deficits, stomatal closure is induced to avoid excessive water loss and carbon fixation rates are
60 reduced due to low gas exchange (Chaves et al., 2002; Haldimann and Feller, 2004; Rodriguez-
61 Dominguez et al., 2016). Climate change can also affect light-dependent processes of
62 photosynthesis in tree seedlings. This is because elevated thermal and/or water stress reduce the
63 flow of electrons in the thylakoid membrane of chloroplasts and this decreases the conversion of
64 light energy into chemical energy for fixing carbon (Maxwell and Johnson, 2000; Baker and
65 Oxborough, 2004). Thus, the combined effects of warmer and drier conditions expected in forest
66 ecosystems can alter water and carbon balances of tree seedlings and reduce their survival.

67 The availability of soil nitrogen is another important factor that can be altered by climate
68 change. Nitrogen is the most important mineral nutrient that regulates plant growth (Poorter et
69 al., 2012) and its natural inputs in forest ecosystems mainly depend on the activity of soil bacteria
70 (Brookshire et al., 2011). These microorganisms are extremely sensitive to changes in soil
71 temperature and moisture and, thus, the expected variations in climatic conditions can modify
72 their efficiency for mineralizing nitrogen (Brookshire et al., 2011; Chen et al., 2011). However,
73 the net effects of climate change on soil nitrogen are still controversial. This is because warming
74 may stimulate the metabolic activity of nitrogen-fixing bacteria and increase soil nitrogen
75 contents (Melillo et al., 2002; Brookshire et al., 2011; Butler et al., 2012), but reductions in soil
76 moisture content due to reduced water inputs are expected to cause the opposite effects (Borken
77 and Matzner, 2009; Chen et al., 2011). Instead, from the perspective of tree seedlings, the
78 combined effects of increasing temperatures and reduced precipitation levels can jeopardize their
79 ability to acquire nitrogen irrespective of whether its concentrations increase or decrease in the
80 future. This is because these climatic conditions can induce strong water deficits and decrease the
81 mobility of the mineral forms of nitrogen in the soil matrix, reducing the capability of plants to
82 uptake this resource through the roots (Rennenberg et al., 2009; He and Dijkstra, 2014). In
83 consequence, nitrogen contents and growth rates of tree seedlings in forest ecosystems are likely
84 to decrease with the advance of climate change.

85 This study assesses how climate change could affect the emergence, survival and performance
86 of recently emerged oak seedlings in a temperate forest of Mexico. For this, we performed a field
87 experiment in which temperature and precipitation were manipulated during the period of the
88 year in which the emergence of oak seedlings occurs, as these early life-cycle stages of oaks are
89 extremely sensitive to changing environmental conditions (Zavala-Chávez and García-Moya,
90 1996; Badano et al., 2011, 2015). In these ecosystems, adult oaks produce acorns (nut-type,

91 single-seeded fruits) in spring and release them in late summer, concurring with the second half
92 of the rainy season (Zavala-Chávez and García-Moya, 1996; González-Salvatierra et al., 2013;
93 Badano et al., 2015). As seeds of Mexican oaks are extremely recalcitrant (embryos die if seed
94 water content drops below 25-30%) and they require elevated hydration levels to germinate
95 (above 80% of seed water content), seedling emergence in the field occurs in short time after
96 acorns are released from parental trees (Zavala-Chávez and García-Moya, 1996; Zavala-Chávez
97 2008). The development of recently emerged oak seedlings mainly depends on the nutritional
98 reserves accumulated in their cotyledons, but they are extremely susceptible to die because of
99 thermal and water stress (Badano et al., 2009; González-Salvatierra et al., 2013). Thus, we
100 hypothesized that warmer and drier conditions expected in climate change scenarios will reduce
101 the emergence and survival of oak seedlings, also decreasing their overall performance. As
102 climate change can reduce nitrogen availability for plants, we also hypothesized that the content
103 of nitrogen in seedlings subjected to climate change conditions will be lower than those of
104 seedling growing under the current climate.

105

106 **2. Materials and Methods**

107 *2.1. Study area*

108 Our experiment was conducted at National Park “El Potosí”, located in the westernmost section
109 of Sierra Madre Oriental (Sierra de Álvarez), state of San Luis Potosí, Mexico (include the file
110 “Pérez Ruiz et al. Experimental sites.kmz” here). Climate in this region is temperate (mean
111 annual temperature 18 °C) and up to 80% of rainfalls occurs between June and November
112 (annual precipitation 500-600 mm), but sparse precipitation events can happen until January
113 (Vargas-Márquez, 1997). Soils are shallow, with maximum depths of 10-15 cm, and vegetation is
114 composed by semi-deciduous oak forests. In these ecosystems, despite the shallowness of soils,

115 tree roots penetrate the bedrock through small fissures, anchoring these organisms to the substrate
116 (Zavala-Chávez and García-Sánchez, 1999). As seedling emergence mainly occurs in the upper
117 section of the soil profile of the forest understory (Badano et al., 2015), we located the
118 experimental site beneath the canopy of a well-preserved oak forest dominated by *Quercus*
119 *crassifolia* Humb. & Bonpl., *Quercus eduardii* Trel., and *Quercus jonesii* Trel. (21°55'10" N,
120 100°19'34" W, 2123 m elevation).

121 To determine how much climate will change in the study site, we computed the difference
122 between current and future values of air temperature and precipitation using the geodatabases of
123 Fernández-Eguiarte et al. (2012, 2014). These geodatabases provide the values of these two
124 variables with a spatial resolution of 1-km² per pixel, where current values are monthly averages
125 that integrate data from 1902 to 2011 (Fernández-Eguiarte et al., 2012) and future values are
126 estimated for each month of the year with the HadGEM2-ES climate change model (Fernández-
127 Eguiarte et al., 2014). Future values of air temperature and precipitation are provided for the
128 period 2021-2040 and consider the four climate change scenarios (RCP2.6, RCP4.5, RCP6.0 and
129 RCP8.5) proposed in the latest report of IPCC (2013). The HadGEM2-ES model is commonly
130 used make climate change predictions across continental Mexico because it has lower deviations
131 than other models when past climatic conditions are estimated, and this reduces uncertainty about
132 the future climate (Fernández-Eguiarte et al., 2014). Because we focused on the earlier life-cycle
133 stages of oaks, differences between current and future values of these climatic conditions were
134 computed for the period of the year in which seedling emergence occurs, comprised between
135 October and February. These estimates indicated that, during the tree recruitment season, air
136 temperature in the study site will increase about 1.3-1.7° C and precipitation will decrease by 11-
137 18% (Table 1).

Climate change scenario	Air temperature (net change)	Precipitation (percent change)
RCP2.6	+1.7 °C	-18%
RCP4.5	+1.2 °C	-14%
RCP6.0	+1.3 °C	-11%
RCP8.5	+1.3 °C	-16%

Table 1. Short term changes (period 2021-2040) in air temperature and precipitation predicted for the study site during the oak recruitment season (October-February). Predictions were performed with the HadGEM2-ES model considering the four climate change scenarios (RCP2.6, RCP4.5, RCP6.0 and RCP8.5) proposed in the latest IPCC report.

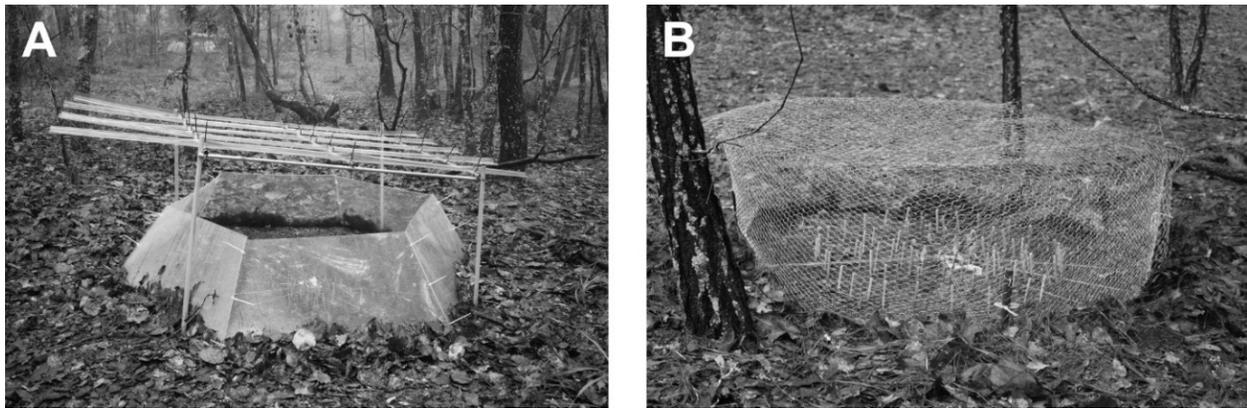
138

139 *2.2. Experimental design*

140 The field experiment was carried out between 17 October 2015 and 27 February 2016,
141 comprising the entire period in which oak seedlings emerge and their early establishment occurs.
142 For this, we draw a quadrant of 50 x 50 m in the study site and randomly selected 20 point-sites
143 within it on September 2015, taking care of maintaining a minimum distance of 5 m among them.
144 Leaf litter and rocks were removed in a radius of 3 m around each of these sites. The half of the
145 sites were randomly assigned to climate change simulation plots (hereafter, CCS plots) where we
146 manipulated temperature and precipitation, while the other half of the sites were assigned to
147 control plots that were maintained under the current environmental conditions. To manipulate
148 temperature in CCS plots we used hexagonal open-top chambers (Marion et al., 1997), which
149 were built with sheets of transparent acrylic (3 mm thick, wavelength transmission 280-750 nm).
150 The resulting open-top chambers were 0.50 m tall, 1.5 m wide at the open-top, and 2.08 m wide
151 at the base (Fig. 1A). Previous studies indicate that this open-top chamber design can increase air
152 temperature by 1-3 °C relative to the external environment, without affecting light transmission
153 within the chambers (Dabros and Fyles, 2010; Aragón-Gastélum et al., 2014). Precipitation
154 within CCS plots was reduced with rainout shelters (Yahdjian and Sala, 2002), which consisted

155 in five U-shaped channels of transparent polycarbonate (1.5 mm thick, 10 cm wide, 3 m length)
156 equidistantly arranged on metallic frames in order to cover 15% of the open-top of chambers
157 (Fig. 1A). Before the experiment, we tested whether rainout shelters reduced precipitation within
158 the ranges predicted for the study site (see above). These trials indicated that these shelters
159 reduced precipitation by 10-20% at each rainfall event. Control plots, on the other hand, were
160 fenced with wire mesh to prevent the access of granivorous and herbivorous vertebrates, such as
161 mice, squirrels and birds (Fig. 1B).

162



163

Fig. 1. Climate change simulation plots (CCS) consisting in open-top chambers covered with rainout shelters (A) and control plots fenced with wire mesh (B).

164

165 Oak seedlings were developed in the field by sowing acorns within the former experimental
166 plots. We only included two dominant oaks of the study site (*Q. crassifolia* and *Q. eduardii*)
167 because the third species (*Q. jonesii*) did not produce acorns in 2015. To gather acorns of *Q.*
168 *crassifolia* and *Q. eduardii*, we marked ten trees of each species with developing fruits on 09
169 August 2015 and removed leaf litter beneath their canopies. On 27 September 2015, after acorns
170 were released from parental trees, we collected 100 mature fruits beneath the canopy of each of
171 them. Because acorns quickly lose viability due to desiccation or infestation by insects, they were

172 taken to the laboratory and the float test was used to separate viable and unviable acorns (Gribko
173 and Jones, 1995). For this, we removed acorn cupules and placed the nuts in 20 L containers
174 filled with water. After 2 h in water, we assumed that sunken acorns were viable and floating
175 acorns were unviable (Gribko and Jones, 1995). This resulted in more than 600 viable acorns of
176 each species, which were stored at 5° C to prevent viability loss due to desiccation.

177 After acorn collection, we weekly visited the study site to assess the moment in which natural
178 emergence of oak seedlings started in the field. This occurred on 14 October 2015 and, at this
179 date, we randomly selected 200 viable acorns of each oak species and placed them in water
180 during 48 h. We did this because acorns require to reach their maximum hydration capacity to
181 germinate (Zavala-Chávez and García-Moya, 1996; Zavala-Chávez, 2008). These acorns were
182 sowed on 17 October 2015. For this, we randomly assigned five controls and five CCS plots to
183 each oak species and sowed 20 acorns within them. Acorns were sowed 2 cm depth to mimic the
184 behavior of scatter-hoarding mice, which are their main secondary dispersers in North American
185 oak forests (Smallwood et al., 2001). Sowing positions were marked with small wood stakes and
186 the emergence of aerial shoots was monitored every seven days until 27 February 2016. At each
187 monitoring date, we also recorded whether emerged seedlings were alive. Seedlings were
188 assumed to have died when their stems and apical buds were completely withered (González-
189 Salvatierra et al., 2013). We also recorded the dates of leaf unfolding on each seedling because, if
190 climate change alters their phenology, leaf age would be required to control for effects of time in
191 the analyses of foliar functional traits described below. To avoid pseudo-replication, leaf ages
192 (i.e., number of days elapsed after leaf unfolding) were averaged across leaves of each oak
193 seedling.

194

195 *2.3. Climate and soil conditions*

196 To determine the extent of the microclimatic changes induced with open-top chambers and
197 rainout shelters, as compared to current climate, we randomly selected three plots of each
198 treatment (controls and CCS plots) and installed automatized sensors to measure temperature and
199 relative humidity of the air, soil temperature and precipitation. Temperature and relative humidity
200 of the air were measured with dual dataloggers fixed on wood stakes 20 cm above ground
201 (HOBO U23-Pro-V2, Onset Computer Corporation, USA), while soil temperature was measured
202 with dataloggers buried 5 cm depth (HOBO Pendant-UA-002, Onset Computer Corporation,
203 USA). In this way, temperature was measured in the environments in which acorns germinate
204 (belowground) and oak seedlings develop (aboveground). These dataloggers were programed to
205 measure climatic variables every hour during the entire experiment. Precipitation was measured
206 with automatized pluviometers programed to record this variable at each rainfall event (HOBO S-
207 RGB-M002, Onset Computer Corporation, USA). All these sensors were arranged at the center
208 of each plot taking care of avoiding interference among them.

209 We also measured soil volumetric water content during the entire experiment. For this, we
210 randomly selected five experimental plots of each climate treatment and marked them. In these
211 plots, we performed six point-measures every week using a time-domain reflectometer
212 (FieldScout TDR 300, Spectrum Technologies, USA) and these values were averaged per plot at
213 each measurement date. This latter variable was measured at 3.8 cm depth, corresponding to the
214 environment in which acorns were sowed and can influence seedling emergence, and 12.0 cm
215 depth, corresponding to the environment from which the roots of emerged seedlings can uptake
216 water and nutrients.

217 To assess whether the climate change induction alters soil nitrogen content, we measured the
218 concentrations of nitrate (NO_3^-) and ammonium (NH_4^+) in controls and CCS plots. We focused
219 on NO_3^- and NH_4^+ because these are the mineral forms of nitrogen that plants can use (Fitter,

220 1997). Concentrations of NO_3^- and NH_4^+ were measured on the five plots of each climate
221 treatment that we selected to perform water content measures. Nevertheless, as measuring NO_3^-
222 and NH_4^+ required to extract soil samples and this could damage oak seedlings, these
223 measurements were only conducted at the beginning (before climate manipulation) and at the end
224 (after climate manipulation) of the experiment. At both times, four topsoil cores (5 cm depth, 2
225 cm width) were collected within each experimental unit and they were pooled in a mixed sample.
226 This sample was meshed (2 mm mesh) and its NO_3^- content was determined in the field with
227 portable ion-selective meters (LAQUATwin NO_3^- meter B-742, Horiba, Japan). This procedure
228 was repeated three times for each soil sample and NO_3^- readings were averaged. The remaining
229 fraction of each soil sample was placed in a hermetic plastic bag and stored in ice to reduce the
230 metabolic activity of soil microorganism. After arriving to the laboratory (about 3 h after soil
231 collection), these soil samples were spread on aluminum sheets and dried in an air-forced stove at
232 40°C until their weight remained constant (about 48 h). Once samples were fully dry, we took 5
233 g of soil and NH_4^+ was extracted with 2 M potassium chloride solution. These extracts were
234 treated with Berthelot's reagent and read in a UV-visible spectrophotometer (Genesys 10S,
235 Thermo Fisher Scientific). This procedure was repeated three times for each soil sample and
236 NH_4^+ values were averaged.

237

238 *2.4. Oak seedling responses*

239 At the end of the experiment, ungerminated acorns were recovered from the experimental plots
240 and taken to the laboratory to determine their viability. For this, we made a small incision on the
241 pericarp of acorns and incubated them in 1% triphenyltetrazolium chloride solution during 24 h
242 in dark. If seed embryos are alive, the colorless tetrazolium salt reacts with the hydrogen released
243 by active dehydrogenase enzymes and this generates formazan, a non-diffusible red dye (Bewley

244 et al., 2013). As no recovered acorn was viable (i.e., the embryos were not red-stained), we
245 assumed that no additional germination would occur after the experiment was finished. These
246 data were used to compare the proportion of germinated acorns and surviving seedlings between
247 controls and CCS plots. At this time, we also measured the length of the aerial shoots of all
248 surviving seedlings (i.e., length from the ground to the apical bud) with a digital caliper. These
249 data were used to estimate their net growth rates as the ratio between the length of the aerial
250 shoot and the age of each seedling (i.e., the number of days elapsed after shoot emergence) and
251 these values were compared between climatic treatments.

252 We later assessed a series of foliar functional responses on three surviving seedlings from each
253 experimental plot to determine whether climate change can affect their performance. All these
254 seedlings had at least three mature leaves without symptoms of wilting or insect damage, which
255 were marked and numbered with small paper tags. In the field, we used a portable fluorometer
256 (MINI-PAM II, Heinz Walz, Germany) to assess foliar temperature and photosynthetic photon
257 flux density (*PPFD*) on each leaf, and estimated the effective quantum yield (ϕ_{PSII}) by measuring
258 their chlorophyll fluorescence. Values of ϕ_{PSII} were computed as $(F'_m - Ft) / F'_m$, where F_t is
259 the fluorescence emitted by leaves under steady-state illumination and F'_m is their maximum
260 fluorescence when a saturating pulse of actinic light ($\approx 2500 \mu\text{mol photon m}^{-2} \text{s}^{-1}$, in our case) is
261 superimposed on the environmental light level (Maxwell and Johnson, 2000). These
262 measurements were conducted between 10:00 and 12:00 h because, after midday, the elevated
263 levels of light could mask the effects of other environmental factors on photosynthetic quantum
264 yield (Franco and Lüttge, 2002). Values of leaf temperature, *PPFD* and ϕ_{PSII} were averaged
265 across leaves belonging to the same seedling to avoid pseudo-replication.

266 Tagged leaves were later harvested, stored in hermetic plastic bags and placed in ice to reduce
267 water loss. These leaves were weighed immediately after they arrived at the laboratory (about 3 h
268 after harvesting) to determine their fresh biomass (*FB*). After that, leaves were dried in an air-
269 forced stove at 60° C until their dry biomass (*DB*) remained constant (about 24-36 h). Values of
270 *FB* and *DB* were determined using an analytical balance with resolution of 0.1 mg (Cubis,
271 Sartorius, Germany) and used to estimate the percent water content (*PWC*) of leaves as
272 $(FB - DB) / FB$. These values were averaged across the leaves of each seedling and used as
273 surrogate of their water status at the moment of harvesting (Gond et al., 1999). Further, to assess
274 whether climate change can affect the carbon and nitrogen balance of recently emerged oak
275 seedlings, dried leaves were ground in a ball mill (MM200, Retsch, Germany) and processed to
276 determine the percent contents of carbon and nitrogen with an elemental analyzer (ECS 4010,
277 Costech, USA). Because of their small size, leaves collected on each seedling were pooled in a
278 single sample to reach the minimum biomass (0.8 g) required for these analyses.

279

280 2.5. Statistical analysis

281 We used repeated measures ANOVAs to compare all environmental variables between climate
282 treatments. For these analyses, values of air and soil temperature, as well as values of relative
283 humidity of the air, were averaged at each week of the experiment for each datalogger (n = 3 per
284 climate treatment; repeated measures = 19 weeks). On the other hand, precipitation recorded by
285 each pluviometer was accumulated for every week of the experiment (n = 3 per climate
286 treatment) but, as ANOVAs does not admit groups with zero variance (Zar, 2010), we excluded
287 those weeks with no rainfalls from this analysis (repeated measures = 11 weeks). Comparisons of
288 soil water contents at 3.8 and 12.0 cm depth were conducted with the values measured every
289 week of the experiment (n = 5 per climate treatment; repeated measures = 19 weeks), while soil

290 concentrations of NO_3^- and NH_4^+ were compared by only including the two dates in which these
291 variables were measured ($n = 5$ per climate treatment; repeated measures = 2 dates).

292 The proportion of germinated acorns and surviving seedlings were compared between climate
293 treatments and oak species with two-way factorial ANOVA (2 climate treatments x 2 oak species
294 = 4 treatments in total; $n = 5$ per treatment). The identity of oaks was included in these analyses
295 because responses to climate change may vary between species. The same analyses were used to
296 compare the age of seedlings, their net growth rates and leaf ages, *PWC*, foliar temperature,
297 *PPFD*, ϕ_{PSII} , and foliar contents of nitrogen and carbon. The values of all these variables were
298 averaged across seedlings from the same experimental unit to avoid pseudo-replication. All
299 datasets that support the former analyses were deposited in Mendeley Data
300 (<http://dx.doi.org/10.17632/bsvxh88ypz.1>).

301

302 **3. Results**

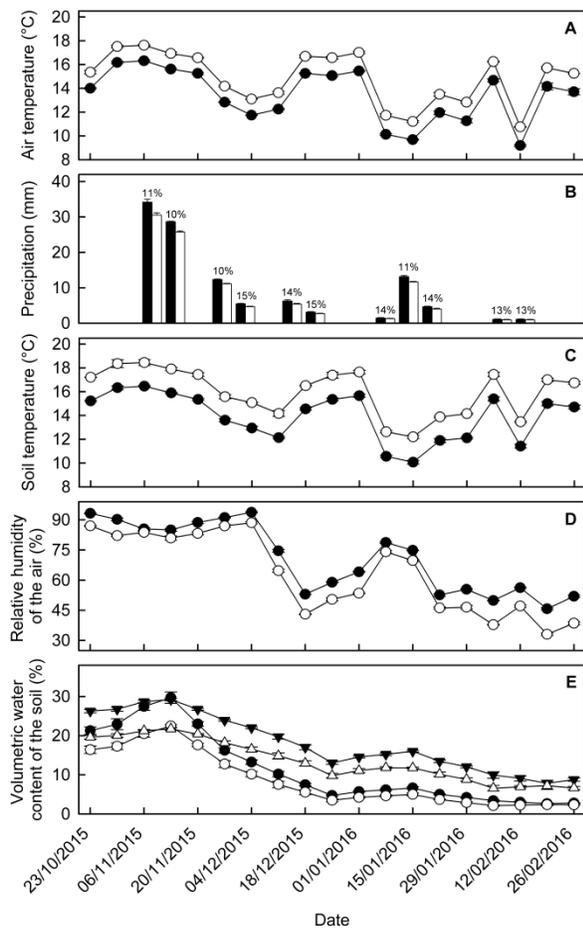
303 *3.1. Climate and soil conditions*

304 Air temperature was higher in CCS plots than in controls across the entire experiment, while the
305 converse occurred with precipitation (Table 2). Differences in air temperature between treatments
306 increased from the beginning to the end of the experiment; CCS plots were about 1.3°C warmer
307 than controls until 18 December 2015, but this difference increased up to 1.5°C after that date
308 (Fig. 2A). Precipitation was 10-11% lower in CCS plots than in controls on those weeks where
309 rainfalls surpassed 10 mm, while CCS plots received 14-15% less precipitation on those weeks
310 where rainfalls were below that value (Fig. 1B).

Environmental variable	Climate treatment	Time (weeks)	Interaction climate treatment-time
Air temperature	$F_{(4,1)} = 853.759^*$	$F_{(18,72)} = 5178.385^*$	$F_{(18,72)} = 3.163^*$

Precipitation	$F_{(1,4)} = 339.846^*$	$F_{(8,32)} = 10846.486^*$	$F_{(8,32)} = 31.435^*$
Soil temperature	$F_{(1,4)} = 5310.901^*$	$F_{(18,72)} = 1482.021^*$	$F_{(18,72)} = 0.229$
Air relative humidity	$F_{(1,4)} = 1076.431^*$	$F_{(18,72)} = 14174.897^*$	$F_{(18,72)} = 109.919^*$
Soil water content-3.8 cm depth	$F_{(1,8)} = 180.184^*$	$F_{(18,144)} = 1722.143^*$	$F_{(18,144)} = 32.258^*$
Soil water content-12.0 cm depth	$F_{(1,8)} = 743.359^*$	$F_{(18,144)} = 1522.408^*$	$F_{(18,144)} = 35.600^*$
Soil nitrate concentration	$F_{(1,8)} = 181.203^*$	$F_{(1,8)} = 562.389^*$	$F_{(1,8)} = 299.475^*$
Soil ammonium concentration	$F_{(1,8)} = 405.474^*$	$F_{(1,8)} = 1111.181^*$	$F_{(1,8)} = 577.643^*$

Table 2. Results of repeated measures ANOVA addressed to compare climatic variables and soil features between climate treatments. The table shows the F-values obtained for each factor (climate treatments and time) and their interaction. Significant effects on each response variable are indicated with an asterisk (critical $\alpha = 0.05$).



311

Fig. 2. Average values ($\pm 95\%$ C.I.) of air temperature (A), precipitation (B), soil temperature (C), relative humidity of the air (D), and soil water content (E) at each week of the experiment in controls (black symbols) and CCS plots (white symbols). Numbers above precipitation bars indicate the percent difference between climate treatments. Soil water content is reported at 3.8 cm (circles) and 12.0 cm depth (triangles).

312

313 Soil temperature, relative humidity of the air and soil water contents also differed between

314 climate treatments (Table 2). Soil temperature within CCS plots was about 2.0° C higher than in

315 controls and this difference was sustained along the entire experiment (Fig. 2C). Air relative

316 humidity was always lower in CCS plots than in controls, but these differences decreased in rainy

317 weeks (Fig. 2D). Water contents at both, 3.8 cm and 12.0 cm depth, were lower in CCS plots than

318 in controls and these differences increased when rainfalls decreased (January-February; Fig. 2E).

319 Soil concentrations of NO_3^- and NH_4^+ differed between climate treatments and sampling dates

320 (Table 2). At the beginning of the experiment (i.e., before climate manipulation was applied),

321 there were no differences in the concentrations of NO_3^- (Fig. 3A) or NH_4^+ (Fig. 3B) between

322 plots assigned to controls and CCS. However, concentrations of both mineral nutrients were

323 lower in CCS plots than in controls by the end of the experiment (Fig. 3).

324

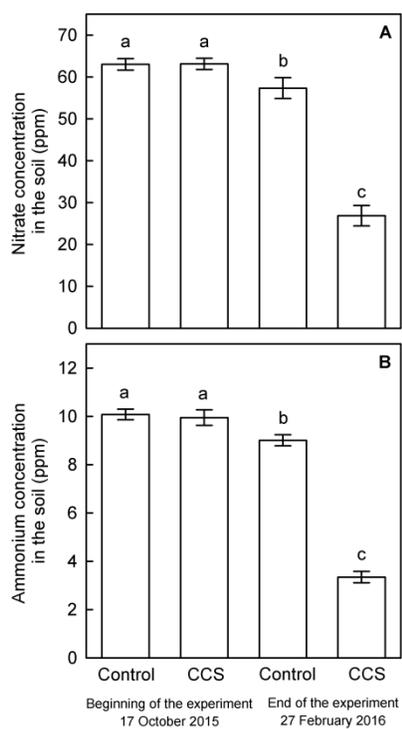


Fig. 3. Soil concentrations (average \pm 95% C.I.) of nitrate (A) and ammonium (B) in controls and CCS plots at the beginning (17 October 2015) and at the end of the experiment (27 February 2016). Different letters above the bars indicate significant differences between treatments (Tukey tests critical $\alpha = 0.05$).

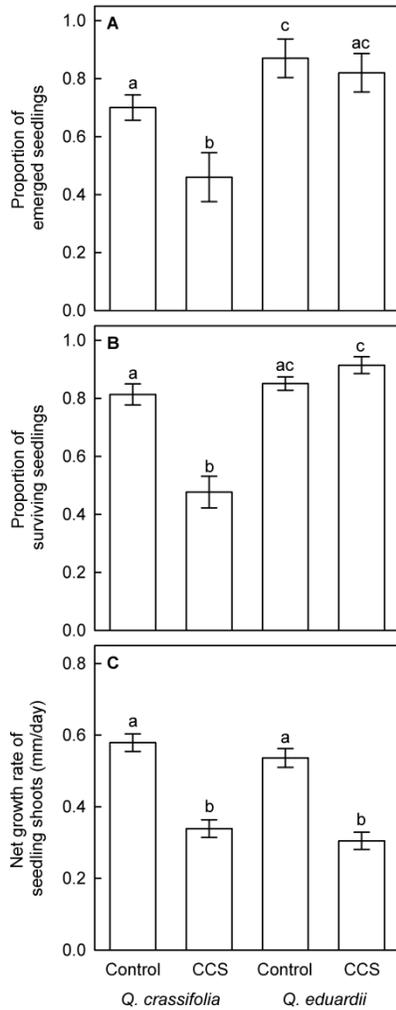
326

327 3.2. Seedling responses

328 The proportion of emerged and surviving seedlings differed between climate treatments
329 (emergence: $F_{(1,16)} = 18.086$, $p = 0.006$; survival: $F_{(1,16)} = 50.585$, $p < 0.001$) and oak species
330 (emergence: $F_{(1,16)} = 60.409$, $p < 0.001$; survival: $F_{(1,16)} = 152.669$, $p < 0.001$). Interactive effects
331 of these two factors (climate treatment \times oak species) were also found on both response variables
332 (emergence: $F_{(1,16)} = 7.763$, $p = 0.013$; survival: $F_{(1,16)} = 108.411$, $p < 0.001$). Emergence of *Q.*
333 *crassifolia* seedlings was lower in CCS plots than in controls, while emergence of *Q. eduardii*
334 seedlings did not differ between climate treatments (Fig. 4A). However, in both controls and
335 CCS plots, the proportion of emerged seedlings was higher for *Q. eduardii* than for *Q. crassifolia*
336 (Fig. 4A). The proportion of surviving seedlings in controls did not differ between oak species,
337 but seedlings of *Q. eduardii* had higher survival than those of *Q. crassifolia* in CCS plots (Fig.
338 4B).

339 Age of surviving seedlings neither differed between climate treatments nor oak species,
340 averaging 102 days across all experimental units (data not shown). Conversely, shoot growth
341 rates differed between climate treatments ($F_{(1,16)} = 346.225$, $p < 0.001$) and oak species ($F_{(1,16)} =$
342 9.181 , $p = 0.008$), but no interactive effects between these two factors were found ($F_{(1,16)} = 0.113$,
343 $p = 0.741$). This analysis indicated that seedlings of both, *Q. eduardii* and *Q. crassifolia*, had
344 higher shoot growth rates in controls than in CCS plots (Fig. 4C).

345



346

Fig. 4. Proportion (average \pm 95% C.I.) of emerged seedlings (A) and surviving seedlings (B) of *Q. crassifolia* and *Q. eduardii* in controls and CCS plots. The figure also shows the average (\pm 95% C.I.) net growth rates of seedling shoots (C) in both climate treatments. Different letters above the bars indicate significant differences between treatments (Tukey tests critical $\alpha = 0.05$).

347

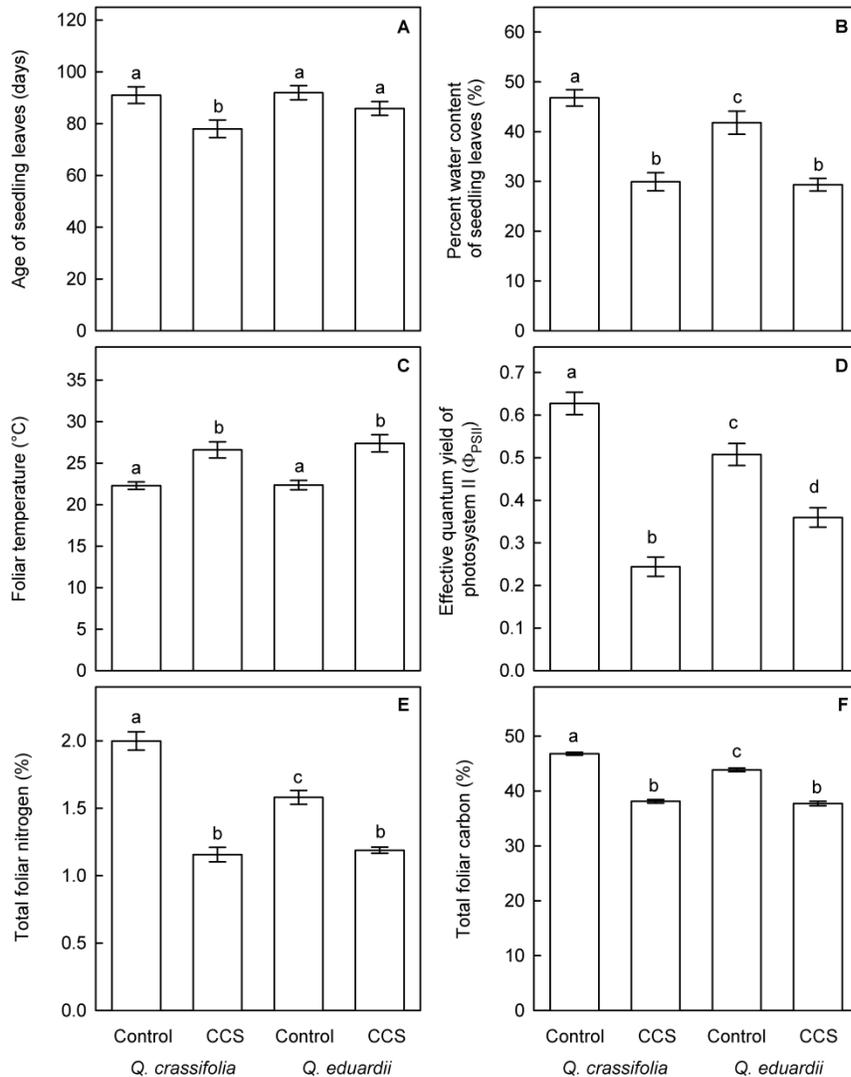
348 Age of seedling leaves differed between climate treatments ($F_{(1,16)} = 38.267$, $p < 0.001$) and
 349 oak species ($F_{(1,16)} = 8.152$, $p = 0.011$), and significant effects were also found for the interaction
 350 of these two factors ($F_{(1,16)} = 5.060$, $p = 0.039$). In this case, leaf unfolding occurred later in *Q.*
 351 *crassifolia* seedlings than in seedlings from all the other treatments (Fig. 5A). For this reason,
 352 leaf age was included as continuous concomitant variable in the analyses comparing the other

353 foliar functional traits (i.e., we used ANCOVA instead of ANOVA). Values of *PWC* were not
 354 affected by leaf age, but significant differences were found between climate treatments and oak
 355 species (Table 3). Leaves of both oak species had lower *PWC* in CCS plots than in controls and
 356 these differences were higher for *Q. crassifolia* than for *Q. eduardii* (Fig. 5B). Neither leaf age
 357 nor oak species affected foliar temperatures (Table 3), but both oak species had higher foliar
 358 temperatures CCS plots than in controls (Fig. 5C).

Environmental variable	Leaf age	Climate treatment	Oak species	Interaction climate treatment-oak species
Leaf percent water content	$F_{(1,15)} = 1.501$	$F_{(1,15)} = 60.208^*$	$F_{(1,15)} = 10.332^*$	$F_{(1,15)} = 2.299$
Foliar temperature	$F_{(1,15)} = 0.285$	$F_{(1,15)} = 32.095^*$	$F_{(1,15)} = 1.331$	$F_{(1,15)} = 1.061$
ϕ_{PSII}	$F_{(1,15)} = 1.397$	$F_{(1,15)} = 115.490^*$	$F_{(1,15)} = 0.693$	$F_{(1,15)} = 60.504^*$
Total foliar nitrogen	$F_{(1,15)} = 0.640$	$F_{(1,15)} = 142.040^*$	$F_{(1,15)} = 40.424^*$	$F_{(1,15)} = 48.803^*$
Total foliar carbon	$F_{(1,15)} = 0.034$	$F_{(1,15)} = 501.276^*$	$F_{(1,15)} = 60.225^*$	$F_{(1,15)} = 37.492^*$

Table 3. Results of ANCOVA comparing leaf traits and associated variables between climate treatments and oak species. The table shows the F-values obtained for each factor (climate treatments and time) and their interaction. Significant effects on each response variable are indicated with an asterisk (critical $\alpha = 0.05$). In these analyses, leaf age was included as concomitant continuous variable.

359
 360 Values of *PPFD* neither differed between climate treatments nor oak species (data not shown),
 361 indicating that leaves of all seedlings were exposed to similar levels of light ($578.606 \mu\text{mol m}^{-2} \text{s}^{-1} \pm 6.643$ 95% I.C.) during the chlorophyll fluorescence measurements. Values of ϕ_{PSII} were not
 362 affected by leaf age and they did not differ between oak species (Table 3). However, seedlings
 363 both of oak species displayed higher ϕ_{PSII} values in controls than CCS plots (Table 3; Fig. 5D).
 364
 365



366

Fig. 5. Foliar functional traits (average \pm 95% C.I.) of *Q. crassifolia* and *Q. eduardii* seedlings from controls and CCS plots, including age of leaves (A), leaf percent water content (B), foliar temperature (C), effective quantum yield of photosystem II (D), foliar content of nitrogen (E) and foliar content of carbon (F). Different letters above the bars indicate significant differences between treatments (Tukey tests critical $\alpha = 0.05$).

367

368 No effects of leaf age were found on foliar contents of nitrogen and carbon, but values of these
 369 two variables differed between climate treatments and oak species (Table 3). Both oak species
 370 had higher contents of foliar nitrogen (Fig. 5E) and foliar carbon (Fig. 5F) in controls than in

371 CCS plots, and these differences were larger in seedlings of *Q. crassifolia* than in seedlings of *Q.*
372 *eduardii*.

373

374 **4. Discussion**

375 Our study indicates that increasing temperature and drought will impair the development of
376 recently emerged oak seedlings in the field and, as far as we are aware, this is the first
377 experimental study addressing these issues in temperate forests of North America. Indeed, as
378 open-top chambers and rainout shelters properly simulated the conditions of temperature and
379 rainfall expected in the short term (2021-2040), our experimental results suggest that these
380 negative impacts of climate change would occur much sooner than predicted by other studies,
381 which have used species distribution models to simulate the future distribution of tree species and
382 proposed that climate change effects on these forests will be evident after the middle of this
383 century (Gómez-Mendoza and Arriaga, 2007; Mclaughlin and Zavaleta, 2012). The induction of
384 higher temperature and lower rainfall in the field also modified other environmental variables that
385 can impair the development of oak seedlings, and it is important to note that the future shifts in
386 the values of these variables are not directly predicted by climate change models. For instance,
387 we observed an overall decrease of air relative humidity within CCS plots, which can be
388 attributed to the direct inverse relationship between the values of this variable and air temperature
389 (Hardwick-Jones et al., 2010). Further, higher soil temperatures resulting from warming could
390 have increased soil water evaporation within CCS plots (Parlange et al., 1998) and this, together
391 with the smaller water inputs due to rainout shelters, may be responsible for the lower soil water
392 contents recorded in these experimental units. In this way, the climatic changes that will occur
393 during the next decades in temperate forest due to the increased emission of greenhouse gases
394 may reduce the recruitment rates of oaks in these ecosystems. Nevertheless, our results also

395 indicated that the two oak species included in this study somewhat differed in their responses to
396 climate change.

397 Emergence and survival of *Q. crassifolia* seedlings were lower in CCS plots than in controls,
398 while these differences were not found for *Q. eduardii*. This suggests that the early life-cycle
399 stages of *Q. crassifolia* would be more sensitive to climate change than those of *Q. eduardii*.
400 Because acorn of most oak species quickly lose viability when external factors enhance seed
401 water loss (Zavala-Chávez, 2008; Joët et al., 2013), the higher soil temperatures induced in CCS
402 plots may have accelerated acorn desiccation and, consequently, this may have reduced the
403 emergence of *Q. crassifolia* seedlings. Further, as oak seeds require elevated hydration levels to
404 stimulate the development of their embryos (Zavala-Chávez and García-Moya, 1996), the lower
405 soil moisture contents in CCS plots may have prevented that the acorns of *Q. crassifolia* reached
406 the water contents they needed to germinate. Thus, the combined effects of higher temperature
407 and reduced rainfall may be responsible of the lower proportion of emerged seedlings of this oak
408 species in CCS plots.

409 The lower survival of recently emerged *Q. crassifolia* seedlings in CCS plots can also be
410 attributed to the combination of increased warming and drought. Several authors have reported
411 that, under the current climatic conditions, oak seedlings quickly wilt if they face thermal and/or
412 water stress during their recruitment season, which drastically increases their mortality in the
413 field (Rey-Benayas et al., 2005; González-Salvatierra et al., 2013; Montes-Hernández and López-
414 Barrera, 2013; Badano et al., 2011, 2015). In our study site, climate change models predict that
415 these two stress factors will intensify in the near future, and our experimental results indicate that
416 such increased environmental harshness will reduce the survival of *Q. crassifolia* seedlings.
417 However, neither emergence nor survival of *Q. eduardii* seedlings were reduced within CCS

418 plots, as compared to controls, suggesting that this oak species may be able to deal with climate
419 change.

420 These differences in seedling emergence and survival between *Q. crassifolia* and *Q. eduardii*
421 could be attributed to differential capabilities they have for recruiting in contrasting habitats. In
422 temperate oak forest of Mexico, *Q. crassifolia* is considered a late successional species whose
423 seedlings mainly recruit in shaded understory habitats and, although they have an elevated
424 competitive ability for soil resources in comparison with seedlings of other tree species, they do
425 not tolerate elevated temperatures or water shortage (Castillo-Lara et al., 2008). Instead, even
426 when *Q. eduardii* is also a conspicuous species within these forest, this oak is considered a
427 pioneer tree because it colonizes disturbed open areas, where temperature and drought are much
428 higher than beneath forest canopies (Alfonso-Corrado et al., 2007). Thus, lack of effects of
429 climate change on the emergence and survival of *Q. eduardii* seedlings may be due to adaptations
430 they have for establishing in harsh environments.

431 Despite the higher emergence and survival of *Q. eduardii* seedling in CCS plots, as compared
432 with those of *Q. crassifolia*, our results suggest that climate change will negatively affect the
433 development of the early life-cycle stages in both species. For instance, aboveground growth of
434 these seedlings was about twice higher in controls than in CCS plots. This contrasts with the
435 results of experiments that have manipulated temperature in cold and temperate forests, which
436 have indicated that warming can hasten aerial shoot development of tree seedlings (Danby and
437 Hik, 2007; Xu et al., 2012; Piper et al., 2013). Nevertheless, experiments conducted in arid and
438 semi-dry environments have reported that the occurrence of drought events during the plant
439 recruitment season can strongly impair the growth rates of tree seedling (Valladares and Sánchez-
440 Gómez, 2006; Mclaughlin and Zavaleta, 2012). Thus, the lower growth rates of oak seedlings in
441 CCS plots could be due to this latter factor, as the environmental variables measured in these

442 experimental units suggested that the combined effect of increased temperatures and reduced
443 rainfalls generated strong soil water deficits.

444 Climate manipulation within CCS plots also induced other detrimental effects on oak
445 seedlings, including reduced leaf water content (*PWC*), lower carbon accumulation in their
446 leaves, and diminished photosynthetic quantum yield (ϕ_{PSII}). The lower *PWC* of seedlings in CCS
447 plots can be linked with their higher foliar temperatures, which may have increased water loss
448 through transpiration (Haldimann and Feller, 2004; Rodriguez-Dominguez et al., 2016). Further,
449 the lower air relative humidity within CCS plots may have reduced the boundary vapor layer of
450 leaves, also enhancing foliar transpiration (Martin et al., 1999; Will et al., 2013). Since leaf
451 dehydration induces stomatal closure (Brodribb and Holbrook, 2003), these lower *PWC* could
452 have led to reduced gas exchange and this may partially explain why seedlings from CCS plots
453 had lower carbon contents than those from the controls. The reduced values of ϕ_{PSII} in these
454 seedlings can also be related with their lower carbon contents, as thermal and/or water stress
455 decrease ϕ_{PSII} in plants (Xu and Zhou, 2006; González-Salvatierra et al., 2013; Aragón-Gastélum
456 et al., 2014; Badano et al., 2015) and causes energetic deficits for fixing carbon through the
457 Calvin cycle (Baker and Oxborough 2004; Xu and Zhou, 2006). Therefore, our results suggest
458 that climate change can also impair the photosynthetic efficiency of oak seedlings.

459 The lower contents of NO_3^- and NH_4^+ in the soil CCS plots concur with the results of other
460 experiments conducted in forest ecosystems, which showed that climate change may cause
461 nitrogen deficits for plants (Rennenberg et al., 2009; Chen et al., 2011; Roa-Fuentes et al., 2015).
462 Decreases in soil nitrogen content under climate change conditions have been mainly attributed to
463 the detrimental effects of water deficits on the metabolic activity of nitrogen-fixing bacteria,
464 rather than increasing temperatures (Borken and Matzner, 2009; Chen et al., 2011). This

465 decreased availability of nitrogen in the soil may explain the lower foliar nitrogen contents of
466 seedlings that grew within CCS plots. However, it is also important to note that the lower soil
467 moisture in these plots may have reduced the mobility of this mineral nutrient in the soil matrix
468 and, consequently, impaired its uptake through the roots of seedlings. These effects of drought on
469 the activity of nitrogen-fixing organisms and/or the mobility of nitrogen in the soil matrix, may
470 explain the lower growth rates of oak seedlings in CCS plots, as nitrogen is a key nutrient that
471 regulates plant development (Rennenberg et al., 2009; He and Dijkstra, 2014).

472 Overall, this study indicates that climate change conditions that will occur across temperate
473 forest of North America have the potential to reduce the recruitment of oak species. Under the
474 current climate, elevated temperatures and drought events that occur during the growing season
475 of oaks constitute important ecological barriers for their establishment in temperate forest
476 (Zavala-Chávez and García-Moya, 1996; González-Salvatierra et al., 2013; Montes-Hernández
477 and López-Barrera, 2013; Badano et al., 2009, 2011, 2015), and our results reveal that the
478 strengthening of these stress factors will impair even more the development of recently emerged
479 oak seedlings. This, in turn, may have strong detrimental effects on the natural regeneration
480 processes that regulate the maintenance of these forests, and this can threaten the provisioning of
481 ecosystem services that support human well-being.

482

483 **5. Conclusion**

484 Our results indicate that the advance of climate can reduce the emergence, establishment, growth
485 and physiological performance of oak seedlings in temperate forest. Nevertheless, the differences
486 in seedling emergence and survival that we observed between the two oaks included in this study
487 suggest that some species could be more susceptible to climate change (e.g., *Q. crassifolia*) than
488 others (e.g., *Q. eduardii*). On this issue, extensive field experiments are still required to identify

489 what tree species are vulnerable to climate change and what species will be able to cope with
490 warming and drought. This would allow to focus conservation actions on those species highly
491 sensitive to climate change, promoting their assisted migration towards zones climatically
492 suitable for their development and hence ensure their prevalence. Conversely, those species with
493 elevated resilience to climate change could be used for developing sustainable forestry and/or
494 addressing restoration programs to recover forests in degraded areas.

495

496 **Statement of authorship**

497 CLPR and EIB conceptualized the study, designed the field experiment, gathered the data and
498 performed the statistical analyses; JPRO, PDS, JF, DD and JAFC contributed with the
499 measurement of seedling variables and foliar traits in the field and the laboratory; all authors
500 contributed interpret the results and co-wrote the article.

501

502 **Acknowledgements**

503 We thank CONANP-Mexico and people of Ejido Cañanda Grande for the logistic support
504 provided during the field experiment. This work was supported by Consejo Nacional de Ciencia
505 y Tecnología de México [grant SEP-CONACYT CB-2013/221623 to EIB and PhD studentship
506 150830 to CLPR].

507

508 **References**

509 Alfonso-Corrado, C., Clark-Tapia, R., Mendoza, A., 2007. Demography and management of two
510 clonal oaks: *Quercus eduardii* and *Q. potosina* (Fagaceae) in central México. Forest Ecol.
511 Manag. 251, 129-141.

512 Aragón-Gastélum, J.L., Flores, J., Yáñez-Espinosa L., Badano, E., Ramírez-Tobías, H.M.,
513 Rodas-Ortíz, J.P., González-Salvatierra, C., 2014. Induced climate change impairs
514 photosynthetic performance in *Echinocactus platyacanthus*, an especially protected
515 Mexican cactus species. *Flora* 209, 499-503.

516 Badano, E.I., Pérez, D., Vergara, C.H., 2009. Love of nurse plants is not enough for restoring oak
517 forests in a seasonally dry tropical environment. *Restor. Ecol.* 17, 571-576.

518 Badano, E.I., Samour-Nieva, O.R., Flores, J., 2011. Emulating nurse plants to restore oak forests.
519 *Ecol. Eng.* 37, 1244-1248.

520 Badano, E.I., Samour-Nieva, O.R., Flores, J., Douterlungne, D., 2015. Microclimate and seeding
521 predation as drivers of tree recruitment in human-disturbed oak forests. *Forest Ecol.*
522 *Manag.* 356, 93-100.

523 Baker, N.R., Oxborough, K., 2004. Chlorophyll fluorescence as a probe of photosynthetic
524 productivity, in: Papageorgiou, G., Govindjee (Eds.), *Chlorophyll a Fluorescence: A*
525 *Signature of Photosynthesis*. Springer, Dordrecht, pp. 65-82.

526 Bewley, J.D., Bradford, K.J., Hilhorst, H.W.M., Nonogaki, H., 2013. *Seeds: Physiology of*
527 *Development, Germination and Dormancy*, third ed. Springer, New York.

528 Borken, W., Matzner, E., 2009. Reappraisal of drying and wetting effects on C and N
529 mineralization and fluxes in soils. *Glob. Change Biol.* 15, 808-24

530 Brodribb, T.J., Holbrook, N.M., 2003. Stomatal closure during leaf dehydration, correlation with
531 other leaf physiological traits. *Plant Physiol.* 132, 2166-73.

532 Brookshire, E.N.J., Gerber, S., Webster, J.R., Vose, J.M., Swank, W.T., 2011. Direct effects of
533 temperature on forest nitrogen cycling revealed through analysis of long-term watershed
534 records. *Glob. Change Biol.* 17, 297-308.

535 Butler, S.M., Melillo, J.M., Johnson J., Mohan, J., Steudler, P.A., Lux, H., Burrows, E., Smith,
536 R.M., Vario, C.L., Scott, L., Hill, T.D., Aponte, N., Bowles, F., 2012. Soil warming alters
537 nitrogen cycling in a New England forest: Implications for ecosystem function and
538 structure. *Oecologia* 168, 819-28.

539 Castillo-Lara, P., Flores-Cano, J.A., Aguirre-Rivera, J.R., Yeaton, R.I., 2008. Dinámica
540 sucesional del encinar de la Sierra Álvarez, San Luis Potosí, México. *Madera y Bosques*
541 14, 21-35.

542 Chaves, M.M., Pereira, J.S., Maroco, J., Rodrigues, M.L., Ricardo, C.P.P., Osório, M.L.,
543 Carvalho, I., Faria, T., Pinheiro, C., 2002. How plants cope with water stress in the field?
544 Photosynthesis and growth. *Ann. Bot.* 89, 907-916.

545 Chen, Y.T., Borke, W., Stange, C.F., Matzner, E., 2011. Effects of decreasing water potential on
546 gross ammonification and nitrification in an acid coniferous forest soil. *Soil Biol. Biochem.*
547 43, 333-38.

548 Clark, J.S., Iverson, L., Woodall, C.W., Allen, C.D., Bell, D.M., Bragg, D.C., D'Amato, A.W.,
549 Davis F.W., Hersh M.H., Ibanez, I., Jackson S.T., Matthews, S., Pederson, N., Peters, M.,
550 Schwartz M.W., Waring, K.M., Zimmermann N.E., 2016. The impacts of increasing
551 drought on forest dynamics, structure, and biodiversity in the United States. *Glob. Change*
552 *Biol.* 22, 2329-52.

553 Dabros, A., Fyles, J.W., 2010. Effects of open-top chambers and substrate type on
554 biogeochemical processes at disturbed boreal forest sites in northwestern Quebec. *Plant*
555 *Soil* 327, 465-79.

556 Danby, R.K., Hik, D.S., 2007. Responses of white spruce (*Picea glauca*) to experimental
557 warming at a subarctic alpine treeline. *Glob. Change Biol.* 13, 437-51.

558 Fernández-Eguiarte, A., Romero-Centeno, R., Zavala-Hidalgo, J., 2012. Atlas Climático Digital
559 de México y Áreas Adyacentes, Vol. 1. Universidad Nacional Autónoma de México,
560 Mexico City.

561 Fernández-Eguiarte, A., Romero-Centeno, R., Zavala-Hidalgo, J., Kucieńska, B., 2014. Atlas
562 Climático Digital de México y Áreas Adyacentes, Vol. 2. Universidad Nacional Autónoma
563 de México, Mexico City.

564 Fitter, A., 1997. Nutrient acquisition, in: Crawley, M.J. (Ed.), Plant Ecology. Blackwell, Oxford,
565 pp. 51-72.

566 Franco, A., Lüttge, U., 2002. Midday depression in savanna trees: Coordinated adjustments in
567 photochemical efficiency, photorespiration, CO₂ assimilation and water use efficiency.
568 Oecologia 131, 356-365

569 Gómez-Mendoza, L., Arriaga, L., 2007. Modeling the effect of climate change on the distribution
570 of oak and pine species of Mexico. Conserv. Biol. 21, 1545-55.

571 Gond, V., De Pury, D.D.G., Veroustraete, F., Ceulemans, R., 1999. Seasonal variations in leaf
572 area index, leaf chlorophyll, and water content; scaling-up to estimate fAPAR and carbon
573 balance in a multilayer, multispecies temperate forest. Tree Physiol. 19, 673-679.

574 González-Salvatierra, C., Badano, E.I., Flores, J., Rodas, J.P., 2013. Shade shelters increase
575 survival and photosynthetic performance of oak transplants at abandoned fields in semi-arid
576 climates. J. For. Res. 24, 23-28.

577 Gribko, L.S., Jones, W.E., 1995. Test of the float method of assessing northern red oak acorn
578 condition. Tree Plant. Notes 46, 143-47.

579 Haldimann, P., Feller, U., 2004. Inhibition of photosynthesis by high temperature in oak
580 (*Quercus pubescens* L.) leaves grown under natural conditions closely correlates with a

581 reversible heat-dependent reduction of the activation state of ribulose-1,5-bisphosphate
582 carboxylase/oxygenase. *Plant Cell Environ.* 27, 1169-83.

583 Hardwick-Jones, R., Westra, S., Sharma, A., 2010. Observed relationships between extreme sub-
584 daily precipitation, surface temperature, and relative humidity. *Geophys. Res. Lett.* 37,
585 L22805.

586 He, M., Dijkstra, F.A., 2014. Drought effect on plant nitrogen and phosphorus: A meta-analysis.
587 *New Phytol.* 204, 924-31.

588 IPCC, 2013. Fifth Assessment Report of the Intergovernmental Panel on Climate Change -
589 Climate Change 2013: The Physical Science Basis. Cambridge University Press,
590 Cambridge.

591 Joët, T., Ourcival, J.M., Dussert, S., 2013. Ecological significance of seed desiccation sensitivity
592 in *Quercus ilex*. *Ann. Bot.* 111, 693-701.

593 Marion, G.M., Henry, G.H.R., Freckman, D.W., Johnstone, J., Jones, C.G., Jones M.H.,
594 Lévesque, E., Molau, U., Molgaard, P., Parsons, A.N., Svoboda J., Virginia R.A., 1997.
595 Open-top designs for manipulating field temperature in high-latitude ecosystems. *Glob.*
596 *Change Biol.* 3, 20-32.

597 Martin, T.A., Hinckley, T.M., Meinzer, F.C., Sprugel, D.G., 1999. Boundary layer conductance,
598 leaf temperature and transpiration of *Abies amabilis* branches. *Tree Physiol.* 19, 435-43.

599 Maxwell, K., Johnson, G., 2000. Chlorophyll fluorescence - A practical guide. *J. Exp. Bot.* 51,
600 659-68.

601 Mclaughlin, B.C., Zavaleta, E.S., 2012. Predicting species responses to climate change:
602 Demography and climate microrefugia in California Valley oak (*Quercus lobata*). *Glob.*
603 *Change Biol.* 18, 2301-12.

604 Melillo, J.M., Steudler, P.A., Aber, J.D., Newkirk, K., Lux, H., Bowles, F.P., Catricala, C.,
605 Magill, A., Ahrens, T., Morrisseau, S., 2002. Soil warming and carbon-cycle feedbacks to
606 the climate system. *Science* 298, 2173-76.

607 Montes-Hernández, B., López-Barrera, F., 2013. Seedling establishment of *Quercus insignis*: A
608 critically endangered oak tree species in southern Mexico. *Forest Ecol. Manag.* 310, 927-
609 34.

610 Parlange, M.B., Cahill, A.T., Nielsen, D.R., Hopmans, J.W., Wendroth, O., 1998. Review of heat
611 and water movement in field soils. *Soil Till. Res.* 47, 5-10.

612 Pérez-Ramos, I.M., Rodríguez-Calcerrada, J., Ourcival, J.M., Rambal, S., 2013. *Quercus ilex*
613 recruitment in a drier world: A multi-stage demographic approach. *Perspect. Plant Ecol.*
614 *Evol. Syst.* 15, 106-17.

615 Piper, F.I., Fajardo, A., Cavieres, L.A., 2013. Simulated warming does not impair seedling
616 survival and growth of *Nothofagus pumilio* in the southern Andes. *Perspect. Plant Ecol.*
617 *Evol. Syst.* 15, 97-105.

618 Poorter, H., Niklas, K.J., Reich, P.B., Oleksyn, J., Poot, P., Mommer, L., 2012. Biomass
619 allocation to leaves, stems and roots: Meta-analyses of interspecific variation and
620 environmental control. *New Phytol.* 193, 30-50.

621 Rennenberg, H., Dannenmann, M., Gessler, A., Kreuzwieser, J., Simon, J., Papen, H., 2009.
622 Nitrogen balance in forest soils: Nutritional limitation of plants under climate change
623 stresses. *Plant Biol.* 11, 4-23.

624 Rey-Benayas, J.M., Navarro, J., Espigares, T., Nicolau, J.M., Zavala, M.A., 2005. Effects of
625 artificial shading and weed mowing in reforestation of Mediterranean abandoned cropland
626 with contrasting *Quercus* species. *Forest Ecol. Manag.* 212, 302-14.

627 Roa-Fuentes, L.L., Templer, P.H., Campo, J., 2015. Effects of precipitation regime and soil
628 nitrogen on leaf traits in seasonally dry tropical forests of the Yucatan Peninsula, Mexico.
629 *Oecologia* 179, 585-97.

630 Rodriguez-Dominguez, C.M., Buckley, T.N., Egea, G., de Cires, A., Hernandez-Santana, V.,
631 Martorell, S., Diaz-Espejo, A., 2016. Most stomatal closure in woody species under
632 moderate drought can be explained by stomatal responses to leaf turgor. *Plant Cell Environ.*
633 39, 2014-26.

634 Smallwood, P.D., Steele, M.A., Faeth, S.H., 2001. The ultimate basis of the caching preferences
635 of rodents, and the oak-dispersal syndrome: Tannins, insects, and seed germination. *Am.*
636 *Zool.* 41, 840-51.

637 Valladares, F., Sánchez-Gómez, D., 2006. Ecophysiological traits associated with drought in
638 Mediterranean tree seedlings: Individual responses versus interspecific trends in eleven
639 species. *Plant Biol.* 8, 688-97.

640 Vargas-Márquez, F., 1997. *Parques Nacionales de México*. Secretaría de Medio Ambiente,
641 Recursos Naturales y Pesca, Mexico City.

642 Will, R.E., Wilson, S.M., Zou, C.B., Hennessey, T.C., 2013. Increased vapor pressure deficit due
643 to higher temperature leads to greater transpiration and faster mortality during drought for
644 tree seedlings common to the forest-grassland ecotone. *New Phytol.* 200, 366-74.

645 Xu, Z.Z., Zhou, G.S., 2006. Combined effects of water stress and high temperature on
646 photosynthesis, nitrogen metabolism and lipid peroxidation of a perennial grass *Leymus*
647 *chinensis*. *Planta* 224, 1080-90.

648 Xu, Z., Hu, T., Zhang, Y., 2012. Effects of experimental warming on phenology, growth and gas
649 exchange of treeline birch (*Betula utilis*) saplings, eastern Tibetan Plateau, China. *Eur. J.*
650 *For. Res.* 131, 811-19.

- 651 Yahdjian, L., Sala, O.E., 2002. A rainout shelter design for intercepting different amounts of
652 rainfall. *Oecologia* 133, 95-101.
- 653 Zar, J.H., 2010. *Biostatistical Analysis*, fifth ed. Prentice Hall, New Jersey.
- 654 Zavala-Chávez, F., 2008. Efecto del almacenamiento sobre la viabilidad y la germinación de
655 bellotas de *Quercus rugosa* y *Quercus glabrescens*. *Rev. Mex. Cs. For.* 33, 15-25.
- 656 Zavala-Chávez, F., García-Moya, E., 1996. *Frutos y semillas de encinos*. Universidad Autónoma
657 Chapingo, Texcoco.
- 658 Zavala-Chávez, F., García-Sánchez, F., 1999. Aspectos fisonómicos de los encinares de la Sierra
659 de Álvarez, San Luis Potosí, México. *Rev. Chapingo Ser. Cs. For. Amb.* 51, 27-35.