

This is the Post-print version of the following article: *Esáú Bojórquez-Velázquez, Aida Jimena Velarde-Salcedo, Antonio De León-Rodríguez, Hugo Jimenez-Islas, Jose Luis Pérez-Torres, Alfredo Herrera-Estrella, Eduardo Espitia-Rangel, Ana Paulina Barba de la Rosa, Morphological, proximal composition, and bioactive compounds characterization of wild and cultivated amaranth (Amaranthus spp.) species, Journal of Cereal Science, Volume 83, 2018, Pages 222-228*, which has been published in final form at: <https://doi.org/10.1016/j.jcs.2018.09.004>

© 2018. This manuscript version is made available under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) license <http://creativecommons.org/licenses/by-nc-nd/4.0/>

# Accepted Manuscript

Morphological, proximal composition, and bioactive compounds characterization of wild and cultivated amaranth (*Amaranthus* spp.) species

Esaú Bojórquez-Velázquez, Aida Jimena Velarde-Salcedo, Antonio De León-Rodríguez, Hugo Jimenez-Islas, Jose Luis Pérez-Torres, Alfredo Herrera-Estrella, Eduardo Espitia-Rangel, Ana Paulina Barba de la Rosa

PII: S0733-5210(18)30399-0

DOI: [10.1016/j.jcs.2018.09.004](https://doi.org/10.1016/j.jcs.2018.09.004)

Reference: YJCRS 2635

To appear in: *Journal of Cereal Science*

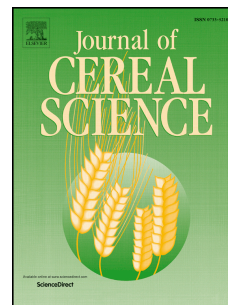
Received Date: 18 May 2018

Revised Date: 4 September 2018

Accepted Date: 6 September 2018

Please cite this article as: Bojórquez-Velázquez, Esaú., Velarde-Salcedo, A.J., De León-Rodríguez, A., Jimenez-Islas, H., Pérez-Torres, J.L., Herrera-Estrella, A., Espitia-Rangel, E., Barba de la Rosa, A.P., Morphological, proximal composition, and bioactive compounds characterization of wild and cultivated amaranth (*Amaranthus* spp.) species, *Journal of Cereal Science* (2018), doi: 10.1016/j.jcs.2018.09.004.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



1 **Morphological, proximal composition, and bioactive compounds**  
2 **characterization of wild and cultivated amaranth (*Amaranthus***  
3 **spp.) species**

4  
5 Esaú Bojórquez-Velázquez<sup>a</sup>, Aida Jimena Velarde-Salcedo<sup>a</sup>, Antonio De León-  
6 Rodríguez<sup>a</sup>, Hugo Jimenez-Islas<sup>b</sup>, Jose Luis Pérez-Torres<sup>a</sup>, Alfredo Herrera-  
7 Estrella<sup>c</sup>, Eduardo Espitia-Rangel<sup>d,\*</sup>, Ana Paulina Barba de la Rosa<sup>a,\*</sup>

8

9 <sup>a</sup>*IPICYT, Instituto Potosino de Investigación Científica y Tecnológica A.C.*  
10 *Camino a la Presa San José 2055, Col. Lomas 4<sup>a</sup> Sección, C.P. 78216, San Luis*  
11 *Potosí, Mexico.*

12 <sup>b</sup>*Tecnológico Nacional de México. Instituto Tecnológico de Celaya.*  
13 *Departamento de Ingeniería Bioquímica. Antonio García Cubas Pte. 600 esq AV.*  
14 *Tecnológico, Celaya, Guanajuato, 38010, México.*

15 <sup>c</sup>*Laboratorio Nacional de Genómica para la Biodiversidad, CINVESTAV-Irapuato,*  
16 *Irapuato, Irapuato, Guanajuato, 36824, México.*

17 <sup>d</sup>*INIFAP, Instituto Nacional de Investigaciones Forestales Agrícolas y Pecuarias.*  
18 *Campo Experimental Valle de México, Km 13.5 Carr Los Reyes-Texcoco, C.P.*  
19 *56250 Texcoco Estado de México.*

20

21

22 **Running title:** Characterization of wild and cultivated amaranth seeds

23

24

25 Corresponding authors:

26 E-mail: apbarba@ipicyt.edu.mx (A.P. Barba de la Rosa);

27 espitia.eduardo@inifap.gob.mx (E. Espitia-Rangel)

28

29

30

31 **ABSTRACT**

32 Amaranth seeds have gained renewed interest due to the presence of encrypted  
33 peptides with several biological functions, among which the inhibition of  
34 dipeptidyl peptidase IV (DPPIV) and angiotensin converting enzyme (ACE)  
35 stands out. Amaranth seeds also contain an oily fraction rich in squalene, an  
36 unsaturated hydrocarbon, which have been attributed diverse beneficial health  
37 effects. Up to date, only cultivated species have been analyzed and no  
38 information exists about bioactive peptides and lipid composition of wild  
39 amaranths. The aim of this work was to carry out the morphological and  
40 biochemical comparison of cultivated species *A. hypochondriacus* (waxy and  
41 non-waxy cultivars) and *A. cruentus* with wild species *A. powellii* and *A. hybridus*.  
42 The highest protein and fat contents were observed in *A. powellii*, but *A. cruentus*  
43 showed the highest squalene content. The electrophoretic protein profile showed  
44 differences in protein accumulation among species. *In vitro* gastrointestinal  
45 digestion was used to release the encrypted peptides and their inhibitory action  
46 against DPPIV and ACE activities was evaluated. Our results showed that  
47 peptides from *A. hypochondriacus* cv Opaca presented the highest inhibition  
48 against both DPPIV and ACE activities. This information is valuable in order to  
49 design strategies to obtain new amaranth varieties with higher nutraceutical  
50 quality.

51

52

53 **Keywords:** ACE; biopeptides; DPPIV; cultivated and wild species; squalene

## 54 **1. Introduction**

55 The world's human population is predicted to reach over 9.3 billion by the year  
56 2050, which will increase the demand of food. At the same time, climate change  
57 and soil deterioration represents an increasingly serious threat for food  
58 production (Lobell et al., 2011). To face these challenges, crop improvements  
59 must intend not only to ensure food availability, but crops must meet higher  
60 nutritious food standards (McCouch et al., 2013; Muñoz et al., 2017). For this  
61 reason, a sustainable diet supply, that means not only foods with high nutritional  
62 value but also foods containing bioactive compounds, which could represent  
63 additional benefits for health, are increasingly important for present and future  
64 generations (Siró et al., 2008).

65 With an increasing interest in new food sources, the need to better explore  
66 untapped plant biodiversity has gained importance, including wild relatives of our  
67 current crops (Muñoz et al., 2017). Wild species have survived several climate  
68 changes and therefore, they are valuable resources of genetic information for  
69 improving crops in the face of climate change (Lobell et al., 2011).

70 Because populations of many wild relatives are under threat from habitat loss  
71 and degradation, there is an urgent need to collect and preserve their  
72 germplasm. Such wild relatives need to be characterized to take advantage of  
73 them in the development of strategies to solve current and future agricultural  
74 challenges (McCouch et al., 2013). Another essential component to support the  
75 rediscovery of ancient species for food diversity is the marketing of these crops  
76 and their products (McCouch et al., 2013; Muñoz et al., 2017), for example, the

77 gluten-free food market worth almost \$1.6 billion in 2011. This resurgent interest  
78 is expressed in re-discovering ancestral crops as functional foods, which may  
79 offer an important alternative for people affected by celiac disease (Cooper,  
80 2015).

81 Amaranth is one of the oldest cultivated plants, which had great importance for  
82 the Aztec, Mayan, and Incas. Amaranth was grown as staple crop together with  
83 corn, but was banned during the Spanish Conquest. Since the 70's amaranth  
84 resurged as an alternative crop not only due to its high nutritional value (high  
85 lysine and methionine content) but also because amaranth prolamins content is  
86 negligible (Huerta-Ocampo and Barba de la Rosa, 2011), which are the seed  
87 storage proteins responsible for the manifestation of celiac disease and  
88 cerebropathias. In this new century amaranth gained renewed importance due to  
89 its nutraceutical properties; amaranth proteins contain encrypted peptides  
90 amongst the most studied are those with antihypertensive action (Huerta-  
91 Ocampo and Barba de la Rosa, 2011). Furthermore, the inhibitory peptides in  
92 amaranth seed proteins against dipeptidyl peptidase IV (DPPIV) activity have  
93 been identified and characterized (Velarde-Salcedo et al., 2013). The oily fraction  
94 of amaranth seeds is rich in squalene, an unsaturated hydrocarbon to which has  
95 been attributed hypocholesterolaemic properties (Chaturvedi et al., 1993).

96 In addition to nutritional characteristics, amaranth plants have attractive  
97 agronomic features; they grow where cereals and vegetables cannot such as dry  
98 soils, high altitudes, and high temperatures (Huerta-Ocampo and Barba de la  
99 Rosa et al., 2011). Amaranth cultivation has increased and breeders produced a

100 large number of new varieties adapted to different environments, however, some  
101 of these new varieties are only new names for old varieties or landraces, hence  
102 the use of wild amaranth species with remarkable tolerance to several abiotic  
103 stresses such as *A. powellii* and *A. hybridus* are of great interest. Although these  
104 wild species are proposed as the ancestors of the main cultivated species used  
105 for seed production such as *A. hypochondriacus* and *A. cruentus*, still its  
106 molecular relationships have not been established. The aim of the present work  
107 was to compare the morphological characteristics and bioactive compounds  
108 content of cultivated and wild amaranth species.

109

## 110 **2. Materials and Methods**

### 111 *2.1. Amaranth genotypes*

112 Amaranth seeds of wild (*A. hybridus* and *A. powellii*) species as well as the  
113 most cultivated and studied species, *A. hypochondriacus* cv Nutrisol, and *A.*  
114 *cruentus* cv Amaranteca, were provided by the National Institute for Forestry,  
115 Agriculture and Livestock Research (INIFAP), Mexico. Two more cultivars of *A.*  
116 *hypochondriacus* were included in the study; Cristalina (non-waxy type) and  
117 Opaca (waxy type), which are derived from a heterozygous plant for this  
118 character by six generations of single seed descendent and were collected from  
119 Atzitzintla, Tlaxcala, Mexico.

120

### 121 *2.2. Morphological and structural seeds characterization*

122 Seed weight was calculated by weighing 100 seeds on an electronic balance  
123 DV215CD Discovery (Ohaus, Parsippany, NJ, USA) with 0.01/0.1 mg accuracy.  
124 The weight of 100 seeds was extrapolated to 1000 seeds. Seed dimensions  
125 (diameter and width) were taken with the SteREO Discovery V8 (Carl-Zeiss,  
126 Oberkochen, GE). All measurements were done in triplicates.

127 Images of whole seeds were obtained with the SteREO Discovery V8 (Carl-  
128 Zeiss). Paradermal sections were visualized by scanning electron microscopy  
129 captured with an ESEM model Quanta 200 (FEI, Hillsboro, OR, USA) from the  
130 National Laboratory of Nanosciences and Nanotechnology Research (LINAN)  
131 IPICYT.

132

### 133 *2.3. Amaranth flours proximate composition*

134 Seeds were cleaned and milled in liquid nitrogen using a KRUPS GX4100  
135 (Solingen, GE) mill to obtain fine flour. Flour samples were stored in plastic tubes  
136 at -80 °C until analysis. Total nitrogen content was determined by micro-Kjeldahl  
137 method (AOAC, 2007, method 12.960.52), and total protein content was  
138 calculated using a 5.85 factor. Fat content was determined by Soxhlet method  
139 (AOAC, 2007, 996.01 method). Crude fibre and ash contents were obtained  
140 according to AOAC (2007) methods 991.43 and 900.02, respectively. All  
141 determinations were made at least in triplicates.

142

### 143 *2.4. Protein extraction and electrophoretic profile of amaranth seeds proteins*



144 Total protein extracts, from the six amaranth species studied, were obtained  
145 by mixing 0.1 g flours with 2 ml of a solution containing 7 M urea, 2 M thiourea,  
146 2% (w/v) Triton X-100 and 0.05 M DTT. Suspensions were mixed by vortexing for  
147 15 min at 4 °C and centrifuged at 17,000×g at 20 °C, supernatants were  
148 recovered and protein quantified using the Bradford protein assay (Bio-Rad,  
149 Hercules, CA, USA). Each sample was analysed by denaturing polyacrylamide  
150 gel electrophoresis (SDS-PAGE) in a discontinuous Tris-glycine system. The  
151 stacking and resolving gels were 4% and 13.5%, respectively. Protein (15 µg)  
152 was loaded onto the gel and separated in a Mini-Protean III system (Bio-Rad),  
153 gel was run at 10 mA/gel for 30 min followed by 25 mA/gel until bromophenol  
154 blue reached the bottom of the gel. After electrophoresis, the gels were stained  
155 with a 0.05% Coomassie blue R-250 (USB Corporation, Cleveland, OH, USA) in  
156 40% methanolic solution containing 10% acetic acid.

157

#### 158 *2.5. In-gel digestion and LC-MS/MS protein identification*

159 Protein bands were manually excised from gel, destained, reduced with 10  
160 mM dithiothreitol and alkylated with 55 mM iodoacetamide. Protein digestion  
161 was carried out with sequencing-grade trypsin (Promega, Madison, WI, U.S.A.).  
162 Tryptic peptides were analyzed with a nanoACQUITY UPLC System (Waters,  
163 Milford, MA, U.S.A.) coupled to a SYNAPT-HDMS Q-TOF (Waters) mass  
164 spectrometer. MS/MS spectra data sets were used to generate PKL files using  
165 Protein Lynx Global Server v2.4 (PLGS, Waters). Proteins were then identified  
166 using the MASCOT search engine v2.5 (Matrix Science, London, U.K.).

167 Searches were conducted against the *Viridiplantae* subset of the NCBI nr protein  
168 database (6 686 534 sequences, May 2018). Trypsin was used as the specific  
169 protease, and one missed cleavage was allowed. The mass tolerance for  
170 precursor and fragment ions was set to 50 ppm and 0.1 Da, respectively.  
171 Carbamidomethyl cysteine was set as fixed modification and oxidation of  
172 methionine was specified as variable modification. The protein identification  
173 criteria included at least two MS/MS spectra matched at 99% level of confidence,  
174 and identifications were considered successful when significant MASCOT scores  
175 >50 were obtained, indicating the identity or extensive homology at  $p < 0.01$  and  
176 the presence of a consecutive y ion series of more than three amino acids.

177

## 178 2.6. *Gastrointestinal digestion in vitro*

179 A simulated gastrointestinal digestion *in vitro* model was carried out as  
180 reported before (Velarde-Salcedo et al., 2013). Briefly, 1 g of amaranth defatted  
181 flour was resuspended in 20 ml of 0.03 M NaCl pH 2.0. In order to inactivate  
182 proteases, the suspensions were heated in a water bath at 80 °C for 5 min and  
183 allowed to cool down at room temperature. Porcine pepsin (Sigma–Aldrich, St.  
184 Louis, MI, USA) previously dissolved in 0.03 M NaCl pH 2.0 was added in a 1:20  
185 ratio (w/w enzyme to substrate). Samples were digested at constant pH for 3 h at  
186 37 °C and pH was then adjusted to 7.5. A mixture of trypsin (Sigma–Aldrich) and  
187 pancreatin (Sigma–Aldrich) was prepared (1:1 w/w trypsin:pancreatin ratio in 0.1  
188 N NaHCO<sub>3</sub>), added to the digestive solution and incubated at constant pH for an  
189 additional 3 h period (1:20 w/w enzyme to substrate ratio for both the enzymes,

190 trypsin and pancreatin). Digestion was stopped by heating the suspensions at 75  
191 °C for 20 min and centrifuged at 13,000xg for 30 min. Peptides were ultra filtrated  
192 through 10 kDa filters (Amicon Ultra-10 centrifugal filters, Sigma-Aldrich). The  
193 peptides concentration was determined by the Lowry-based DC Protein Assay  
194 (Bio-Rad) using BSA as a standard, and then stored at -20 °C until analysis.

195

#### 196 *2.6.1. Inhibition of dipeptidyl peptidase IV (DPPIV) activity*

197 DPPIV activity was measured using the chromogenic substrate Gly-Pro-  
198 pNitroanilide (Sigma-Aldrich) as previously reported (Velarde-Salcedo, et al.,  
199 2013). Briefly, 10 µl of 100 ng/ml of dipeptidyl peptidase IV (Sigma-Aldrich) were  
200 added to 40 µl amaranth peptides dissolved in 100 mM Tris pH 8 and 50 µl of 1  
201 mM Gly-Pro-pNitroanilide dissolved in Tris buffer. Mix was incubated at 37 °C for  
202 1 h. Absorbance was measured at 415 nm in a Multiskan Go plate reader  
203 (Thermo Fisher Scientific Inc., Waltham, Massachusetts, USA). Results were  
204 expressed as µmol of nitroaniline/min based on a p-nitroaniline (Sigma-Aldrich)  
205 standard curve.

206

#### 207 *2.6.2. Inhibition of Angiotensin Converting Enzyme (ACE) activity*

208 Peptides with inhibitory activity against ACE were measured by  
209 spectrophotometric assay. Briefly 20 µl of sample was added to 0.1 ml of 0.1 M  
210 potassium phosphate buffer (pH 8.3) containing 0.3 M NaCl and 5 mM hippuryl-  
211 histidyl-leucine (HHL, Sigma). ACE (5 mU) (EC 3.4.15.1, 5.1 U/mg, Sigma) was  
212 added and the reaction mixture was incubated at 37 °C for 30 min. The reaction

213 was terminated by the addition of 0.1 ml of 1 M HCl. The hippuric acid formed  
214 was extracted with ethyl acetate, heat-evaporated at 95 °C for 10 min, dissolved  
215 in distilled water and measured spectrophotometrically at 228 nm. The activity of  
216 each sample was tested in triplicate. Captopril was used as a positive control.  
217 The IC<sub>50</sub> value was defined as the peptide concentration (mg/ml) needed to  
218 inhibit 50% ACE activity; it was calculated by an ACE inhibition (%) vs. log  
219 peptide concentration (mg/ml) linear regression.

220

### 221 *2.7. Amaranth lipid extraction, GC-MS analysis, and squalene quantification*

222 Lipids from amaranth seeds were extracted by mixing 0.1 g of amaranth flour  
223 with 1.75 ml of hexane, in constant agitation at room temperature by 5 h.  
224 Samples were centrifuged (17,000×g, 20 min, 25 °C) and supernatants  
225 transferred into new tubes. Extracts were analysed by GC-MS using a  
226 7820A/5977E System (Agilent Technologies, Santa Clara, California, USA), with  
227 a HP-5ms capillary column (Agilent Technologies) of 30 m length, 250 µm of  
228 inner diameter and 0.25 µm-film thickness. Samples, 3 µl, were injected in  
229 splitless mode. The column was held at 80 °C for 1 min after injection, the  
230 temperature programmed at 20 °C/min to 210 °C and held for 10 min more, then  
231 10 °C/min to 280 °C and held for 35 min. Helium was used as carrier gas, at a  
232 constant column flow rate of 1 ml/min. The injector temperature was 250 °C and  
233 the detector temperature was 230 °C. The mass spectrometer was operated  
234 under Electron Impact Ionization at 70 eV with a mass range from 30-500 amu.  
235 Lipids were identified comparing their retention times and the mass spectra

236 against NIST Mass Spectral Library v2.2. Results were expressed as the  
237 individual relative percentage of each lipid present in the sample. For squalene  
238 absolute quantification, an analytical standard (Sigma) was used to construct a  
239 calibration curve from 1-10 mg/l ( $r^2=0.997$ ).

240

## 241 2.8. Statistical Analysis

242 A variance analysis (ANOVA) was carried out with Holm-Sidak test using the  
243 Sigma Plot software analysis v12.3 (Systat Software, Inc., San Jose, CA, USA)  
244 for paired analysis and considering  $p<0.05$  for statistically significant differences.

245

## 246 3. Results

### 247 3.1. Morphological characterization

248 *A. powellii* showed the lowest value for thousand seeds weight (TSW) with  
249 only 0.45 g and *A. hypochondriacus* cv Cristalina presented the highest TWS of  
250 0.90 g (**Supplementary Table S1**). *A. hypochondriacus* cv Nutrisol have the  
251 smallest seed length 1.19 mm and the smallest width was observed in *A. powellii*  
252 (0.88 mm). The largest seeds were those of *A. hybridus* and *A. hypochondriacus*  
253 cv Cristalina with dimensions of 1.32x1.15 mm and 1.33x1.13 mm, respectively.  
254 Although different in length and width, most species conserve the same  
255 diameter/width ratio, with exception of *A. hypochondriacus* cv Cristalina, which  
256 present more oval seeds (1.17 D/W ratio) and *A. cruentus* with the most rounded  
257 seeds (1.11 D/W ratio).

258 Phenotypic characteristics of wild and cultivated seeds are shown in **Figure**  
259 **1**. The wild species are bright black in colour, while seeds of cultivated species  
260 are cream light colour. Different *A. hypochondriacus* cultivars are distinguished  
261 due to the translucent (vitreous) or opaque characteristics; the cv Nutrisol is the  
262 most opaque while cv Cristalina, as its name indicates, is a translucent bright  
263 seed. Vitreous characteristic has been related with the type, degree of cross-  
264 linking, and molecular weight distribution of proteins in seeds.

265 SEM analyses of paradermal cuts showed that wild species *A. hybridus* and  
266 *A. powellii* as well as *A. hypochondriacus* cv Cristalina contain polyhedral defined  
267 structures in the vitreous perisperm, structures that are not observed in *A.*  
268 *cruentus*, *A. hypochondriacus* cvs Nutrisol and Opaca, which perisperm is not  
269 vitreous (**Figure 2**).

270

### 271 3.2. Proximal composition of amaranth seeds flours

272 The flours proximal composition from wild and cultivated amaranth seeds is  
273 shown in **Table 1 and Supplementary Table S2**. Although *A. powellii* is the  
274 smallest seeds, is the species with the highest protein (17.8%) and fat (8.1%)  
275 contents. *A. cruentus* is the species with the lowest protein content (14.8%), but  
276 the highest starch content (73.0%). On the other hand, *A. hybridus* and *A.*  
277 *hypochondriacus* cv Cristalina, with the largest seeds, are the species with the  
278 lowest fat content (5.9 and 5.7%, respectively). Interestingly, *A. hybridus* has the  
279 highest crude fibre (6.1%) and ash (3.7%) contents. It is interesting that among  
280 the *A. hypochondriacus* species, the most commercial cultivar, Nutrisol, showed

281 less protein content (15.8%) in comparison with cvs Opaca and Cristalina  
282 (16.7%).

283

### 284 3.3. Electrophoretic pattern and protein identification

285 The amount of protein quantified by Bradford (**Supplementary Figure S1**)  
286 correlated with the values obtained by Kjeldahl method (**Table 1**), which indicates  
287 that total protein extracted was almost the total protein present in seeds. Total  
288 proteins were analysed by SDS-PAGE (**Figure 3**). In all species and cultivars  
289 were observed the bands located at 35-37 kDa and 18-20 kDa, which represent  
290 the acidic and basic subunits of the canonical 11S globulins. The most  
291 remarkable differences among species and cultivars analysed were observed in  
292 the range of 50 to 70 kDa. Both wild species as well as *A. hypochondriacus* cv  
293 Cristalina have a band around 65 kDa. *A. powellii* and *A. cruentus* share a band  
294 of 60 kDa. *A. hybridus* and all *A. hypochondriacus* cultivars showed a 55 kDa  
295 band. These three bands were cut from gel, analysed by LC-MS/MS and  
296 identified as a Granule Bound Starch Synthase I or GBSSI (**Supplementary**  
297 **Table S3 and Supplementary Figure S2**).

298

### 299 3.4. Amaranth peptides with inhibitory activity against DPPIV and ACE

300 A simulated gastrointestinal digestion *in vitro* method was used to release the  
301 encrypted peptides from all amaranth samples. The capacity of released  
302 amaranth peptides to inhibit both DPPIV and ACE enzymes was measured.  
303 DPPIV inhibition increased in a dose-response relationship (**Figure 4A**), higher

304 inhibition activity was detected at the highest tested concentration (3.2 mg/ml). At  
305 this concentration, *A. hypochondriacus* cv Opaca rendered the highest inhibitory  
306 activity reaching a 60% of DPPIV with an IC<sub>50</sub> of 1.6 mg/ml. *A. hypochondriacus*  
307 cv Cristalina and *A. powellii* showed the least DPPIV inhibition reaching only 40%  
308 at the highest tested peptide concentration (3.2 mg/ml). A similar ACE inhibitory  
309 activity profile was observed (**Figure 4B**). *A. hypochondriacus* cv Opaca peptides  
310 presented the highest activity reaching of 80% inhibition at 3.2 mg/ml with an IC<sub>50</sub>  
311 of 0.6 mg/ml. *A. hypochondriacus* cv Nutrisol, *A. hybridus* and *A. cruentus*  
312 showed an IC<sub>50</sub> of 1.5 mg/ml while *A. hypochondriacus* cv Cristalina and *A.*  
313 *powellii* the IC<sub>50</sub> was of 2.5 mg/ml.

314

### 315 3.5. Characterization of lipids in amaranth seeds

316 The lipid composition analysed by GC-MS showed the presence of palmitic  
317 and linoleic acids in all species. Linoleic ethyl esters was present only in *A.*  
318 *hybridus* and *A. hypochondriacus* cvs Cristalina and Nutrisol, while oleic acid  
319 ethyl ester was only present in *A. hypochondriacus* cvs Cristalina and Nutrisol.  
320 Butyl ester of palmitic and stearic acid were detected in all samples analysed.  
321 Stigmasterol, an important phytosterol, was detected in higher abundance in the  
322 wild species *A. powellii*, followed by *A. hypochondriacus* cvs Cristalina, and *A.*  
323 *hybridus* (**Supplementary Table S4**). Squalene, an unsaturated hydrocarbon,  
324 was detected in all samples but interestingly the highest abundance was  
325 detected in *A. cruentus* (**Supplementary Table S4 and Supplementary Figure**  
326 **S3**). Because the importance of squalene and relative abundance do not reflect



327 the real quantity present in samples, squalene was quantified. Results showed  
328 that squalene concentration ranged from 0.197 to 0.335 g/100 g of seeds and  
329 this values in relation to oil content ranged from 2.85 to 4.86 g/100 g oil (**Table**  
330 **2**).

331

#### 332 **4. Discussion**

333 Wild ancestors of common cereals, such as rice and wheat, have been used  
334 as resources for quality improvement of cultivated grains (Cooper, 2015).  
335 However, despite the potential of a several seeds progenitors to face the  
336 challenges of modern agriculture, there are few collections of wild relatives and  
337 even the available wild genetic resources are still under-utilized (McCouch et al.,  
338 2013).

339 Mexico is rich in genetic diversity of amaranth species such as *A. powellii* and  
340 *A. hybridus* (Espitia-Rangel et al., 2012). These wild accessions have been  
341 considered as the ancestors of the cultivated species *A. hypochondriacus* and *A.*  
342 *cruentus* (Sauer et al., 1967), but concerns have been raised about the  
343 hypothesis of amaranths ancestors. Hence, that morphological and molecular  
344 analysis of wild and cultivated species could help validate the amaranth  
345 phylogeny and evolutionary relationships (Espitia-Rangel et al., 2012).

346 For years, plant wild species have survived to abiotic and biotic stresses.  
347 Seeds have used dark or bright colours, as a signal of toxic materials, as  
348 protective action against predators (Lev-Yadun, 2016). These pigmentations are  
349 due to polyphenols, plant metabolites that play a role in the protection of plants

350 against ultraviolet radiation, pathogens, and herbivores (Alvarez-Jubete et al.,  
351 2010). The absence of these pigments in cultivated amaranths is considered as a  
352 trait of domestication.

353 However, there are black seeds such as *Pisum humile* and *P. fulvum*, which  
354 are highly edible but mimic various toxic seeds of legumes that grow in the same  
355 region such as *Lathyrus ochrus* (Lev-Yadun, 2016). Wild black seeded  
356 amaranths (a hybrid between *A. hypochondriacus* and *A. hybridus*), are grown in  
357 Michoacán-Mexico to make special black tamales (Sauer et al., 1967). Another  
358 characteristic of wild seeds is the hardness of their testa as protective tissues for  
359 mechanical defences against granivores attacks. However, a light color seed with  
360 soft testa has been the target for domestication. This also can be observed in  
361 amaranth, light seeds with soft testa were selected for cultivation and  
362 domestication (**Figure 1**).

363 Seed size is another characteristic related to the profitability of agricultural  
364 operations. Selection of big seeds, in terms of genetic changes, is related to  
365 breakdown of seed dispersal and seed dormancy (Fernández-Marín, 2014). The  
366 reported size for amaranth seeds is 0.9-1.7 mm diameter and TSW ranged from  
367 0.6-1.0 g (Assad et al., 2017), values that agree with our results, except for *A.*  
368 *powellii* that have the smallest diameter and TSW of 0.88 mm 0.45 g,  
369 respectively. Interestingly, the cultivated species *A. hypochondriacus* cv  
370 Cristalina, *A. cruentus*, and the wild *A. hybridus* bear the largest seeds.  
371 Genotypes with small seeds are correlated with low seed quality, since larger  
372 seed size is probably advantageous because of their better standability under

373 agricultural conditions, and because of the greater plantlets size arising from  
374 them, and particularly important for crops with edible seeds (Lush and Wien,  
375 1980; Espitia-Rangel et al., 2012). The weight has been also widely used as  
376 characteristic for seeds improvement and increased yield. Our results showed  
377 that the cultivated species *A. hypochondriacus* cvs Cristalina and Opaca, as well  
378 *A. cruentus* are the cultivars that presented the highest values of TSW.

379 Although several morphological characteristics have been studied in relation to  
380 seeds breeding, less attention has been paid and it is poorly understood how  
381 agricultural selection and cultivation affected the nutritional quality of seeds,  
382 which are important traits for crop improvement and meet with Food Sovereignty  
383 (Muñoz et al., 2017).

384 The scarce literature available on cultivars and their wild relatives are focused  
385 on protein and amino acid contents and very little information exists on other  
386 important nutritional traits such oils or starch. In general, a decrease in protein,  
387 fibre, and minerals is related with the increase in carbohydrates. Seeds with  
388 higher carbohydrate content are bigger and with higher TSW. This is in  
389 agreement with our results, *A. powellii*, wild species with the smallest size and  
390 TSW presented the highest protein and lowest carbohydrate contents (17.8%  
391 and 64.5%, respectively). Among the cultivated species, which are bigger seeds,  
392 *A. cruentus* showed the lowest protein but highest carbohydrate content (14.8  
393 and 73.9%, respectively).

394 Amaranth lipid content varies from 5.7% to 8.1% (Table 1), values that are in  
395 the range of reported values from 6 to 20% (Assad et al., 2017). In soybean, oil

396 content is higher in domesticated seeds as compared with its wild counterpart  
397 (Zong et al., 2017), however in amaranth this relationship is not clear. The wild *A.*  
398 *powellii*, is the species with the highest fat content but *A. hybridus*, also a wild  
399 species, showed the lowest fat content. Our results are in agreement with  
400 Fernández-Marín et al. (2014) who reported that in some legumes such as  
401 soybean, peanut, lens, among others, the domestication caused a decreased in  
402 total carotenoid content, especially a reduction of  $\alpha$ - and  $\gamma$ -tocopherol was  
403 detected as domestication increased. In amaranth was observed a reduction in  
404 abundance of stigmasterol from 3.17% presented in the wild species *A. powellii*,  
405 while in the most domesticated species, *A. hypochondriacus* cv Nutrisol, the  
406 value was 1.44% (**Supplementary Table S4**).

407 Amaranth grain is considered as a good source of crude fibre, content that is  
408 higher than in rice, sorghum, oat, barley, and potato (USDA,  
409 [ndb.nal.usda.gov/ndb/foods/list](http://ndb.nal.usda.gov/ndb/foods/list)). *A. hybridus* showed the highest crude fibre  
410 content (6.1%) and the lowest content was found in *A. cruentus* and *A.*  
411 *hypochondriacus* cv Nutrisol, species with the highest starch content. Fibre is an  
412 important part of human nutrition; sufficient fibre intake is related with prevention  
413 of colon cancer. It has been reported that fibre in amaranth could be responsible  
414 for the control of blood cholesterol level preventing the development of  
415 atherosclerosis and its complications (Caselato-Souza et al., 2014).

416 Carbohydrates in amaranth are of especial attention due to the very small size  
417 of the starch granules (0.5 - 2  $\mu$ m), which gives functional characteristics of great  
418 interest in food applications (Kong et al., 2009). Carbohydrates were very

419 important during amaranth domestication that is thought to have occurred during  
420 the prehispanic times. Aztecs used sticky grain amaranths to make cakes as part  
421 of religious ceremonies (Sauer, 1967). The sticky grain selection was the origin  
422 of so called waxy varieties of cereals and other starch-producing crops (Hunt et  
423 al., 2010). Amaranth waxy types were selected and nowadays are the cultivated  
424 species (*A. cruentus*, *A. hypochondriacus*). Sticky starch type is characterized by  
425 very low content of amylose, a character that modifies the glycaemic index. Waxy  
426 starch types generally have a higher glycaemic loads, its consumption is related  
427 with a better physical performance and quicker recovery associated with intense  
428 physical activity and with the reloading of glycogen storages after exercise  
429 (Wright, 2005). Therefore, higher-glycaemic cereal grains, which may have been  
430 an advantage and better tasting, treat in past cultures, today may impose a  
431 disadvantage for modern civilization where exercise and physical activity has  
432 decreased and glycaemic loads are related with type-2 diabetes risk (WHO,  
433 2016). So non-waxy varieties should be reconsidered for the generation of new  
434 amaranth cultivars and wild species are an important source of non-waxy  
435 starches and high protein contents.

436 The waxy and non-waxy amaranth types are related with the morphological  
437 observation, the wild species showed a well-defined polyhedral perisperm  
438 (**Figure 2**), but also with the differential accumulation of proteins (**Figure 3**). The  
439 differentially accumulated proteins were identified as GBSSI (**Supplementary**  
440 **Table S3 and Supplementary Figure S2**), enzyme responsible for amylose  
441 synthesis. It is important to mention that only the species that presented

442 polyhedral structures observed by SEM (**Figure 2**), have the GBSSI band at 65  
443 kDa (**Figure 3**). GBSSI isoforms of 60 and 55 kDa could be not functional leading  
444 to the synthesis of starches with different ratios of amylose/amylopectin, and  
445 therefore different rheological and physicochemical characteristics, which may be  
446 highly valued in the food and beverage science and technology industry.

447 In relation to nutraceutical characteristic, although limited data are available in  
448 this sense, it has been reported that ancient wheat are not really healthier than  
449 modern wheat (Shewry and He, 2015). In this work we have shown that one  
450 cultivated species, *A. hypochondriacus* cv Opaca showed the highest inhibition  
451 against the DPPIV and ACE activities.

452 Type-2 diabetes is a chronic metabolic disorder considered as one of the  
453 major global health problems (WHO, 2016). The actual therapies to lower the  
454 hyperglycaemic state in patients with diabetes are based on the inhibition of  
455 DPPIV, enzyme responsible for degradation and inactivation of glucose-  
456 dependent insulintropic polypeptide (GIP) and glucagon-like peptide-1 (GLP-1).  
457 GIP and GLP are incretin hormones, which have a function to induce insulin  
458 secretion in the  $\beta$ -pancreatic cells. Synthetic DPPIV inhibitors are used as drugs  
459 therapies, however, there is a risk of side effects, DPPIV has several other  
460 functions than incretins inhibition (Matteucci and Giampietro, 2011) and natural  
461 DPPIV inhibitors are of great interest as therapy to promote a healthy life (Siró et  
462 al., 2008). The possible mechanisms of action of amaranth peptides with  
463 inhibitory activity against DPPIV have been described (Velarde-Salcedo et al.,  
464 2013).

465 On the other hand, hypertension is one of the main risk factors for  
466 cardiovascular diseases, hypertension might affect as many as 1 billion  
467 individuals worldwide. ACE plays an important role in the regulation of blood  
468 pressure by catalysing the production of the vasoconstrictor Angiotensin II and  
469 inactivating the vasodilator Bradykinin. Thus, ACE-inhibitory drugs are commonly  
470 used to control high blood pressure in hypertensive subjects. It is reported that  
471 the main source of ACE inhibitory peptides is fermented milk with IC<sub>50</sub> values  
472 ranged from 0.47 to 1.70 mg/ml (Gonzalez-Gonzalez et al., 2011). Amaranth IC<sub>50</sub>  
473 value was 0.6 mg/ml, which is in the range of milk peptides. Peptides YP, LPP,  
474 LRP, VPP, and IKP peptides have been detected in amaranth seed protein  
475 hydrolyzates, the IKP peptide has been described as one of the most potent  
476 inhibitor of ACE activity (Huerta-Ocampo and Barba de la Rosa, 2011).

477 Interestingly, although cultivated amaranth species presented higher  
478 carbohydrates content, the amount of fat was no dramatically decreased (5.7 to  
479 8.1%). We found that the oil composition in amaranth was characterized for the  
480 high levels of squalene, reaching values from 2.85 g/100g oil in *A.*  
481 *hypochondriacus* cv Nutrisol and up to 4.86 g/100g oil in *A. cruentus* (**Table 2**).  
482 He and Corke (2003) reported values from 1.0% to 7.3% squalene/oil, depending  
483 on the amaranth cultivar/specie analysed, while D'Amico and Schoenlechner  
484 (2017) reported concentrations from 2.26 to 11.19%.

485 Amaranth has the ability to modulate cholesterol levels in serum, which is due  
486 to its content of squalene (Chaturvedi et al., 1993; D'Amico and Schoenlechner,  
487 2017). The recommended squalene intake (0.25 to 0.5 mg a day) may lower

488 blood cholesterol levels reducing the risk of atherosclerosis and heart attack  
489 (Reddy and Couvreur, 2009).

490 Palmitic, linoleic, and cis-octadecanoic acids were detected (**Supplementary**  
491 **Table S4**). The ethyl esters of linoleic and oleic acids as butyl esters of palmitic  
492 and stearic acids were detected as minor components. In this regard, it was  
493 reported that in carob (*Ceratonia siliqua* L.) seeds, the most abundant fatty acids  
494 were the methyl-esters of oleic acid (C18:1), linoleic acid (C18:2n6), palmitic acid  
495 (C16:0), and stearic acid (C18:0) (Gubbuk et al., 2018).

496 Several clinical studies have shown that a high trans-fatty acid diet causes  
497 adverse changes in the plasma lipoprotein profile, with an increase in LDL and a  
498 decrease in HDL (Siddhuraju and Becker, 2001). In the present study, no trans-  
499 fatty acids such as elaidic and linolelaidic, myristic, behenic, eruci and lignoceric  
500 acids were detected.

501

## 502 **Conclusions**

503 Based on these results, we propose *A. powellii* as an interesting option to  
504 generate amaranth cultivars with higher protein contents in their grains. *A.*  
505 *hybridus* showed the highest crude fibre content (6.1%), while *A. cruentus* and *A.*  
506 *hypochondriacus* cv Nutrisol had the highest starch content. Wild species and *A.*  
507 *hypochondriacus* cv Cristalina presented a perisperm with polyhedral well-  
508 defined structures and share the presence of a 65 kDa band corresponding to  
509 GBSSI; while *A. hypochondriacus* cvs Nutrisol and Opaca and *A. cruentus* cv  
510 Amaranteca showed a starch with low or no amylose content. The higher



511 inhibition of DPPIV activity was detected at the highest concentration of peptides  
512 (3.2 mg/ml). *A. hypochondriacus* cv Opaca rendered the highest activity reaching  
513 until 60% of DPPIV inhibition with an IC<sub>50</sub> of 1.6 mg/ml. Regarding ACE inhibitory  
514 activity, also *A. hypochondriacus* cv Opaca showed the highest activity reaching  
515 of 80% inhibition at 3.2 mg/ml with an IC<sub>50</sub> of 0.6 mg/ml. Lipids in amaranth  
516 varied among species and cultivars, squalene highest concentrations were  
517 detected in *A. cruentus* followed by *A. hybridus*.

518 Further efforts are needed in order to improve amaranth phenotyping with  
519 special focus on food quality and health-promoting compounds, hence wild  
520 species rediscovery will provide more information to support amaranth breeding.

521

## 522 **Acknowledgments**

523 EBV thanks to CONACYT fellowship 298096 and we thank to CONACYT-  
524 Problemas Nacionales “Amaranto en la Soberanía Alimentaria Grant No.  
525 248415”. JLPT thanks to support from CONACYT. Thanks to Dr. O.A. Patrón-  
526 Soberano for her technical assistance in microscopic techniques and Dr. V.E.  
527 Balderas-Hernández for his assistance with GC-MS.

528

529

## 530 **References**

531 Alvarez-Jubete, L., Wijngaard, H., Arendt, E.K., Gallagher, E. 2010. Polyphenol  
532 composition and in vitro antioxidant activity of amaranth, quinoa,  
533 buckwheat and wheat as affected by sprouting and baking. Food  
534 Chemistry 119, 770-778.  
535 AOAC, 2007. Official Methods of Analysis, 18<sup>th</sup> ed. Association of Official  
536 analytical Chemists, Washington, D.C.

- 537 Assad, R., Reshi, Z.A., Jan, S., Rashid, I., 2017. Biology of amaranths. The  
538 Botanical Review 83, 382-436.
- 539 Caselato-Sousa, V.M., Ozaki, M.R., de Almeida, E.A., Amaya-Farfan, J., 2014.  
540 Intake of heat- expanded amaranth grain reverses endothelial dysfunction  
541 in hypercholesterolemic rabbits. Food and Function 5, 3281.
- 542 Cooper, R., 2015. Re-discovering ancient wheats varieties as functional foods.  
543 Journal of Traditional and Complementary Medicine 5, 138-143.
- 544 Chaturvedi, A., Sarojini, G., Devi, N.L., 1993. Hypocholesterolemic effect of  
545 amaranth seeds (*Amaranthus esculantus*). Plant Foods for Human  
546 Nutrition 44, 63-70.
- 547 D'Amico, S., Schoenlechner, R., 2017. Amaranth: Its Unique Nutritional and  
548 Health-Promoting Attributes. In: Taylor, J.R.N., Awika, J.M. (Eds.), Gluten-  
549 Free Ancient Grains Cereals, Pseudocereals, and Legumes: Sustainable,  
550 Nutritious, and Health-Promoting Foods for the 21st Century. Woodhead  
551 Publishing, Elsevier Ltd., pp. 131-159.
- 552 Espitia-Rangel, E., Escobedo-Lopez, D., Mapes-Sánchez, C., de la O Olán, M.,  
553 Aguilar-Delgado, M., Hernández-Casillas, J.M., Ayala-Garay, A.V., Rivas-  
554 Valencia, P., Martínez-Trejo, G., Ramírez-Vazquez, M.L., Morán-  
555 Bañuelos, S.H., 2012. Conservation of amaranth (*Amaranthus* spp.)  
556 genetic resources in Mexico. In: Amaranto Ciencia y Tecnología. Eduardo  
557 Espitia Rangel (Ed). Libro Científico No. 2. INIFAP/SINAREFI. México. pp  
558 147-163.
- 559 Fernández-Marín, B., Milla, R., Martín-Robles, N., Arc, E., Kranner, I., Becerril,  
560 J.M., García-Plazaola, I., 2014. Side-effects of domestication: cultivated  
561 legume seeds contain similar tocopheroles and fatty acids but less  
562 carotenoids than their wild counterparts. BMC Plant Biology 14, 1599.
- 563 Gonzalez-Gonzalez, C.R., Tuohy, K.M., Jauregi, P., 2011. Production of  
564 angiotensin-I-converting enzyme (ACE) inhibitory activity in milk fermented  
565 with probiotic strains: Effect of calcium, pH and peptides on the ACE-  
566 inhibitory activity. International Dairy Journal 21, 615-622.
- 567 Gubbuk, H., Kafkas, E., Guven, E., Gunes, E., 2010. Physical and phytochemical  
568 profile of wild and domesticated carbo (*Ceratonia siliqua* L.) genotypes.  
569 Spanish Journal of Agricultural Research 8, 1129-1136.
- 570 He, H.-P., Corke, H., 2003. Oil and squalene in *Amaranthus* grain and leaf.  
571 Journal of Agricultural and Food Chemistry, 51, 7913-7920.
- 572 Huerta-Ocampo, J.A., Barba de la Rosa, A.P., 2011. Amaranth: a pseudo-cereal  
573 with nutraceutical properties. Current Nutrition and Food Science 7, 1-9.
- 574 Hunt, H.V., Denyer, K., Packman, L.C., Jones, M.K., Hower, Ch.J., 2010.  
575 Molecular basis of the waxy endosperm starch phenotype in Broomcorn  
576 Millet (*Panicum miliaceum* L.). Molecular Biology and Evolution 27, 1478-  
577 1494.
- 578 Kong, X., Bao, J., Corke, H., 2009. Physical properties of *Amaranthus* starch.  
579 Food Chemistry 113, 371-376.
- 580 Lev-Yadun, S., 2016. Defensive (anti-herbivory) coloration in land plants. Seed  
581 Camouflage. 31-39. eBook Springer 2016.

- 582 Lobell, D.B., Schlenker, W., Costa-Roberts, J., 2011. Climate trends and global  
583 crop production since 1980. *Science* 333, 616–620.
- 584 Lush, W.M., Wein, H.C., 1980. The importance of seed size in early growth of  
585 wild and domesticated cowpeas. *Journal of Agriculture Science* 94, 177-  
586 182.
- 587 Matteucci, E., Giampietro, O., 2011. Dipeptidyl peptidase-4 inhibition: Linking  
588 chemical properties to clinical safety. *Current Medicinal Chemistry* 18,  
589 4753–4760.
- 590 McCouch, S., Baute, G.J., Bradeen, J., Bramel, P., Bretting, P.K., Buckler, E.,  
591 Burke, J.M., Charest, D., Cloutier, S., Cole, G., Dempewolf, H., Dingkuhn,  
592 M., Feuillet, C., Gepts, P., Grattapaglia, D., Guarino, L., Jackson, S.,  
593 Knapp, S., Langridge, P., Lawton-Rauh, A., Lijua, Q., Lusty, Ch., Michael,  
594 T., Myles, S., Naito, K., Nelson, R.L., Pontarollo, R., Richards, ChM.,  
595 Rieseberg, L., Ross-Ibarra, J., Rounsley, S., Hamilton R.S., Schurr, U,  
596 Stein, N., Tomooka, N., van der Knaap, E., van Tassel, D., Toll, J., Valls,  
597 J., Varshney, R.K., ward, J., Waugh, R., Wenzl, P., Zamir, D., 2013.  
598 *Agriculture: feeding the future. Nature* 499, 23–24.
- 599 Muñoz, N., Liu, A., Kan, L., Li, M.-W., Lam, H.-M., 2017. Potential uses of wild  
600 germplasms of grain legumes for crop improvement. *International Journal*  
601 *of Molecular Sciences* 18, 328.
- 602 Reddy, L.H., Couvreur, P., 2009. Squalene: A natural triterpene for use in  
603 disease management and therapy. *Advanced Drug Delivery Reviews* 61,  
604 1412-1426.
- 605 Sauer, J.D., 1967. The grain amaranths and their relatives: a revised taxonomic  
606 and geographic survey. *Annals of the Missouri Botanical Garden* 54, 103-  
607 137.
- 608 Shewry, P.R., Hey, S., 2015. Do “ancient” wheat species differ from modern  
609 bread wheat in their contents of bioactive components? *Journal of Cereal*  
610 *Science* 65, 236-243.
- 611 Siddhuraju, P., Becker, K., 2001. Species/variety differences in biochemical  
612 composition and nutritional value of Indian tribal legumes of the genus  
613 *Canavalia*. *Nahrung/Food* 4, 224-233.
- 614 Siró, I., Kápolna, E., Kápolna, B., Lugasi, A., 2008. Functional food product  
615 development, marketing and consumer acceptance. A review. *Appetite* 51,  
616 456–467.
- 617 Velarde-Salcedo, A.J., Barrera-Pacheco, A., Lara-González, S., Montero-Morán,  
618 G.M., Díaz-Gois, A., González de Mejia, E., Barba de la Rosa, A.P., 2013.  
619 *In vitro* inhibition of dipeptidyl peptidase IV by peptides derived from the  
620 hydrolysis of amaranth (*Amaranthus hypochondriacus* L.) proteins. *Food*  
621 *Chemistry* 136, 758-764.
- 622 WHO, 2016. World Health Organization. Diabetes fact sheet. Accessed May 1,  
623 2018. <http://www.who.int/mediacentre/factsheets/fs312/en/>.
- 624 Wright, H.H., 2005. The glycaemic index and sports nutrition, *South African*  
625 *Journal of Clinical Nutrition* 18, 222–228.
- 626 Zong, Y., Yao, Sh., Crawford, G.W., Fang, H., Lang, J., Fan, J., Sun, Zh., Liu, Y.,  
627 Zhang, J., Duan, X., Zhou, G., Xiao, T., Luan, F., Wang, Q., Chen, X.,

628 Jiang, H., 2017. Selection for oil content during soybean domestication  
629 revealed by X-ray tomography of ancient beans. *Scientific Reports* 7,  
630 43595.

631  
632

633

#### 634 **Figure Legends**

635 **Figure 1.** Morphological characteristics of amaranth wild and cultivated species

636 **Figure 2.** Scanning Electron Microscopy (SEM) micrograph of transversal cuts of  
637 amaranth seeds

638 **Figure 3.** 1-DE electrophoretic pattern of total proteins extracted from amaranth  
639 seed. Lanes: M=molecular weight marker; 1=*A. hybridus*, 2=*A. powellii*,  
640 3=*A. cruentus* cv *Amaranteca*, 4=*A. hypochondriacus* cv *Opaca* (waxy),  
641 5=*A. hypochondriacus* cv *Cristalina* (non-waxy), and 6=*A.*  
642 *hypochondriacus* cv *Nutrisol*.

643 **Figure 4.** Inhibitory activity of amaranth peptides released by gastrointestinal  
644 simulated digestion *in vitro* against A) DPPIV and B) ACE. Peptides  
645 bigger than 10 kDa were removed by ultrafiltration and ACE activity was  
646 measured at different peptides concentrations.

647

648 **Table 1**  
 649 Proximate composition of wild and cultivated amaranth species (%db)  
 650

Amaranth species	Protein <sup>1</sup>	Fat	Crude Fibre	Ash	Carbohydrates <sup>2</sup>
<i>A. hybridus</i>	15.9 ±0.1 <sup>b</sup>	5.9 ±0.0 <sup>c</sup>	6.1 ±0.0 <sup>a</sup>	3.7 ±0.1 <sup>a</sup>	68.5 ±0.2 <sup>d</sup>
<i>A. powellii</i>	17.8 ±0.1 <sup>a</sup>	8.1 ±0.0 <sup>a</sup>	5.9 ±0.0 <sup>b</sup>	3.6 ±0.1 <sup>a</sup>	64.5 ±0.2 <sup>e</sup>
<i>A. cruentus</i>	14.8 ±0.4 <sup>c</sup>	6.9 ±0.0 <sup>b</sup>	2.5 ±0.0 <sup>e</sup>	2.8 ±0.0 <sup>b</sup>	73.0 ±0.5 <sup>a</sup>
<i>A. hypochondriacus</i> cv Opaca	16.7 ±0.8 <sup>b</sup>	6.9 ±0.0 <sup>b</sup>	3.5 ±0.1 <sup>d</sup>	3.0 ±0.0 <sup>b</sup>	69.9 ±0.8 <sup>c</sup>
<i>A. hypochondriacus</i> cv Cristalina	16.7 ±0.1 <sup>b</sup>	5.7 ±0.1 <sup>c</sup>	3.9 ±0.0 <sup>c</sup>	2.9 ±0.0 <sup>b</sup>	70.9 ±0.3 <sup>b</sup>
<i>A. hypochondriacus</i> cv Nutrisol	15.8 ±0.1 <sup>b</sup>	6.9 ±0.1 <sup>b</sup>	2.4 ±0.0 <sup>f</sup>	3.5 ±0.2 <sup>a</sup>	71.4 ±0.1 <sup>b</sup>

<sup>1</sup>N×5.85; <sup>2</sup>As difference; Mean values of three replicates ± standard deviation; different superscript letters by column indicate statistically significant differences at  $p<0.05$ .

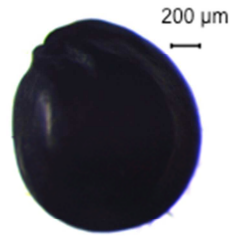
651  
 652  
 653  
 654  
 655  
 656  
 657  
 658

659 **Table 2**  
 660 Squalene quantification by GG-MS in wild and cultivated amaranth species

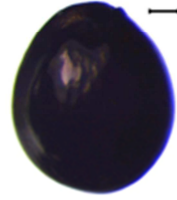
Amaranth species	Squalene (g/100g)	
	in seeds	in oil
<i>A. hybridus</i>	0.246 ±0.03 <sup>b</sup>	4.17 ±0.45 <sup>b</sup>
<i>A. powellii</i>	0.252 ±0.02 <sup>b</sup>	3.12 ±0.27 <sup>c</sup>
<i>A. cruentus</i>	0.335 ±0.02 <sup>a</sup>	4.86 ±0.31 <sup>a</sup>
<i>A. hypochondriacus</i> cv Opaca	0.271 ±0.01 <sup>b</sup>	3.93 ±0.08 <sup>b</sup>
<i>A. hypochondriacus</i> cv Cristalina	0.217 ±0.00 <sup>c</sup>	3.80 ±0.05 <sup>b</sup>
<i>A. hypochondriacus</i> cv Nutrisol	0.197 ±0.01 <sup>d</sup>	2.85 ±0.12 <sup>c</sup>

Values are the mean ± SD of three determinations.

661



*A. hybridus*



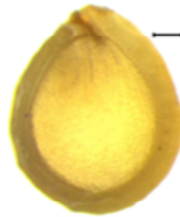
*A. powellii*



*A. cruentus* cv Amaranteca



*A. hypochondriacus*  
cv Opaca



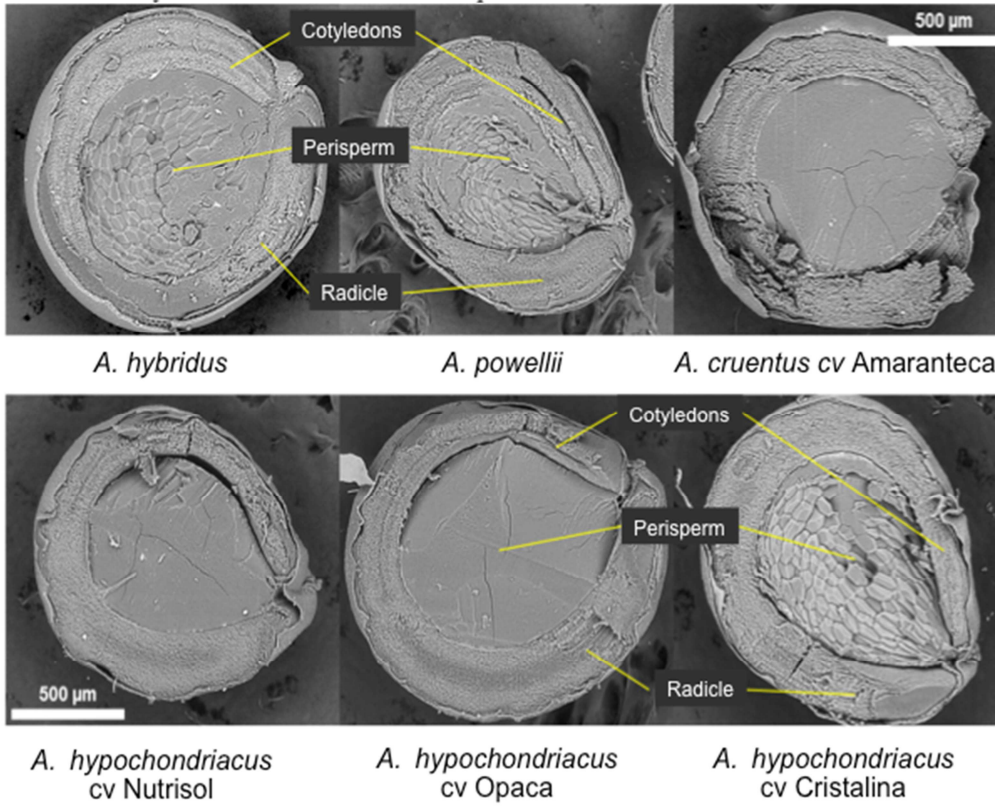
*A. hypochondriacus*  
cv Cristalina



*A. hypochondriacus*  
cv Nutrisol

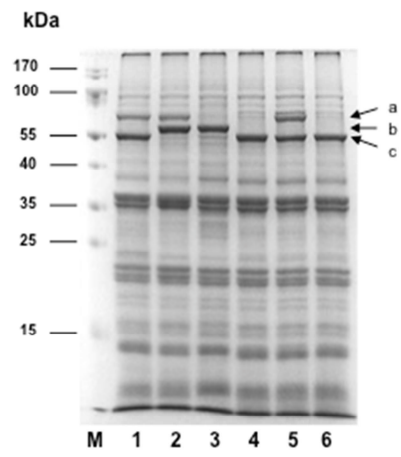
ACCEPTED MANUSCRIPT





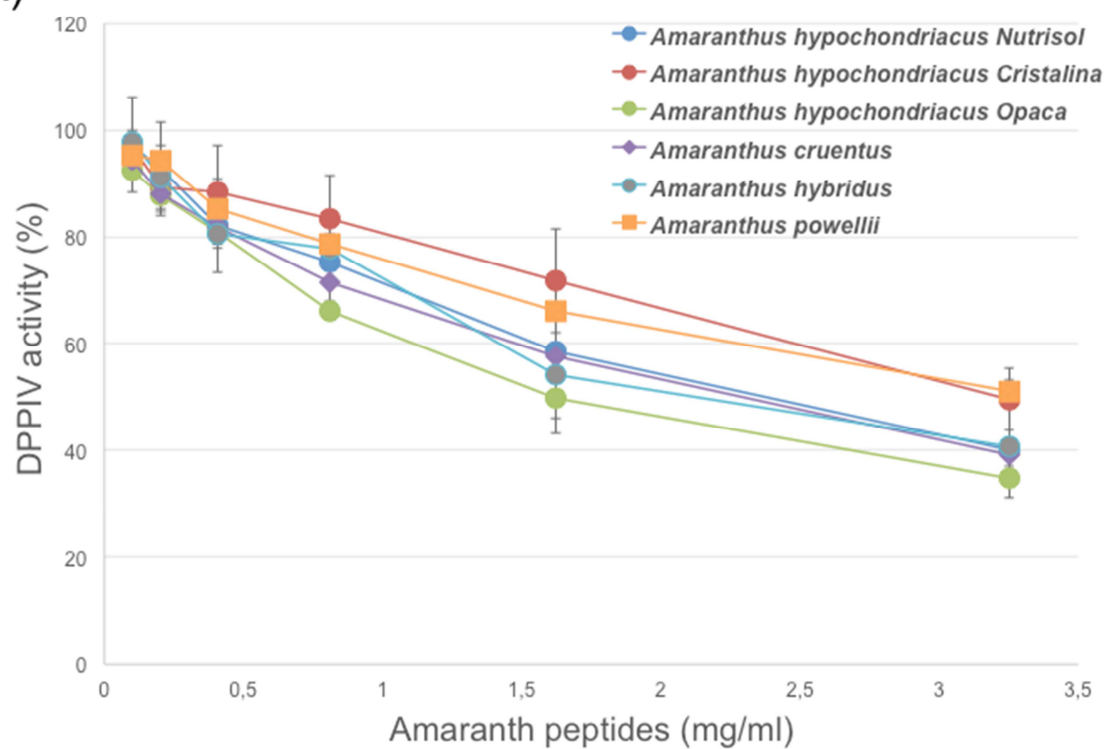
ACCEPTED MANUSCRIPT



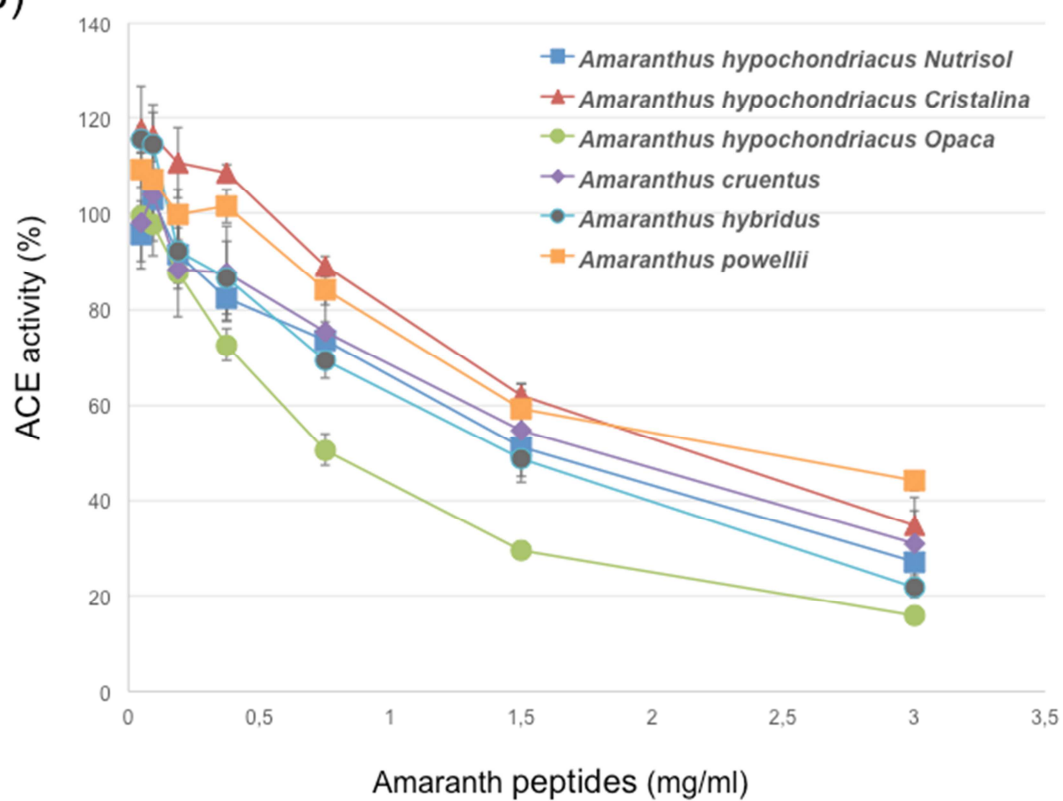


ACCEPTED MANUSCRIPT

A)



B)



## Highlights

- *A. powellii*, a wild species, showed the higher protein content among the amaranth species analyzed
- Protein profiles of wild and cultivated amaranth species showed high polymorphism in the high molecular weight region
- *A. hypochondriacus* cv Opaca released peptides inhibited 80% of angiotensin converting enzyme activity
- Squalene content in amaranth oil extract varies from 53-94% depending of the species