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## Article

# Expression of *EPL1* from *Trichoderma atroviride* in *Arabidopsis* Confers Resistance to Bacterial and Fungal Pathogens

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**Abstract:** During plant interaction with beneficial microorganisms, fungi secrete a battery of elicitors that trigger plant defenses against pathogenic microorganisms. Among the elicitor molecules secreted by *Trichoderma* are cerato-platanin proteins, such as *EPL1*, from *Trichoderma atroviride*. In this study, *Arabidopsis thaliana* plants that express the *TaEPL1* gene were challenged with phytopathogens to evaluate whether expression of *EPL1* confers increased resistance to the bacterial pathogen *Pseudomonas syringae* and the necrotrophic fungus *Botrytis cinerea*. Infection assays showed that *Arabidopsis EPL1-2*, *EPL1-3*, *EPL1-4* expressing lines were more resistant to both pathogens in comparison to WT plants. After *Pseudomonas syringae* infection, there were reduced disease symptoms (e.g., small chlorotic spots) and low bacterial titers in the three *35S::TaEPL1* expression lines. Similarly; *35S::TaEPL1* expression lines were more resistant to *Botrytis cinerea* infection, showing smaller lesion size in comparison to WT. Interestingly, an increase in ROS levels was detected in *35S::TaEPL1* expression lines when compared to WT. A higher expression of SA- and JA-response genes occurred in the *35S::TaEPL1* lines, which could explain the resistance of these *EPL1* expression lines to both pathogens. We propose that *EPL1* is an excellent elicitor, which can be used to generate crops with improved resistance to broad-spectrum diseases.

**Keywords:** *Arabidopsis thaliana*; *Botrytis cinerea*; cerato-platanin proteins; elicitor; *Pseudomonas syringae*; *Trichoderma atroviride*



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## 1. Introduction

The genus *Trichoderma* comprises many rhizocompetent filamentous fungi found in different ecosystems [1]. *Trichoderma* species are fast-growing, opportunistic invaders and prolific producers of secondary metabolites with antibiotic activity that suppresses diverse phytopathogens such as bacteria, fungi, and nematodes [2–4]. These properties make these fungi an ecologically dominant species. Other biocontrol mechanisms used by *Trichoderma* include mycoparasitism and competition for space and nutrients [5]. Due to the ability of *Trichoderma* spp. to suppress different phytopathogenic fungi, some species have been widely used in agriculture as biological control agents [6,7].

Some *Trichoderma* species have the ability to establish symbiotic relationships with plants. This symbiosis is achieved through crosstalk, in which plants and *Trichoderma* produce a wide range of chemical compounds that modify the transcriptomes, proteomes, and metabolomes of host plants [8,9]. As a result, *Trichoderma* species produce a wide range

of compounds such as elicitors, siderophores, phytohormones, and volatile organic compounds (VOCs) that modulate plant growth and defense [10–12]. Elicitors are molecules that originate from the host plant (endogenous elicitors) or from the microbe (exogenous elicitors) and can induce biochemical and/or structural responses associated with the expression of resistance to plant diseases [13]. Among the elicitors produced by *Trichoderma* are the cerato-platanin proteins, which can activate the plant defense system against different kinds of phytopathogens [14].

Cerato-platanins (CPs) are proteins secreted by fungi that participate in diverse stages of the host-fungus interaction, acting as virulence factors or elicitors [15]. These CPs are composed of small cysteine-rich proteins of approximately 100 to 130 residues, and they have a signal peptide for their secretion [16]. The first CPs elicitors studied in *Trichoderma* were SM1 from *T. virens* and Eliciting Protein-Like (EPL1) from *T. atroviride* [14,17]. Exogenous application of SM1 purified from *T. virens* [14] as well as recombinant SM1 generated in the yeast *Pichia pastoris* [18] increased the expression of both local and systemic defense-related genes in cotton and maize plants. It was shown that the exogenous application of purified SM1 on cotton cotyledons reduced the size of the lesion caused by the pathogenic fungus *Colletotrichum* sp. [14]. In addition, maize and tomato plants inoculated with *T. virens* Gv29-8 that overexpressed SM1 were more resistant to necrotrophic and biotrophic phytopathogens, respectively [19,20].

The Eliciting Plant Response-Like 1 (Epl1) elicitor is produced and exported by *T. atroviride* to the plant, and it is important in the interaction of fungi with plants because it stimulates the induction of defense responses in the plant [20,21]. *T. atroviride* strains (TaOE) that overexpress the *EPL1* gene increase disease resistance against different phytopathogens in tomato plants, whereas *Epl1* knockout mutants conferred less protection in tomato plants against *Alternaria solani* and *B. cinerea* [20]. While there is evidence that the EPL1 elicitor of *T. atroviride* activates the plant's defense system, to the best of our knowledge, there are no reports on the molecular mechanism of this elicitor in plants.

The exogenous application of Epl1 protein from *T. asperellum* strain T4 (expressed and purified from *Pichia pastoris*) to soybean leaves showed protective activity against the pathogen *Cercosporidium sofinum* [22]. A similar study showed that the application of EPL1 protein of *T. asperellum* also produced in *P. pastoris* significantly reduced the infection of *Populus* plantlets by *A. alternata* [23].

Other fungal CPs have been shown to induce resistance in plants by exogenous treatments. In the fungus *Botrytis cinerea*, CP BcSpl1 is one of the most abundant proteins in the secretome of this plant pathogen. Knockout mutants of *bcspl1* showed a reduction in virulence on hosts [24]. It was shown that the exogenous application of the BcSpl1 protein by infiltration in tobacco leaves induces resistance against *Pseudomonas syringae* and *B. cinerea*, which correlated with the induction of the PR-1 $\alpha$  and PR-5 genes [25]. In addition, the CP Pop1 secreted by *Ceratocystis populicola*, a poplar pathogen, has been shown to induce phytoalexin synthesis, production of ROS and NO, and the induction of defense related genes in *Platanus acerifolia* leaves [15].

The heterologous expression of the SM1 elicitor of the phytopathogenic fungus *Magnaporthe oryzae* in *Arabidopsis* and rice plants has also been reported. Transgenic lines of *Arabidopsis* and rice that express SM1 have a greater resistance against fungal and bacterial diseases [26,27]. However, transient expression of MoSM1 in rice leaves or high expression levels of the MoSM1 gene in transgenic lines of *Arabidopsis* generated HR-like necrosis. This could be due to the fact that some CPs of phytopathogenic fungi, such as CP from *Ceratocystis fimbriata* f. sp. *platani* [28], BcSpl1 from *B. cinerea* [24], and MoSM1 or MoMSP1 from *M. oryzae*, cause phytotoxic effects in different plants [26,29].

To determine if the SM1 homolog of a symbiotic fungus expressed in *Arabidopsis* confers disease resistance but without producing the toxic effect on the plant, we generated *Arabidopsis thaliana* plants that express the *EPL1* gene from *T. atroviride*. These *Arabidopsis* TaEPL1 expressing lines were infected with the bacterium *P. syringae* and the necrotrophic fungus *B. cinerea*. We achieved a higher resistance against these pathogens in plants

expressing *TaEPL1*. We also observed high levels of hydrogen peroxide accumulation in *TaEPL1* expressing lines compared to WT plants. Expression analysis of plant hormone-related genes showed that the transgenic *EPL1* lines induced SA and JA related genes, which could explain the enhanced resistance of these transgenic lines to the tested pathogens.

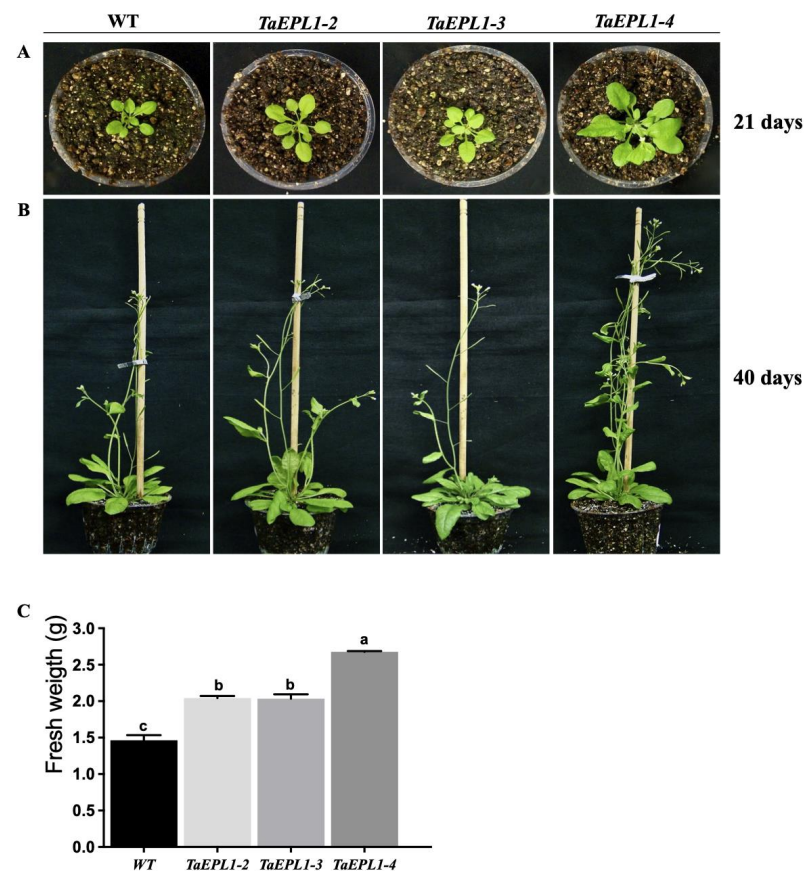
## 2. Results

### 2.1. Generation of Arabidopsis *EPL1* Expressing Lines (*35S::TaEPL1*)

We generated *Arabidopsis* transgenic plants expressing the *EPL1* elicitor gene from the beneficial fungus *T. atroviride* (*TaEPL1*). The *35S::TaEPL1* construct (Supplementary Figure S1A) was transformed by floral dip in *A. thaliana* ecotype Col-0 (WT). The *EPL1* gene expression of three *35S::TaEPL1* lines (*EPL1-2*, *EPL1-3* and *EPL1-4*) was quantified by RT-qPCR in 15-day-old plants. The highest *EPL1* mRNA expression level was found in the *EPL1-3* line, followed by the *EPL1-4* and *EPL1-2* expression lines. As expected, *EPL1* transcript was not detected in the *Arabidopsis* WT plants (Supplementary Figure S1B).

### 2.2. The Expression of the *EPL1* Gene in Arabidopsis Results in Accelerated Growth

The expression of the Trichoderma *EPL1* gene in Arabidopsis (*35S::TaEPL1* lines) resulted in a positive impact on its growth, as observed in 21-day-old plants grown in soil (Figure 1A). In particular, the *35S::TaEPL1-4* line displayed the highest development among the three *TaEPL1* expressing lines (Figure 1A). As previously observed, 40-day-old plants of the *35S::TaEPL1-4* exhibited the most significant development, while the parental Col-0 (WT) displayed the lowest growth (Figure 1B). We evaluated the fresh weight of the aerial parts of each 40-day-old plant, and all three *TaEpl1* expressing lines exhibited higher fresh weights than the WT. Among them, the *35S::TaEpl1-4* line exhibited the highest biomass, as evidenced by a greater number of developed inflorescences (Figure 1B,C).

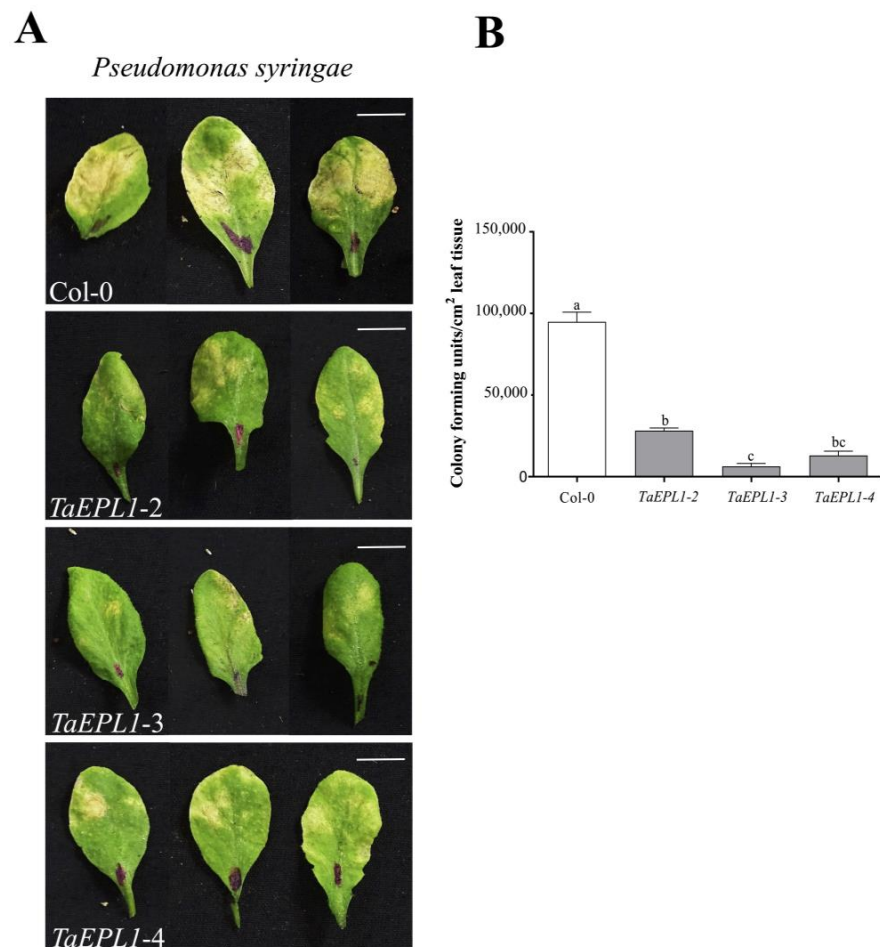


**Figure 1.** Phenotype of Arabidopsis *35S::TaEPL1* expressing lines. Photographs of Arabidopsis plants grown on soil pots under long day conditions, showing the WT, *35S::EPL1-2*, *35S::EPL1-3*, and

35S::EPL1-4 lines at two different stages: 21-day-old (A) and 40-day-old (B). Data on fresh weight (g) of the leaf area in 40-day-old plants for the following lines: WT, 35S::EPL1-2, 35S::EPL1-3, and 35S::EPL1-4 (C). Data are means  $\pm$  SE ( $n = 6$ ). Statistical analysis between genotypes was determined by One-way ANOVA, and the letters indicate statistically significant differences by Tukey's test at  $p < 0.05$ .

### 2.3. The Arabidopsis 35S::TaEPL1 Lines Are More Resistant to Infection by *Pseudomonas Syringae*

The Arabidopsis TaEPL1 expression lines were inoculated with *P. syringae* pv. tomato DC3000 strain (*Pst*) to examine whether the expression of the TaEPL1 elicitor confers resistance against bacterial infection. Four-week-old Arabidopsis EPL1-2, EPL1-3, EPL1-4, and WT plants were inoculated with *Pst* bacterial suspension. Disease symptoms and bacteria colony-forming units were recorded after 72 hpi (Figure 2). We observed more severe symptoms with larger chlorotic spots in the WT plants than the 35S::TaEPL1 expression lines (Figure 2A). This is in agreement with the low levels of bacterial titers detected in the Arabidopsis 35S::TaEPL1 expression lines, which were up to 14.8-fold lower than that obtained in the WT plants (Figure 2B). The EPL1-3 line presented the lowest bacterial titers ( $6.3 \times 10^3$ ) at 72 hpi and showed milder symptoms of *Pst* infection in comparison to the other two overexpressing lines (Figure 2B). This correlates with the EPL1-3 line having the highest EPL1 transcript levels among the transgenic lines (Supplementary Figure S1). Our data reveal that expression of the EPL1 gene from the beneficial fungus *Trichoderma* in Arabidopsis plants confers marked resistance to *P. syringae* infection.

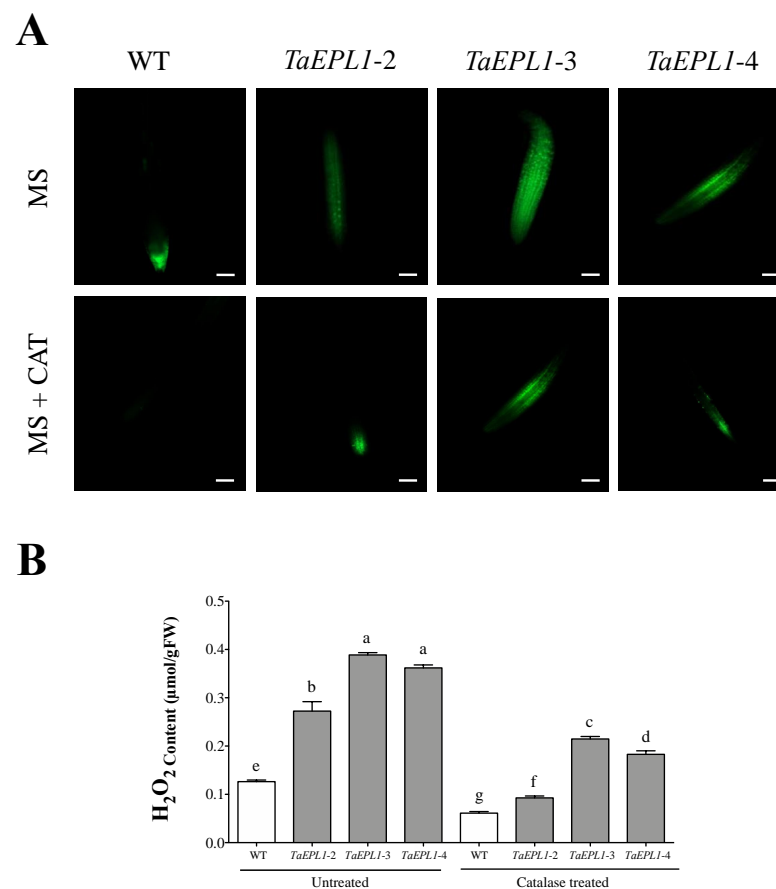


**Figure 2.** *Pseudomonas syringae* infection of Arabidopsis 35S::TaEPL1 and WT leaves. (A) Disease symptoms in leaves of four-week-old 35S::TaEPL1 (TaEPL1-2, TaEPL1-3 and TaEPL1-4) and WT plants

72 h after infection with *P. syringae* DC3000 strain. In the images, the scale bar indicates 1 cm. **(B)** To determine colony-forming units (CFU), five independent 0.5 cm<sup>2</sup> leaf disks from 35S::TaEPL1 and WT plants were ground, and then the ground leaves extract was spotted onto LB plates with rifampicin to count the bacterial colonies formed. Data represented graphically. The bars denote the  $\pm$ SE from five biological replicates ( $n = 5$ ). Statistical analysis between genotypes was determined by One-way ANOVA, and the letters indicate statistically significant differences by Tukey's test at  $p < 0.05$ .

#### 2.4. The Arabidopsis 35S::TaEPL1 Lines Display Increased Reactive Oxygen Species (ROS)

In plants, ROS play an important role in plant defense against phytopathogen infection [30]. We analyzed whether the 35S::TaEPL1 lines produced a higher amount of ROS as a possible mechanism for resistance against microbes. We detected ROS in root tips of 10-day-old 35S::TaEPL1 and WT plantlets using 2,7-dichlorofluorescein diacetate (DCFH2-DA) dye and fluorescence microscopy. ROS signal, detected as green fluorescence, was more intense in 35S::TaEPL1 lines compared to WT roots (Figure 3A). Catalase (CAT) is a key antioxidant enzyme that alleviates oxidative stress through decomposition of hydrogen peroxides to water and oxygen. The CAT treatment was carried out to determine its effect on H<sub>2</sub>O<sub>2</sub> levels between the 35S::TaEPL1 and WT lines. When we applied 250 U/mL CAT on 35S::TaEPL1 and WT plants, H<sub>2</sub>O<sub>2</sub>-associated fluorescence in the root tips in all transgenic lines was decreased, whereas in WT plants, no signal was detected (Figure 3A). A quantitative analysis of H<sub>2</sub>O<sub>2</sub> confirmed that 35S::TaEPL1 lines accumulated a higher amount of hydrogen peroxide, with the highest levels observed in the 35S::TaEPL1-3 line (3-fold more than WT) (Figure 3B). Moreover, upon the application of CAT enzyme (250 U/mL), both the transgenic 35S::TaEPL1 lines and the WT had decreased H<sub>2</sub>O<sub>2</sub> levels, but the transgenic lines maintained the highest values (Figure 3B).

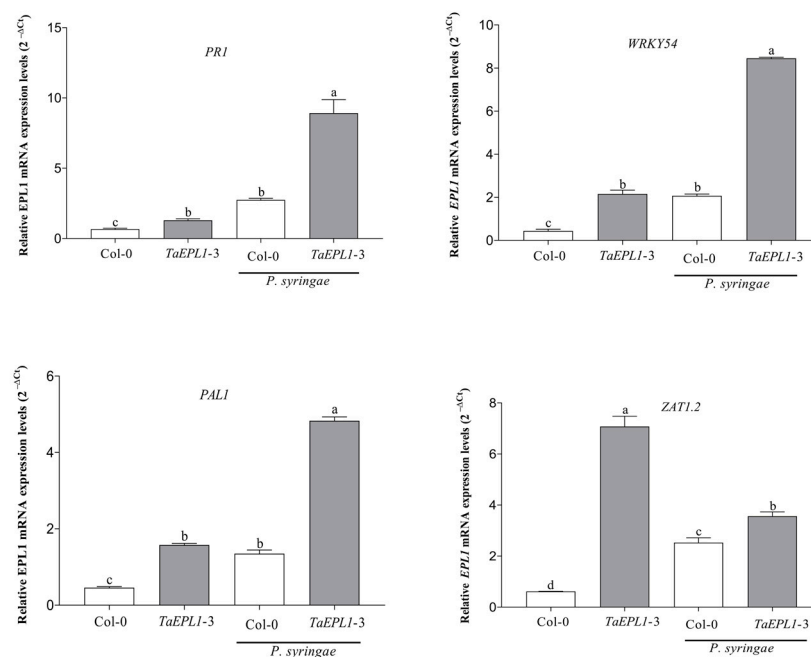


**Figure 3.** Detection of DCFH2-DA fluorescence in *Arabidopsis* 35S::TaEPL1 lines and WT root tips. Representative images in root tips of ten-days-old WT and 35S::TaEPL1 seedlings (TaEPL1-2, TaEPL1-3,

and *TaEPL1-4*). (A) Untreated (0.2X MS liquid medium) and catalase treated (0.2X MS + 250 U/mL CAT) root tips. Images were acquired on a Zeiss Axio Imager M2 microscope under 10× magnification. The scale bar corresponds to 100 μm. (B) H<sub>2</sub>O<sub>2</sub> quantification was performed using KI in *A. thaliana* WT and *35S::TaEPL1* seedlings. Conditions shown are untreated (0.2X MS liquid medium) and catalase treated (0.2X MS + 250 U/mL CAT). The data show the means ± SE from three biological replicates ( $n = 3$ ). The variance analysis was done by Two-way ANOVA (genotype × catalase), and the letters indicate statistically significant differences by Tukey's test at  $p < 0.05$ .

### 2.5. The *35S::TaEPL1-3* Line Shows a Higher Accumulation of Transcripts of SA-Related Genes under *Pseudomonas Syringae* Infection

Salicylic acid (SA) and reactive oxygen species (ROS) play important functions in the activation of plant defense under pathogen attacks. The expression levels of SA-mediated defense markers such as *PR1*, *PAL1*, and *WRKY54*, as well *ZAT1.2* (signaling ROS gene), were quantified by RT-qPCR in rosette leaves of 28-day-old WT and *35S::TaEPL1-3* plants inoculated with *P. syringae* (Figure 4). After 24 h of bacterial infection, we observed higher mRNA expression levels of the four genes analyzed (*PR1*, *PAL1*, *WRKY54* and *ZAT1.2*) in the *35S::TaEPL1-3* line compared to the WT (Figure 4). This same behavior was observed in the mock inoculated plants, in which all the measured markers showed a higher expression in the *35S::TaEPL1-3* line than the WT (Figure 4). This result is in accordance with the observation that this *EPL1* expressing line is also the most resistant to *P. syringae*.

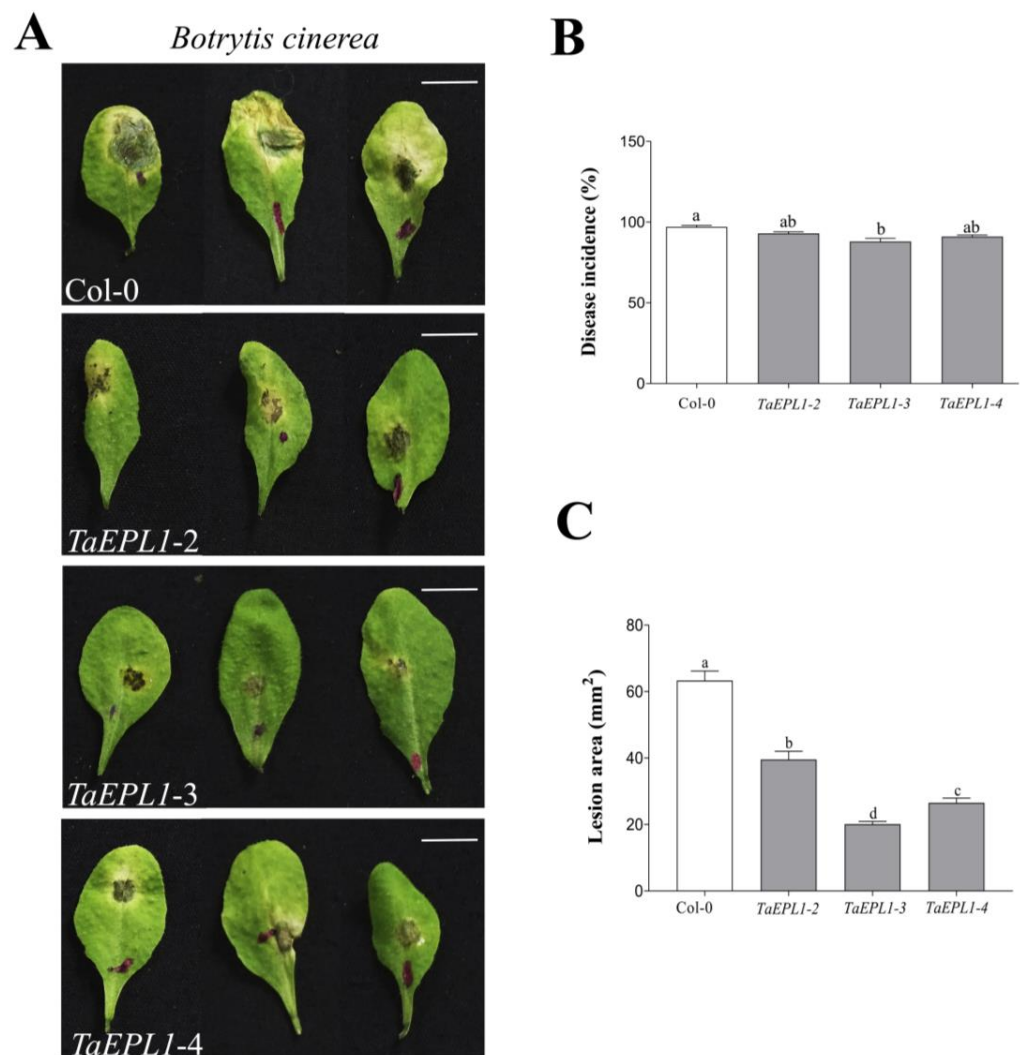


**Figure 4.** Expression analysis of plant defense genes in WT and *35S::TaEPL1-3* line during *Pseudomonas syringae* infection. Leaves of four-week-old WT and the *35S::TaEPL1-3* plants were infected or not with *P. syringae* for 24 h. The expression levels of *PR1*, *WRKY54*, *PAL1*, and *ZAT1.2* genes were measured by RT-qPCR. Values were expressed as relative expression levels ( $2^{-\Delta C_t}$ ) calculated after normalization to the *A. thaliana* *UBQ5* gene. For each sample, three biological replicates were analyzed with their respective technical replicates. The variance analysis was done by Two-way ANOVA (genotype × infection), and the letters indicate statistically significant differences by Tukey's test at  $p < 0.05$ .

### 2.6. The *Arabidopsis 35S::TaEPL1* Lines Were More Resistant to *Botrytis cinerea* Infection

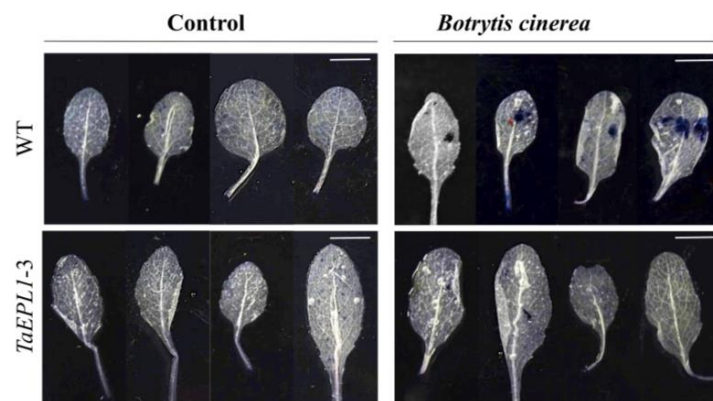
The resistance of *Arabidopsis* *EPL1*-expressing plants against infection by the necrotrophic fungus *B. cinerea* strain B05.10 was assessed. Four-week-old plants expressing *EPL1* gene (*EPL1-2*, *-3*, *-4*) were inoculated with the *B. cinerea* spore solution and compared with WT plants. After 72 hpi, the incidence of infection and leaf lesion size were evaluated. Figure 5A

shows that the *35S::TaEPL1* expressing lines were more resistant to fungus infection than WT plants, as observed by decreased leaf necrosis and less water-soaking development. Necrotic lesion formation in the *35S::TaEPL1* lines showed reductions in a lesion size of  $4.39 \pm 0.1 \text{ mm}^2$  for *EPL1-3*,  $5.07 \pm 0.1 \text{ mm}^2$  for the *EPL1-4* line and  $6.13 \pm 0.2 \text{ mm}^2$  for the *EPL1-2* line, while the WT leaves had lesions of  $7.77 \pm 0.2 \text{ mm}^2$  (Figure 5C). Even though disease incidence was not strongly reduced in the *EPL1* expressing lines, the *EPL1-3* line had the lowest necrotic lesion size caused by the fungus (Figure 5B), which agrees that this *EPL1* expressing line is the most resistant to the bacteria (Figure 2). In addition, we selected the *EPL1-3* line for trypan blue staining after inoculation with *B. cinerea* (Figure 6). We observed that the blue staining was lower in the *EPL1-3* line, while a strong signal was observed in the WT leaves (Figure 6). This confirms a decrease in plant cell death that correlates with a smaller lesion size in the transgenic *EPL1-3* line.



**Figure 5.** Leaf infection of *Arabidopsis 35S::TaEPL1* and WT by *Botrytis cinerea*. **(A)** Disease symptoms in leaves from four-week-old *35S::TaEPL1* (*TaEPL1-2*, *TaEPL1-3*, and *TaEPL1-4*) and WT plants at 72 h after infection with *B. cinerea* strain B05.10. For each line, three representative images of the infected leaves are shown. In the images, the scale bar indicates 1 cm. **(B)** The disease incidence graph shows the number of infection events with respect to the total inoculated leaves ( $n = 50$ ) and expressed as a percentage. **(C)** Lesion area measurements ( $\text{mm}^2$ ) of total infection events. The bars denote the  $\pm$ SE. Statistical analysis between genotypes was determined by One-way ANOVA, and the letters indicate statistically significant differences by Tukey's test at  $p < 0.05$ .

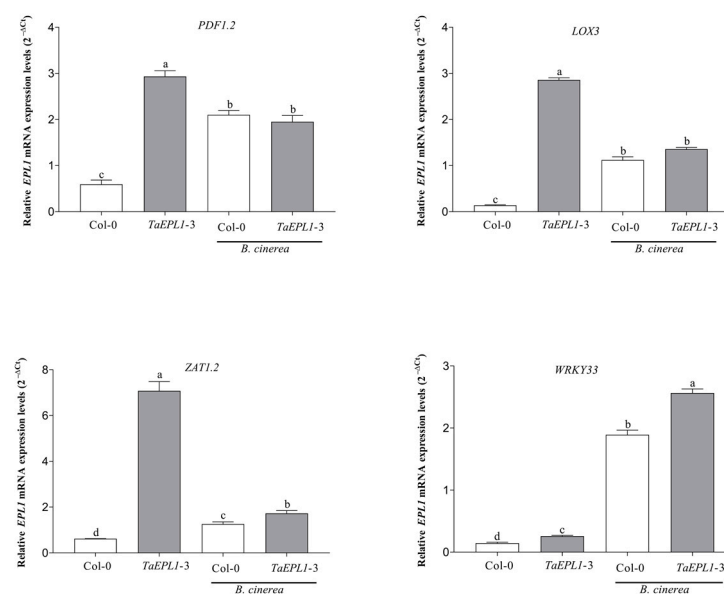




**Figure 6.** Trypan blue staining indicating cell death caused by *Botrytis cinerea* infection in leaves of *35S::TaEPL1-3* line and WT. Representative images of the area of cell death in *EPL1-3* and WT leaves inoculated with *B. cinerea*. In the images, the scale bar indicates 1 cm. Leaves of four-week-old WT and *EPL1-3* plants were infected with  $2 \times 10^5$  spores of *B. cinerea* strain B05.10. Leaves were collected 24 h after inoculation, and then stained in a trypan blue solution. As control, PDB (0.25X) was used as a mock treatment.

### 2.7. Expression Changes Plant Defense Genes in the *35S::TaEPL1-3* Line under *Botrytis cinerea* Infection

We analyzed the expression levels of the *LOX3* and *PDF1.2* genes involved in JA biosynthesis and response, respectively, *WRKY33* gene, which is a transcription factor implicated in plant defense against necrotrophic pathogens [31], and also *ZAT1.2* gene under *B. cinerea* infection. The gene expression levels were evaluated by RT-qPCR in rosette leaves of 28-day-old WT and *35S::TaEPL1-3* plants inoculated with *B. cinerea* for 24 h. In plants inoculated with the fungus, a higher expression was noticed in the *ZAT1.2* and *WRKY33* genes in the *35S::TaEPL1-3* line than WT; whereas, the *PDF1.2* and *LOX3* genes were similarly induced by the fungus between the WT and the *35S::TaEPL1-3* line (Figure 7). In the mock treatment, the *35S::TaEPL1-3* line had higher expression of all marker genes than the parental WT (Figure 7). Our data show that the *35S::TaEPL1-3* line has up-regulated plant defense genes against necrotrophic pathogens, which could prevent the spread of the fungus when plants were inoculated.

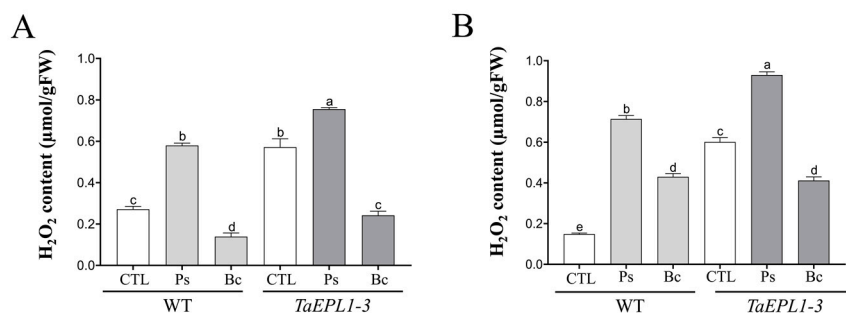


**Figure 7.** Expression analysis of plant defense genes in *A. thaliana* WT and *35S::TaEPL1-3* line during *Botrytis cinerea* infection. Leaves of four-week-old WT and the *35S::TaEPL1-3* plants were infected or

not with fungus for 24 h. Expression patterns of *PDF1.2*, *LOX3*, *WRKY33*, and *ZAT1.2* genes of *A. thaliana* were assessed by RT-qPCR. Values were expressed as relative expression levels ( $2^{-\Delta Ct}$ ) calculated after normalization to the *A. thaliana* *UBQ5* gene. For each sample, three biological replicates were analyzed with their respective technical replicates. The variance analysis was done by Two-way ANOVA (genotype x infection), and the letters indicate statistically significant differences by Tukey's test at  $p < 0.05$ .

### 2.8. Determination of $H_2O_2$ Content in *Arabidopsis* WT and *35S::TaEPL1-3* Lines under *B. cinerea* and *P. syringae* Infection

We analyzed the  $H_2O_2$  accumulation in the 15 day-old WT and *35S::TaEPL1-3* plantlets under *B. cinerea* and *P. syringae* infection at 24 and 48 hpi (Figure 8). As observed in Figure 3, the *35S::TaEPL1-3* line accumulates a higher amount of ROS than WT under mock conditions (Figure 8). Despite the WT line showing an increase in  $H_2O_2$  levels during bacterial infection at both 24 and 48 hpi, the *35S::TaEPL1-3* line exhibited the highest  $H_2O_2$  accumulation (Figure 8). During *B. cinerea* infection, a decrease in  $H_2O_2$  levels was observed at 24 and 48 hpi in the *35S::TaEPL1-3* line compared to their respective mock controls, whereas in the WT ecotype, there was a reduction in  $H_2O_2$  levels at 24 hpi followed by an increase at 48 hpi compared to their mock controls (Figure 8).



**Figure 8.** Hydrogen peroxide quantification in *A. thaliana* WT and *35S::TaEPL1-3* lines under *P. syringae* and *B. cinerea* infection.  $H_2O_2$  quantification was performed using KI in *A. thaliana* 15 day-old WT and *35S::TaEPL1-3* plantlets. The data were represented graphically at 24 h (A) and 48 h (B) after the infection with *P. syringae* (Ps) or *B. cinerea* (Bc). The data show the means  $\pm$  SE from three biological replicates ( $n = 3$ ). The variance analysis was done by Two-way ANOVA (genotype x infection), and the letters indicate statistically significant differences by Tukey's test at  $p < 0.05$ .

### 3. Discussion

*Trichoderma* species are widely used in agriculture as a beneficial fungus due to its biocontrol activity and promotion of plant growth [1,32]. In the plant-*Trichoderma* interaction, both organisms secrete molecules for their recognition, initiating a molecular communication to achieve symbiosis between them [12,33–39]. Among the molecules reported to be secreted by *Trichoderma* are elicitors, such as Eliciting Plant Response-Like 1 (Epl1), which is produced and secreted by *T. atroviride*. There is evidence of a protective role of Epl1 protein from *T. atroviride* against foliar maize pathogen *C. graminicola* [21], and also strains of *T. atroviride* that overexpress *TaEPL1* gene increased resistance in tomato against *A. solani*, *B. cinerea*, and *P. syringae* infection [20]. Although the generation of *Arabidopsis* and rice plants expressing the SM1 elicitor of the phytopathogenic fungus *M. oryzae* has been reported, the effect of the expression in *A. thaliana* of an elicitor as EPL1 from beneficial fungus like *Trichoderma* and its resistance against pathogens has not been studied.

We describe here the response of *A. thaliana* plants that express the EPL1 elicitor to the infection by two phytopathogenic microorganisms. One of these pathogens was the hemibiotrophic *P. syringae*, in which *A. thaliana* *35S::TaEPL1* lines were more resistant to bacterial infection. These *Arabidopsis* *35S::TaEPL1* lines exhibited less damage caused by infection as well as lower bacterial load per leaf compared to the WT plants. This suggests

that the EPL1 elicitor activates a defense response within the plant cells, thus generating a lower susceptibility to *P. syringae*.

We report that the expression of the EPL1 elicitor in *Arabidopsis* triggers H<sub>2</sub>O<sub>2</sub> accumulation even in the absence of pathogen inoculation. Furthermore, despite applying CAT, a H<sub>2</sub>O<sub>2</sub> scavenger, it did not completely reduce the H<sub>2</sub>O<sub>2</sub>-associated fluorescence signal in root tips of the transgenic lines, as observed in the WT. In this sense, the 35S::TaEPL1-3 line showed a higher ZAT1.2 transcript level due to the infection of the bacterium as well as in the control plants (mock), which correlates with ROS accumulation in these transgenic lines. The ZAT1.2 gene encoding a zinc finger protein is involved in oxidative stress responses [40]. After inoculation with *P. syringae*, the TaEPL1-3 expression line had a higher accumulation of PAL1, PR1, and WRKY54 transcripts, which are involved in the salicylic acid (SA) biosynthesis, response, and regulation, respectively. An interesting fact is that the highest accumulation of H<sub>2</sub>O<sub>2</sub> was observed in the 35S::TaEPL1-3 line after bacterial infection, both at 24 and 48 hpi. The accumulation of ROS and the induction of these SA response genes could explain the resistance shown by the 35S::TaEPL1 lines to infection by *Pseudomonas*. It was previously reported that the plant protective activity of SM1 and other CP proteins are associated with the accumulation of ROS and phytoalexins [26,41,42].

Likewise, we found that the expression of the TaEPL1 gene in *Arabidopsis* confers enhanced resistance against *B. cinerea* infection. The *Arabidopsis* 35S::TaEPL1 plants showed a smaller lesion area and a reduction of leaf cell death caused by the fungus compared to the WT plants. This is consistent with a higher expression of the WRKY33 gene in the 35S::TaEPL1 line upon infection with the fungus *B. cinerea*. The WRKY33 transcription factor has been reported to be involved in defense responses against *B. cinerea* and other necrotrophic pathogens [31]. TaEPL1 expression in *Arabidopsis* reduces fungal colonization of plant tissue, thus making them more resistant to this phytopathogen. An interesting observation was that the LOX3, PDF1.2, WRKY33, and ZAT1.2 genes were up-regulated in the 35S::TaEPL1-3 line under mock conditions (i.e., no fungal infection), which could suggest that EPL1 expression in the plant causes a priming of genes involved in plant defense against this necrotrophic pathogen. An important finding was that a reduction in H<sub>2</sub>O<sub>2</sub> levels was observed in the 35S::TaEPL1-3 line during infection with *B. cinerea* at 24 and 48 hpi compared to the mock controls. This behavior suggests that the EPL1 expressing line is actively regulating its ROS levels in response to the presence of this necrotrophic pathogen.

Our data provide evidence that TaEPL1 expressed in *Arabidopsis* plants can activate defense responses against pathogens such as *P. syringae* and *B. cinerea*, which activate the SA- and JA/ET pathways. An ortholog to EPL1 exists in *M. oryzae* called MoSM1 [27]. Expression of MoSM1 in rice significantly increased SA and JA content, and also induced SA- and JA-related biosynthesis genes and signaling under normal growth conditions [27]. In addition, the heterologous expression of the MgSM1 in *A. thaliana* plants confers a broad-spectrum resistance against *B. cinerea*, *Alternaria brassicicola*, and *P. syringae* [26]. In the MgSM1-expressing plants, some defense genes such as PR1, PR5, and PDF1.2 were upregulated, and an accumulation of ROS was reported [26]. The authors changed the 35S viral promoter to an inducible promoter, since large amounts of MgSM1 protein caused a hypersensitive response in *Arabidopsis*, while this phenotype has not been reported in the case of *T. virens* SM1 purified protein [18], nor in our study here with TaEPL1. We did not observe a visible hypersensitive response phenotype in any 35S::TaEPL1 lines. This could be due to the fact that *M. grisea* is a phytopathogenic fungus that causes severe disease in rice and other grasses, and the constitute expression of SM1 *in planta* generated a hypersensitive response with a greater accumulation of ROS, while species of the genus *Trichoderma* are plant symbionts, and so the higher expression of EPL1 did not generate any negative phenotype in the plant. In addition to SM1 from *M. grisea*, phytotoxic effects have been reported for cerato-platanin, such as BcSpl1 from *B. cinerea* [24], a CP of 12.4 kDa from *Ceratocystis fimbriata* f. sp. *platani* [28].

On the other hand, Arabidopsis plants carrying the 35S::TaEPL1 construct exhibited increased biomass, with the TaEPL1-4 line showing the most pronounced accelerated growth. It is noteworthy that expressing a fungal elicitor, such as EPL1 (from the strain *T. atroviride* IMI 206040), in Arabidopsis results in the generation of more vigorous plants. Several species of *Trichoderma* have been reported to exert beneficial effects on plant growth [11]. In particular, the plant growth-promoting effect has been tested using the strain *T. atroviride* (IMI 206040) in tomato plants [20] and *A. thaliana* plants [12,32,34]. Further studies focused on the mechanism of EPL1 gene are needed to elucidate its role in plant growth.

## 4. Materials and Methods

### 4.1. Plant Growth Conditions

Seeds of Arabidopsis 35S::TaEPL1 lines generated in this work and parental Col-0 (WT) were sterilized using 20% (v/v) commercial sodium hypochlorite (6% free chlorine) solution for 5 min, and washed seven times in sterile distilled water. Aseptic seeds were stratified during 48 h at 4 °C and afterwards germinated and grown on agar plates containing 0.2X MS pH 7, 0.5% (w/v) sucrose, and 0.8% (w/v) agar. Seedlings were incubated in a growth chamber with a photoperiod of 16 h (100  $\mu\text{molm}^{-2} \text{s}^{-1}$ )/8 h with a light/dark cycle at a temperature of  $22 \pm 1$  °C until used in the experiments described above.

### 4.2. Generation of Arabidopsis thaliana 35S::TaEPL1 Lines

The EPL1 ORF (417 bp) was amplified from cDNA of mycelial tissue of the fungus *Trichoderma atroviride* with Pfu DNA Polymerase high fidelity (Thermo Scientific™, Waltham, MA, USA) using the primers: EPL1-Fw 5'ATGCAGTTCTCCAGCCTCTTCAAG3' and EPL1-Rv 5'TTAGAGGCCGCGAGTTGCTCACAGC3'. The product was cloned into the pCR8/GW/TOPO vector (Invitrogen, Carlsbad, CA, USA). The entry clone was verified by sequencing and recombined into pMDC32 binary vector by the Gateway LR Clonase enzyme mix (Invitrogen, Carlsbad, CA, USA) to generate the pMDC32-EPL1 construct. The pMDC32-EPL1 was transferred into *Agrobacterium tumefaciens* strain GV2260 by electroporation and transformed into *Arabidopsis thaliana* WT (Col-0) plants by the floral dip method [43]. Transformant seeds were selected based on their capacity to grow on 0.2X Murashige and Skoog medium (MS) supplemented with hygromycin at a concentration of 50 mg/mL. Three independent T4 TaEPL1 overexpression lines were obtained and used in this work (35S::TaEPL1-2, -3 and -4).

### 4.3. Measurement of Fresh Weight of 35S::TaEPL1 and WT Lines

*Arabidopsis thaliana* seeds of the WT (Col-0) and 35S::TaEPL1 expression lines (TaEPL1-2, -3 and -4) were grown on 0.2X MS plates for 7 days. Subsequently, the seedlings were transferred to soil pots containing a mixture of Sunshine Mix #3 commercial substrate, perlite, and vermiculite (3:1:1). The pots were placed in a growth chamber with a temperature of  $22 \pm 1$  °C with a photoperiod of 16 h (100  $\mu\text{molm}^{-2} \text{s}^{-1}$ )/8 h, light/dark. The fresh weight (g) of the aerial part of each 40-day-old plant was measured using an analytical balance. Data analysis was done from six biological replicates ( $n = 6$ ).

### 4.4. Pathogen Inocula Preparation

*Pseudomonas syringae* pv. Tomato DC3000 was grown to an OD600 nm of 0.8 in Luria Bertani (LB) medium (pH 7.0) supplemented with 50 mg/mL of rifampicin at 28 °C with shaking. The culture was centrifuged at 13,000 rpm for 10 min. The cells were washed twice with 10 mM MgCl<sub>2</sub> and adjusted at an OD600 nm of 0.2 ( $1 \times 10^8$  CFU/mL) for use in the infection assays. *Botrytis cinerea* strain B05.10 was grown on Potato Dextrose Agar (PDA) in darkness at 28 °C for two weeks. Spores were collected in sterile water, filtered through glass wool to remove hyphae, and quantified in a Neubauer chamber under a Motic model BA-300 microscope with 40 $\times$  magnification (Motic®, San Antonio, TX, USA).

The inoculum was diluted in Potato Dextrose Broth (PDB, 6 g/L) to a concentration of  $1 \times 10^6$  spores/mL<sup>-1</sup>.

#### 4.5. *Arabidopsis* 35S::TaEPL1 Expression Lines—*Pseudomonas syringae* Inoculation Assays

Fourteen-day-old 35S::TaEPL1 and WT plants were transferred to 50 cell tray inserts (4.5 cm cell diameter) containing a sterile mixture of sunshine Mix#3 commercial substrate:vermiculite:perlite (3:1:1), and plants were watered every three days. Four-week-old plants were inoculated with *P. syringae* (50 plants of each line). For each plant, three leaves were infected with bacteria suspension at an OD<sub>600 nm</sub> of 0.2 in 10 mM MgCl<sub>2</sub> using the abaxial injection method [44]. Mock inoculations of 35S::TaEPL1 and WT plants were made by infiltration with 10 mM MgCl<sub>2</sub>. Infiltrated plants were covered with a plastic dome to maintain humidity and incubated in a growth chamber with a photoperiod of 16 h (100 μmolm<sup>-2</sup> s<sup>-1</sup>)/8 h with a light/dark cycle at a temperature of 22 ± 1 °C. For infection symptom evaluation, the leaves were collected 72 h post-inoculation (hpi), photographed, and sterilized using 70% ethanol for 1 min, and then rinsed with distilled water. Five independent 0.5 cm<sup>2</sup> leaf disks from 35S::TaEPL1 and WT plants (*n* = 5) were ground in 10 mM MgCl<sub>2</sub> and serially diluted. Ten microliters of a 1:5 dilution was spotted onto LB agar plates containing 50 mg/mL rifampicin, and colonies were counted after 2 days growth in the dark at 28 °C [44]. All interaction assays with bacteria were done three times with similar results.

#### 4.6. *Arabidopsis* 35S::TaEPL1 Expression Lines—*Botrytis cinerea* Inoculation Assays

Plants of 35S::TaEPL1 and WT were cultivated and prepared for inoculation as described above. Four-week-old 35S::TaEPL1 lines and WT plants were inoculated with *B. cinerea* (50 plants of each line, *n* = 50). For each plant, three leaves were infected on the adaxial surface of leaves avoiding the vascular system with 10 μL drops containing  $1 \times 10^6$  spores. Control PDB (0.25X) was used as a mock treatment. Inoculated plants were covered with a plastic dome to maintain the humidity and placed in darkness for 72 h at a temperature of 22 ± 1 °C. After this period, the disease incidence (percentage of leaves that showed disease symptoms over the total number of inoculated leaves) and leaf lesion area using Image J software IJ 1.46r version (<http://rsb.info.nih.gov/> accessed on 14 June 2023) were determined. Each assay was repeated three times with a similar result.

#### 4.7. Plant Tissue Trypan Blue Staining after *Botrytis cinerea* Infection

Trypan blue dye was used to detect plant cell death after fungal infection. Infected leaves from four-week-old from *A. thaliana* WT and EPL1-3 plants were collected 24 hpi with  $2 \times 10^5$  spores of *B. cinerea* strain B05.10. PDB medium (0.25X) spore diluent was applied as a mock inoculation (control). Leaves were stained by immersing in a staining solution (10 mg of trypan blue, 10 mL of lactic acid, 10 mL of phenol, 10 mL of glycerol, and 10 mL of distilled H<sub>2</sub>O) for 12 h. Leaves were cleared with consecutive washes in ethanol (98%) until chlorophyll was completely removed and photographed.

#### 4.8. Hydrogen Peroxide (H<sub>2</sub>O<sub>2</sub>) Detection by DCFH<sub>2</sub>-DA Labeling

In situ localization of reactive oxygen species (ROS) was performed in root tips of ten-day-old WT and 35S::TaEPL1 lines using the fluorescent molecule 2',7'-dichlorodihydrofluorescein diacetate (DCFH<sub>2</sub>-DA, Sigma-Aldrich, Burlington, MA, USA), as described by [45], with some modifications. The effect of the exogenous application of catalase (CAT, Sigma Aldrich, Burlington, MA, USA) on the H<sub>2</sub>O<sub>2</sub> levels in the WT and 35S::TaEPL1 lines was also analyzed. Seedlings were placed in 24-well culture plates containing 0.2X MS liquid supplemented with 0 or 250 U/mL of CAT (dissolved in pH7 phosphate buffer) and incubated at 22 ± 1 °C in a growth chamber for 4 h with continuous light. The seedlings were then immersed in 25 μmol DCFH<sub>2</sub>-DA in Tris buffer (10 mM Tris, 50 mM KCl, pH 7.2) for 30 min in complete darkness. After rinsing with buffer to remove excess DCFH<sub>2</sub>-DA and CAT, the roots were observed and photographed with a fluorescence microscope

(Zeiss Axio Imager M2; Carl Zeiss Microscopy, Pleasanton, CA, USA) at 10× magnification using excitation and emission wavelengths of 480 nm and 500–550 nm, respectively. Fifteen seedlings of WT and 35S::TaEPL1 lines were analyzed by treatment; each assay was repeated at least three times.

#### 4.9. Determination of Hydrogen Peroxide Content with Potassium Iodide (KI)

Hydrogen peroxide content was determined according to Jungle et al., 2014. Ten-day-old plantlets of WT and 35S::TaEPL1 lines were transferred to plates containing 0.2X MS liquid supplemented with 0 or 250 U/mL of CAT and incubated at  $22 \pm 1$  °C in a growth chamber for 4 h with continuous light. The CAT treatment was carried out to determine the reduction of the H<sub>2</sub>O<sub>2</sub> content between the WT and 35S::TaEPL1 lines. In another experiment, we measured the hydrogen peroxide content in 15-day-old *A. thaliana* WT and 35S::TaEPL1-3 plants following inoculation with pathogens. Plantlets were infected with 2 µL of *P. syringae* or *B. cinerea* inocula for 24 h and 48 h, and as mock treatments, MgCl<sub>2</sub> and PDB (0.25X) were used, respectively. Samples of 100 mg of fresh tissue from whole *Arabidopsis* plantlets were homogenized in an ice bath with 375 µL 0.1% (*w/v*) trichloroacetic acid. The mixture was centrifuged at 4 °C, 7000 rpm for 20 min; 250 µL of supernatant was transferred to a new tube and 250 µL of 10 mM potassium phosphate buffer (pH 7.0) was added, followed by 500 µL of 1 M KI. The absorbance was read at 390 nm using an Epoch-2 microplate reader (Biotek®, Winooski, VT, USA). The H<sub>2</sub>O<sub>2</sub> content was determined using a standard curve (10, 20, 30, 50, 70, and 100 µM H<sub>2</sub>O<sub>2</sub>). Data analysis was done from three biological replicates (*n* = 3).

#### 4.10. RNA Isolation and RT-qPCR Gene Expression Analysis

Total RNA was extracted from 10-day-old seedlings of WT and 35::EPL1-2, -3, and -4 lines using Concert reagent (Invitrogen, Carlsbad, CA, USA), followed by DNAase Turbo digestion (Ambion, Austin, TX, USA) for genomic DNA removal. Total RNA concentration was calculated with an Epoch-2 microplate reader (Biotek®, Winooski, VT, USA). cDNA was synthesized using 1 µg total RNA and a SuperScript™ II Reverse Transcriptase kit (Invitrogen, Carlsbad, CA, USA). RT-qPCR was performed using the Step One Real-Time PCR Detection System (Applied Biosystems, Waltham, MA, USA). The reaction mixture was made in a volume of 10 µL containing 100 ng of cDNA, 200 nM of each primer (Supplementary Table S1), and 5 µL of Maxima SYBR Green/ROX qPCR Master Mix (2×) Thermo Scientific™ (Waltham, MA, USA). First, the relative expression of the ELP1 gene in 35S::TaEPL1 *Arabidopsis* lines was quantified. Then, the EPL1-3 line was selected to measure the expression levels of SA-mediated defense markers, such as *PR1*, *PAL1*, and *WRKY54* and JA-mediated defense markers *LOX3* and *PDF1.2*, *WRKY33*, as well *ZAT1.2* (signaling ROS gene). Expression levels were quantified in rosette leaves of 28-day-old WT and 35S::TaEPL1-3 plants inoculated with *B. cinerea* or *P. syringae* for 24 h. The sequences of oligonucleotides used are included in the Supplementary Table S1. Gene expression was analyzed by the delta Ct method [46]. The *AtUBQ5* gene was used as a reference. For each cDNA sample, three biological replicates (*n* = 3) were analyzed with their respective technical replicates.

#### 4.11. Statistical Analysis

Results from representative experiments are shown as means ± SE. Statistical significance (*p* ≤ 0.05) among data were determined regarding genotypes by One-Way ANOVA or Two-Way ANOVA (genotype × infection). Tukey's post-tests were performed using the GraphPad Prism version 8.0 software (GraphPad Software, San Diego, CA, USA).

## 5. Conclusions

Our data show that *Arabidopsis* lines expressing the EPL1 elicitor of *T. atroviride* are clearly resistant to pathogens with different lifestyles and modes of nutrition, namely, the hemibiotrophic bacteria *P. syringae* and the necrotrophic fungus *B. cinerea*. We observed that

the *Arabidopsis EPL1-3* line, which had the highest expression of the *TaEPL1* gene, resulted in greater protection against pathogens. This reduction in pathogen infection in *EPL1* expressing plants could be correlated with the increase in ROS levels and the induction of SA- and JA-related genes. Therefore, *EPL1* elicitor is an excellent candidate for use as a molecule that provides protection against diverse phytopathogens.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/plants12132443/s1>, Figure S1: Generation of 35S::*TaEPL1* *Arabidopsis* lines. Table S1: Oligonucleotide sequences used to measure gene expression by RT-qPCR analysis.

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## References

1. Harman, G.E.; Howell, C.R.; Viterbo, A.; Chet, I.; Lorito, M. Trichoderma species—Opportunistic, avirulent plant symbionts. *Nat. Rev. Genet.* **2004**, *2*, 43–56. [\[CrossRef\]](#)
2. Khan, R.A.A.; Najeeb, S.; Hussain, S.; Xie, B.; Li, Y. Bioactive Secondary Metabolites from *Trichoderma* spp. against Phytopathogenic Fungi. *Microorganisms* **2020**, *8*, 817. [\[CrossRef\]](#)
3. Guo, Y.; Fan, Z.; Yi, X.; Zhang, Y.; Khan, R.A.A.; Zhou, Z. Sustainable Management of Soil-Borne Bacterium *Ralstonia solanacearum* In Vitro and In Vivo through Fungal Metabolites of Different *Trichoderma* spp. *Sustainability* **2021**, *13*, 1491. [\[CrossRef\]](#)
4. Ruan, J.; Yao, X.; Zhang, K.; Zhao, M. Trichoderma and Its Role in Biological Control of Plant Fungal and Nematode Disease. *Front. Microbiol.* **2023**, *14*, 1275. [\[CrossRef\]](#)
5. Vinale, F.; Sivasithamparam, K.; Ghisalberti, E.; Marra, R.; Barbetti, M.; Li, H.; Woo, S.; Lorito, M. A novel role for Trichoderma secondary metabolites in the interactions with plants. *Physiol. Mol. Plant Pathol.* **2008**, *72*, 80–86. [\[CrossRef\]](#)
6. Mukherjee, P.K.; Horwitz, B.A.; Singh, U.S.; Mukherjee, M.; Schmoll, M. Trichoderma in agriculture, industry and medicine: An overview. In *Trichoderma Biology and Applications*; CAB International: Boston, MA, USA, 2013; Volume 16, pp. 1–9. [\[CrossRef\]](#)
7. Zin, N.A.; Badaluddin, N.A. Biological functions of *Trichoderma* spp. for agriculture applications. *Ann. Agric. Sci.* **2020**, *65*, 168–178. [\[CrossRef\]](#)
8. Hermosa, R.; Viterbo, A.; Chet, I.; Monte, E. Plant-beneficial effects of Trichoderma and of its genes. *Microbiology* **2012**, *158*, 17–25. [\[CrossRef\]](#)
9. Alfiky, A.; Weisskopf, L. Deciphering *Trichoderma*–Plant–Pathogen Interactions for Better Development of Biocontrol Applications. *J. Fungi* **2021**, *7*, 61. [\[CrossRef\]](#)
10. Vinale, F.; Sivasithamparam, K.; Ghisalberti, E.L.; Ruocco, M.; Woo, S.; Lorito, M. *Trichoderma* Secondary Metabolites that Affect Plant Metabolism. *Nat. Prod. Commun.* **2012**, *7*, 1934578X1200701133. [\[CrossRef\]](#)
11. Nieto-Jacobo, M.F.; Steyaert, J.M.; Salazar-Badillo, F.B.; Nguyen, D.V.; Rostás, M.; Braithwaite, M.; De Souza, J.T.; Jimenez-Bremont, J.F.; Ohkura, M.; Stewart, A.; et al. Environmental Growth Conditions of *Trichoderma* spp. Affects Indole Acetic Acid Derivatives, Volatile Organic Compounds, and Plant Growth Promotion. *Front. Plant Sci.* **2017**, *8*, 102. [\[CrossRef\]](#)
12. González-Pérez, E.; Ortega-Amaro, M.A.; Salazar-Badillo, F.B.; Bautista, E.; Douterlungne, D.; Jiménez-Bremont, J.F. The *Arabidopsis*-*Trichoderma* interaction reveals that the fungal growth medium is an important factor in plant growth induction. *Sci. Rep.* **2018**, *8*, 16427. [\[CrossRef\]](#)
13. Thakur, M.; Sohal, B.S. Role of Elicitors in Inducing Resistance in Plants against Pathogen Infection: A Review. *Int. Sch. Res. Not.* **2013**, *2013*, 762412. [\[CrossRef\]](#)
14. Djonović, S.; Pozo, M.J.; Dangott, L.J.; Howell, C.R.; Kenerley, C.M. Sm1, a Proteinaceous Elicitor Secreted by the Biocontrol Fungus *Trichoderma virens* Induces Plant Defense Responses and Systemic Resistance. *Mol. Plant-Microbe Interact.* **2006**, *19*, 838–853. [\[CrossRef\]](#)

15. Lombardi, L.; Faoro, F.; Luti, S.; Baccelli, I.; Martellini, F.; Bernardi, R.; Picciarelli, P.; Scala, A.; Pazzagli, L. Differential timing of defense-related responses induced by cerato-platanin and cerato-populin, two non-catalytic fungal elicitors. *Physiol. Plant.* **2013**, *149*, 408–421. [[CrossRef](#)]
16. Luti, S.; Sella, L.; Quarantin, A.; Pazzagli, L.; Baccelli, I. Twenty years of research on cerato-platanin family proteins: Clues, conclusions, and unsolved issues. *Fungal Biol. Rev.* **2019**, *34*, 13–24. [[CrossRef](#)]
17. Seidl, V.; Marchetti, M.; Schandl, R.; Allmaier, G.; Kubicek, C.P. Epl1, the major secreted protein of *Hypocrea atroviridis* on glucose, is a member of a strongly conserved protein family comprising plant defense response elicitors. *FEBS J.* **2006**, *273*, 4346–4359. [[CrossRef](#)]
18. Buensanteai, N.; Mukherjee, P.K.; Horwitz, B.A.; Cheng, C.; Dangott, L.J.; Kenerley, C.M. Expression and purification of biologically active *Trichoderma virens* proteinaceous elicitor Sm1 in *Pichia pastoris*. *Protein Expr. Purif.* **2010**, *72*, 131–138. [[CrossRef](#)]
19. Djonović, S.; Vargas, W.A.; Kolomiets, M.V.; Horndeski, M.; Wiest, A.; Kenerley, C.M. A Proteinaceous Elicitor Sm1 from the Beneficial Fungus *Trichoderma virens* Is Required for Induced Systemic Resistance in Maize. *Plant Physiol.* **2007**, *145*, 875–889. [[CrossRef](#)]
20. Salas-Marina, M.A.; Isordia-Jasso, M.I.; Islas-Osuna, M.A.; Delgado-Sánchez, P.; Bremont, J.F.J.; Rodríguez-Kessler, M.; Rosales-Saavedra, M.T.; Herrera-Estrella, A.; Ecasas-Flores, S. The Epl1 and Sm1 proteins from *Trichoderma atroviride* and *Trichoderma virens* differentially modulate systemic disease resistance against different life style pathogens in *Solanum lycopersicum*. *Front. Plant Sci.* **2015**, *6*, 77. [[CrossRef](#)]
21. Vargas, W.A.; Djonović, S.; Sukno, S.A.; Kenerley, C.M. Dimerization Controls the Activity of Fungal Elicitors That Trigger Systemic Resistance in Plants. *J. Biol. Chem.* **2008**, *283*, 19804–19815. [[CrossRef](#)]
22. Wang, Y.; Song, J.; Wu, Y.; Odeph, M.; Liu, Z.; Howlett, B.J.; Wang, S.; Yang, P.; Yao, L.; Zhao, L.; et al. Epl4 Proteinaceous Elicitor Produced in *Pichia pastoris* Has a Protective Effect Against *Cercosporidium sofinum* Infections of Soybean Leaves. *Appl. Biochem. Biotechnol.* **2013**, *169*, 722–737. [[CrossRef](#)]
23. Yu, W.; Mijiti, G.; Huang, Y.; Fan, H.; Wang, Y.; Liu, Z. Functional analysis of eliciting plant response protein Epl1-Tas from *Trichoderma asperellum* ACCC30536. *Sci. Rep.* **2018**, *8*, 7974. [[CrossRef](#)]
24. Frías, M.; González, C.; Brito, N. BcSp11, a cerato-platanin family protein, contributes to *Botrytis cinerea* virulence and elicits the hypersensitive response in the host. *New Phytol.* **2011**, *192*, 483–495. [[CrossRef](#)]
25. Frías, M.; Brito, N.; González, C. The *Botrytis cinerea* cerato-platanin BcSp11 is a potent inducer of systemic acquired resistance (SAR) in tobacco and generates a wave of salicylic acid expanding from the site of application. *Mol. Plant Pathol.* **2013**, *14*, 191–196. [[CrossRef](#)]
26. Yang, Y.; Zhang, H.; Li, G.; Li, W.; Wang, X.; Song, F. Ectopic expression of MgSM1, a Cerato-platanin family protein from *Magnaporthe grisea*, confers broad-spectrum disease resistance in *Arabidopsis*. *Plant Biotechnol. J.* **2009**, *7*, 763–777. [[CrossRef](#)]
27. Hong, Y.; Yang, Y.; Zhang, H.; Huang, L.; Li, D.; Song, F. Overexpression of MoSM1, encoding for an immunity-inducing protein from *Magnaporthe oryzae*, in rice confers broad-spectrum resistance against fungal and bacterial diseases. *Sci. Rep.* **2017**, *7*, srep41037. [[CrossRef](#)]
28. Pazzagli, L.; Cappugi, G.; Manao, G.; Camici, G.; Santini, A.; Scala, A. Purification, Characterization, and Amino Acid Sequence of Cerato-platanin, a New Phytotoxic Protein from *Ceratocystis fimbriata* f. sp. *platani*. *J. Biol. Chem.* **1999**, *274*, 24959–24964. [[CrossRef](#)]
29. Wang, Y.; Wu, J.; Kim, S.G.; Tsuda, K.; Gupta, R.; Park, S.-Y.; Kim, S.T.; Kang, K.Y. *Magnaporthe oryzae*-Secreted Protein MSP1 Induces Cell Death and Elicits Defense Responses in Rice. *Mol. Plant-Microbe Interact.* **2016**, *29*, 299–312. [[CrossRef](#)]
30. Torres, M.A. ROS in biotic interactions. *Physiol. Plant.* **2010**, *138*, 414–429. [[CrossRef](#)]
31. Birkenbihl, R.P.; Diezel, C.; Somssich, I.E. *Arabidopsis* WRKY33 is a key transcriptional regulator of hormonal and metabolic responses toward *Botrytis cinerea* infection. *Plant Physiol.* **2012**, *159*, 266–285. [[CrossRef](#)]
32. Salazar-Badillo, F.B.; Sánchez-Rangel, D.; Becerra-Flora, A.; López-Gómez, M.; Nieto-Jacobo, F.; Mendoza-Mendoza, A.; Jiménez-Bremont, J.F. *Arabidopsis thaliana* polyamine content is modified by the interaction with different *Trichoderma* species. *Plant Physiol. Biochem.* **2015**, *95*, 49–56. [[CrossRef](#)]
33. Benítez, T.; Rincón, A.M.; Limón, M.C.; Codón, A.C. Biocontrol mechanisms of *Trichoderma* strains. *Int. Microbiol.* **2004**, *7*, 249–260.
34. Sáenz-Mata, J.; Jiménez-Bremont, J.F. HR4 Gene is induced in the *Arabidopsis*-*Trichoderma atroviride* beneficial interaction. *Int. J. Mol. Sci.* **2012**, *13*, 9110–9128. [[CrossRef](#)]
35. Mukherjee, P.K.; Horwitz, B.A.; Kenerley, C.M. Secondary metabolism in *Trichoderma*—A genomic perspective. *Microbiology* **2012**, *158*, 35–45. [[CrossRef](#)]
36. Shores, M.; Harman, E.G. Differential expression of maize chitinases in the presence or absence of *Trichoderma harzianum* strain T22 and indications of a novel exo- endo-heterodimeric chitinase activity. *BMC Plant Biol.* **2010**, *10*, 136. [[CrossRef](#)]
37. Howell, C.; Puckhaber, L. A study of the characteristics of “P” and “Q” strains of *Trichoderma virens* to account for differences in biological control efficacy against cotton seedling diseases. *Biol. Control.* **2005**, *33*, 217–222. [[CrossRef](#)]
38. Reino, J.L.; Guerrero, R.F.; Hernandez-Galan, R.; Collado, I.G. Secondary metabolites from species of the biocontrol agent *Trichoderma*. *Phytochem. Rev.* **2008**, *7*, 89–123. [[CrossRef](#)]



39. Sáenz-Mata, J.; Salazar-Badillo, F.B.; Jiménez-Bremont, J.F. Transcriptional regulation of *Arabidopsis thaliana* WRKY genes under interaction with beneficial fungus *Trichoderma atroviride*. *Acta Physiol. Plant.* **2014**, *36*, 1085–1093. [[CrossRef](#)]
40. Rizhsky, L.; Davletova, S.; Liang, H.; Mittler, R. The Zinc Finger Protein Zat12 Is Required for Cytosolic Ascorbate Peroxidase 1 Expression during Oxidative Stress in *Arabidopsis*. *J. Biol. Chem.* **2004**, *279*, 11736–11743. [[CrossRef](#)]
41. Scala, A.; Pazzagli, L.; Comparini, C.; Santini, A.; Tegli, S.; Cappugi, G. Cerato-platanin, an early-produced protein by *Ceratocystis fimbriata* f. sp. *platani*, elicits phytoalexin synthesis in host and non-host plants. *J. Plant Pathol.* **2004**, *86*, 27–33.
42. Bennici, A.; Tani, C.; Pazzagli, L. Effects of cerato-platanin, a toxin from *Ceratocystis fimbriata* f. sp. *platani*, on *Platanus acerifolia* leaves: An ultrastructural study. *Caryologia* **2006**, *59*, 291–298. [[CrossRef](#)]
43. Clough, S.J.; Bent, A.F. Floral dip: A simplified method for *Agrobacterium*-mediated transformation of *Arabidopsis thaliana*. *Plant J.* **1998**, *16*, 735–743. [[CrossRef](#)]
44. Katagiri, F.; Thilmony, R.; He, S.Y. The *Arabidopsis Thaliana*-*Pseudomonas Syringae* Interaction. *Arab. Book* **2002**, *1*, e0039. [[CrossRef](#)]
45. Isabel, C.M.A.; Ignacio, J.R.F.; Margarita, R.K.; Gill, S.S.; Alicia, B.F.; Francisco, J.B.J. Down-regulation of arginine decarboxylase gene-expression results in reactive oxygen species accumulation in *Arabidopsis*. *Biochem. Biophys. Res. Commun.* **2018**, *506*, 1071–1077. [[CrossRef](#)]
46. Livak, K.J.; Schmittgen, T.D. Analysis of relative gene expression data using real-time quantitative PCR and the  $2^{-\Delta\Delta CT}$  Method. *Methods* **2001**, *25*, 402–408. [[CrossRef](#)]

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